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(54) Title: SECRETED PROTEINS

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(57) Abstract: Various embodiments of the invention provide human secreted proteins (SECP) and polynucleotides which identify and encode SECP. Embodiments of the invention also provide expression vectors, host cells, antibodies, agonists, and antagonists. Other embodiments provide methods for diagnosing treating, or preventing disorders associated with aberrant expression of SECP.



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SECRETED PROTEINS

TECHNICAL FIELD

The invention relates to novel nucleic acids, secreted proteins encoded by these nucleic acids, and to the use of these nucleic acids and proteins in the diagnosis, treatment, and prevention of cell proliferative, autoimmune/inflammatory, cardiovascular, neurological, and developmental disorders. The invention also relates to the assessment of the effects of exogenous compounds on the expression of nucleic acids and secreted proteins.

BACKGROUND OF THE INVENTION

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Protein transport and secretion are essential for cellular function. Protein transport is mediated by a signal peptide located at the amino terminus of the protein to be transported or secreted. The signal peptide is comprised of about ten to twenty hydrophobic amino acids which target the nascent protein from the ribosome to a particular membrane bound compartment such as the 15 endoplasmic reticulum (ER). Proteins targeted to the ER may either proceed through the secretory pathway or remain in any of the secretory organelles such as the ER, Golgi apparatus, or lysosomes. Proteins that transit through the secretory pathway are either secreted into the extracellular space or retained in the plasma membrane. Proteins that are retained in the plasma membrane contain one or more transmembrane domains, each comprised of about 20 hydrophobic amino acid residues. Secreted proteins are generally synthesized as inactive precursors that are activated by posttranslational processing events during transit through the secretory pathway. Such events include glycosylation, proteolysis, and removal of the signal peptide by a signal peptidase. Other events that may occur during protein transport include chaperone-dependent unfolding and folding of the nascent protein and interaction of the protein with a receptor or pore complex. Examples of secreted proteins with amino terminal signal peptides are discussed below and include proteins with important roles in cell-to-cell signaling. Such proteins include transmembrane receptors and cell surface markers, extracellular matrix molecules, cytokines, hormones, growth and differentiation factors, enzymes, neuropeptides, vasomediators, cell surface markers, and antigen recognition molecules. (Reviewed in Alberts, B. et al. (1994) Molecular Biology of The Cell, Garland Publishing, New York, NY, pp. 557-560, 582-592.)

Cell surface markers include cell surface antigens identified on leukocytic cells of the immune system. These antigens have been identified using systematic, monoclonal antibody (mAb)-based "shot gun" techniques. These techniques have resulted in the production of hundreds of mAbs

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directed against unknown cell surface leukocytic antigens. These antigens have been grouped into "clusters of differentiation" based on common immunocytochemical localization patterns in various differentiated and undifferentiated leukocytic cell types. Antigens in a given cluster are presumed to identify a single cell surface protein and are assigned a "cluster of differentiation" or "CD"

5 designation. Some of the genes encoding proteins identified by CD antigens have been cloned and verified by standard molecular biology techniques. CD antigens have been characterized as both transmembrane proteins and cell surface proteins anchored to the plasma membrane via covalent attachment to fatty acid-containing glycolipids such as glycosylphosphatidylinositol (GPI). (Reviewed in Barclay, A.N. et al. (1995) The Leucocyte Antigen Facts Book, Academic Press, San Diego, CA, pp. 17-20.)

Matrix proteins (MPs) are transmembrane and extracellular proteins which function in formation, growth, remodeling, and maintenance of tissues and as important mediators and regulators of the inflammatory response. The expression and balance of MPs may be perturbed by biochemical changes that result from congenital, epigenetic, or infectious diseases. In addition, MPs affect leukocyte migration, proliferation, differentiation, and activation in the immune response. MPs are frequently characterized by the presence of one or more domains which may include collagen-like domains, EGF-like domains, immunoglobulin-like domains, and fibronectin-like domains. In addition, MPs may be heavily glycosylated and may contain an Arginine-Glycine-Aspartate (RGD) tripeptide motif which may play a role in adhesive interactions. MPs include extracellular proteins such as fibronectin, collagen, galectin, vitronectin and its proteolytic derivative somatomedin B; and cell adhesion receptors such as cell adhesion molecules (CAMs), cadherins, and integrins. (Reviewed in Ayad, S. et al. (1994) The Extracellular Matrix Facts Book, Academic Press, San Diego, CA, pp. 2-16; Ruoslahti, E. (1997) Kidney Int. 51:1413-1417; Sjaastad, M.D. and W.J. Nelson (1997) BioEssays 19:47-55.)

Mucins are highly glycosylated glycoproteins that are the major structural component of the mucus gel. The physiological functions of mucins are cytoprotection, mechanical protection, maintenance of viscosity in secretions, and cellular recognition. MUC6 is a human gastric mucin that is also found in gall bladder, pancreas, seminal vesicles, and female reproductive tract (Toribara, N.W. et al. (1997) J. Biol. Chem. 272:16398-16403). The MUC6 gene has been mapped to human chromosome 11 (Toribara, N.W. et al. (1993) J. Biol. Chem. 268:5879-5885). Hemoniucin is a novel *Drosophila* surface mucin that may be involved in the induction of antibacterial effector molecules (Theopold, U. et al. (1996) J. Biol. Chem. 217:12708-12715).

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Tuftelins are one of four different enamel matrix proteins that have been identified so far.

The other three known enamel matrix proteins are the amelogenins, enamelin and ameloblastin. Assembly of the enamel extracellular matrix from these component proteins is believed to be critical in producing a matrix competent to undergo mineral replacement (Paine, C.T. et al. (1998) Connect Tissue Res. 38:257-267). Tuftelin mRNA has been found to be expressed in human ameloblastoma tumor, a non-mineralized odontogenic tumor (Deutsch, D. et al. (1998) Connect. Tissue Res. 39:177-184).

Olfactomedin-related proteins are extracellular matrix, secreted glycoproteins with conserved C-terminal motifs. They are expressed in a wide variety of tissues and in a broad range of species, from Caenorhabditis elegans to Homo sapiens. Olfactomedin-related proteins comprise a gene family with at least 5 family members in humans. One of the five, TIGR/myocilin protein, is expressed in the eye and is associated with the pathogenesis of glaucoma (Kulkarni, N.H. et al. (2000) Genet. Res. 76:41-50). Research by Yokoyama, M. et al. (1996; DNA Res. 3:311-320) found a 135-amino acid protein, termed AMY, having 96% sequence identity with rat neuronal olfactomedin-releated ER localized protein in a neuroblastoma cell line cDNA library, suggesting an essential role for AMY in nerve tissue. Neuron-specific olfactomedin-related glycoproteins isolated from rat brain cDNA libraries show strong sequence similarity with olfactomedin. This similarity is suggestive of a matrix-related function of these glycoproteins in neurons and neurosecretory cells (Danielson, P.E. et al. (1994) J. Neurosci. Res. 38:468-478).

Mac-2 binding protein is a 90-kD serum protein (90K), a secreted glycoprotein isolated from both the human breast carcinoma cell line SK-BR-3, and human breast milk. It specifically binds to a human macrophage-associated lectin, Mac-2. Structurally, the mature protein is 567 amino acids in length and is proceeded by an 18-amino acid leader. There are 16 cysteines and seven potential N-linked glycosylation sites. The first 106 amino acids represent a domain very similar to an ancient protein superfamily defined by a macrophage scavenger receptor cysteine-rich domain (Koths, K. et al. (1993) J. Biol. Chem. 268:14245-14249). 90K is elevated in the serum of subpopulations of AIDS patients and is expressed at varying levels in primary tumor samples and tumor cell lines. Ullrich, A. et al. (1994; J. Biol. Chem. 269:18401-18407) have demonstrated that 90K stimulates host defense systems and can induce interleukin-2 secretion. This immune stimulation is proposed to be a result of oncogenic transformation, viral infection or pathogenic invasion (Ullrich et al., supra).

Semaphorins are a large group of axonal guidance molecules consisting of at least 30 different members and are found in vertebrates, invertebrates, and even certain viruses. All semaphorins contain the sema domain which is approximately 500 amino acids in length. Neuropilin, a semaphorin receptor, has been shown to promote neurite outgrowth *in vitro*. The extracellular region of

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neuropilins consists of three different domains: CUB, discoidin, and MAM domains. The CUB and the MAM motifs of neuropilin have been suggested to have roles in protein-protein interactions and are thought to be involved in the binding of semaphorins through the sema and the C-terminal domains (reviewed in Raper, J.A. (2000) Curr. Opin. Neurobiol. 10:88-94). Plexins are neuronal cell surface molecules that mediate cell adhesion via a homophilic binding mechanism in the presence of calcium ions. Plexins have been shown to be expressed in the receptors and neurons of particular sensory systems (Ohta, K. et al. (1995) Cell 14:1189-1199). There is evidence that suggests that some plexins function to control motor and CNS axon guidance in the developing nervous system. Plexins, which themselves contain complete semaphorin domains, may be both the ancestors of classical semaphorins and binding partners for semaphorins (Winberg, M.L. et al (1998) Cell 95:903-916).

Human pregnancy-specific beta 1-glycoprotein (PSG) is a family of closely related glycoproteins of molecular weights of 72 KDa, 64KDa, 62KDa, and 54KDa. Together with the carcinoembryonic antigen, they comprise a subfamily within the immunoglobulin superfamily (Plouzek, C.A. and J.Y. Chou, (1991) Endocrinology 129:950-958) Different subpopulations of PSG have been found to be produced by the trophoblasts of the human placenta, and the amnionic and chorionic membranes (Plouzek, C.A. et al. (1993) Placenta 14:277-285).

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Torsion dystonia is an autosomal dominant movement disorder consisting of involuntary muscular contractions. The disorder has been linked to a 3-base pair mutation in the DYT-1 gene, which encodes torsin A (Ozelius, L.J. et al. (1997) Nat. Genet. 17:40-48). Torsin A bears significant homology to the Hsp100/Clp family of ATPase chaperones, which are conserved in humans, rats, mice, and C. elegans. Strong expression of DYT-1 in neuronal processes indicates a potential role for torsins in synaptic communication (Kustedjo, K. et al. (2000) J. Biol. Chem. 275:27933-27939 and Konakova M. et al. (2001) Arch. Neurol. 58:921-927).

Autocrine motility factor (AMF) is one of the motility cytokines regulating tumor cell migration; therefore identification of the signaling pathway coupled with it has critical importance. Autocrine motility factor receptor (AMFR) expression has been found to be associated with tumor progression in thymoma (Ohta Y. et al. (2000) Int. J. Oncol. 17:259-264). AMFR is a cell surface glycoprotein of molecular weight 78KDa.

Hormones are secreted molecules that travel through the circulation and bind to specific receptors on the surface of, or within, target cells. Although they have diverse biochemical compositions and mechanisms of action, hormones can be grouped into two categories. One category includes small lipophilic hormones that diffuse through the plasma membrane of target cells, bind to cytosolic or nuclear receptors, and form a complex that alters gene expression. Examples of these

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molecules include retinoic acid, thyroxine, and the cholesterol-derived steroid hormones such as progesterone, estrogen, testosterone, cortisol, and aldosterone. The second category includes hydrophilic hormones that function by binding to cell surface receptors that transduce signals across the plasma membrane. Examples of such hormones include amino acid derivatives such as catecholamines (epinephrine, norepinephrine) and histamine, and peptide hormones such as glucagon, insulin, gastrin, secretin, cholecystokinin, adrenocorticotropic hormone, follicle stimulating hormone, luteinizing hormone, thyroid stimulating hormone, and vasopressin. (See, for example, Lodish et al. (1995) Molecular Cell Biology, Scientific American Books Inc., New York, NY, pp. 856-864.)

Pro-opiomelanocortin (POMC) is the precursor polypeptide of corticotropin (ACTH), a hormone synthesized by the anterior pituitary gland, which functions in the stimulation of the adrenal cortex. POMC is also the precursor polypeptide of the hormone beta-lipotropin (beta-LPH). Each hormone includes smaller peptides with distinct biological activities: alpha-melanotropin (alpha-MSH) and corticotropin-like intermediate lobe peptide (CLIP) are formed from ACTH; gamma-lipotropin (gamma-LPH) and beta-endorphin are peptide components of beta-LPH; while beta-MSH is contained within gamma-LPH. Adrenal insufficiency due to ACTH deficiency, resulting from a genetic mutation in exons 2 and 3 of POMC results in an endocrine disorder characterized by earlyonset obesity, adrenal insufficiency, and red hair pigmentation (Chretien, M. et al. (1979) Can. J. Biochem. 57:1111-1121; Krude, H. et al. (1998) Nat. Genet. 19:155-157; Online Mendelian Inheritance in Man (OMIM) 176830).

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Growth and differentiation factors are secreted proteins which function in intercellular communication. Some factors require oligomerization or association with membrane proteins for activity. Complex interactions among these factors and their receptors trigger intracellular signal transduction pathways that stimulate or inhibit cell division, cell differentiation, cell signaling, and cell motility. Most growth and differentiation factors act on cells in their local environment (paracrine 25 signaling). There are three broad classes of growth and differentiation factors. The first class includes the large polypeptide growth factors such as epidermal growth factor, fibroblast growth factor, transforming growth factor, insulin-like growth factor, and platelet-derived growth factor. The second class includes the hematopoietic growth factors such as the colony stimulating factors (CSFs). Hematopoietic growth factors stimulate the proliferation and differentiation of blood cells such as Blymphocytes, T-lymphocytes, erythrocytes, platelets, eosinophils, basophils, neutrophils, macrophages, and their stem cell precursors. The third class includes small peptide factors such as bombesin, vasopressin, oxytocin, endothelin, transferrin, angiotensin II, vasoactive intestinal peptide, and bradykinin, which function as hormones to regulate cellular functions other than proliferation.

Growth and differentiation factors play critical roles in neoplastic transformation of cells in vitro and in tumor progression in vivo. Inappropriate expression of growth factors by tumor cells may contribute to vascularization and metastasis of tumors. During hematopoiesis, growth factor misregulation can result in anemias, leukemias, and lymphomas. Certain growth factors such as interferon are cytotoxic to tumor cells both in vivo and in vitro. Moreover, some growth factors and growth factor receptors are related both structurally and functionally to oncoproteins. In addition, growth factors affect transcriptional regulation of both proto-oncogenes and oncosuppressor genes.

(Reviewed in Pimentel, E. (1994) Handbook of Growth Factors, CRC Press, Ann Arbor, MI, pp. 1-9.)

The Slit protein, first identified in Drosophila, is critical in central nervous system midline formation and potentially in nervous tissue histogenesis and axonal pathfinding. Itoh et al. (1998; Brain Res. Mol. Brain Res. 62:175-186) have identified mammalian homologues of the slit gene (human Slit-1, Slit-2, Slit-3 and rat Slit-1). The encoded proteins are putative secreted proteins containing EGF-like motifs and leucine-rich repeats, both of which are conserved protein-protein interaction domains. Slit-1, -2, and -3 mRNAs are expressed in the brain, spinal cord, and thyroid, respectively (Itoh et al., supra). The Slit family of proteins are indicated to be functional ligands of glypican-1 in nervous tissue and it is suggested that their interactions may be critical in certain stages during central nervous system histogenesis (Liang, Y. et al. (1999) J. Biol. Chem. 274:17885-17892).

Neuropeptides and vasomediators (NP/VM) comprise a large family of endogenous signaling molecules. Included in this family are neuropeptides and neuropeptide hormones such as bombesin, neuropeptide Y, neurotensin, neuromedin N, melanocortins, opioids, galanin, somatostatin, tachykinins, urotensin II and related peptides involved in smooth muscle stimulation, vasopressin, vasoactive intestinal peptide, and circulatory system-borne signaling molecules such as angiotensin, complement, calcitonin, endothelins, formyl-methionyl peptides, glucagon, cholecystokinin and gastrin. NP/VMs can transduce signals directly, modulate the activity or release of other neurotransmitters and hormones, and act as catalytic enzymes in cascades. The effects of NP/VMs range from extremely brief to long-lasting. (Reviewed in Martin, C.R. et al. (1985) Endocrine Physiology, Oxford University Press, New York, NY, pp. 57-62.)

NP/VMs are involved in numerous neurological and cardiovascular disorders. For example, neuropeptide Y is involved in hypertension, congestive heart failure, affective disorders, and appetite regulation. Somatostatin inhibits secretion of growth hormone and prolactin in the anterior pituitary, as well as inhibiting secretion in intestine, pancreatic acinar cells, and pancreatic beta-cells. A reduction in somatostatin levels has been reported in Alzheimer's disease and Parkinson's disease. Vasopressin acts in the kidney to increase water and sodium absorption, and in higher concentrations stimulates

contraction of vascular smooth muscle, platelet activation, and glycogen breakdown in the liver. Vasopressin and its analogues are used clinically to treat diabetes insipidus. Endothelin and angiotensin are involved in hypertension, and drugs, such as captopril, which reduce plasma levels of angiotensin, are used to reduce blood pressure (Watson, S. and S. Arkinstall (1994) The G-protein Linked Receptor Facts Book, Academic Press, San Diego CA, pp. 194; 252; 284; 55; 111).

Neuropeptides have also been shown to have roles in nociception (pain). Vasoactive intestinal peptide appears to play an important role in chronic neuropathic pain. Nociceptin, an endogenous ligand for for the opioid receptor-like 1 receptor, is thought to have a predominantly anti-nociceptive effect, and has been shown to have analgesic properties in different animal models of tonic or chronic pain (Dickinson, T. and S.M. Fleetwood-Walker (1998) Trends Pharmacol. Sci. 19:346-348).

Other proteins that contain signal peptides include secreted proteins with enzymatic activity. Such activity includes, for example, oxidoreductase/dehydrogenase activity, transferase activity, hydrolase activity, lyase activity, isomerase activity, or ligase activity. For example, matrix metalloproteinases are secreted hydrolytic enzymes that degrade the extracellular matrix and thus play an important role in tumor metastasis, tissue morphogenesis, and arthritis (Reponen, P. et al. (1995) Dev. Dyn. 202:388-396; Firestein, G.S. (1992) Curr. Opin. Rheumatol. 4:348-354; Ray, J.M. and W.G. Stetler-Stevenson (1994) Eur. Respir. J. 7:2062-2072; and Mignatti, P. and D.B. Rifkin (1993) Physiol. Rev. 73:161-195). Additional examples are the acetyl-CoA synthetases which activate acetate for use in lipid synthesis or energy generation (Luong, A. et al. (2000) J. Biol. Chem. 275:26458-26466). The result of acetyl-CoA synthetase activity is the formation of acetyl-CoA from acetate and CoA. Acetyl-CoA synthetase share a region of sequence similarity identified as the AMP-binding domain signature. Acetyl-CoA synthetase has been shown to be associated with hypertension (Toh, H. (1991) Protein Seq. Data Anal. 4:111-117; and Iwai, N. et al. (1994) Hypertension 23:375-380).

A number of isomerases catalyze steps in protein folding, phototransduction, and various anabolic and catabolic pathways. One class of isomerases is known as peptidyl-prolyl *cis-trans* isomerases (PPlases). PPlases catalyze the *cis* to *trans* isomerization of certain proline imidic bonds in proteins. Two families of PPlases are the FK506 binding proteins (FKBPs), and cyclophilins (CyPs). FKBPs bind the potent immunosuppressants FK506 and rapamycin, thereby inhibiting signaling pathways in T-cells. Specifically, the PPlase activity of FKBPs is inhibited by binding of FK506 or rapamycin. There are five members of the FKBP family which are named according to their calculated molecular masses (FKBP12, FKBP13, FKBP25, FKBP52, and FKBP65), and localized to different regions of the cell where they associate with different protein complexes (Coss, M. et al. (1995) J. Biol. Chem. 270:29336-29341; Schreiber, S.L. (1991) Science 251:283-287).

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The peptidyl-prolyl isomerase activity of CyP may be part of the signaling pathway that leads to T-cell activation. CyP isomerase activity is associated with protein folding and protein trafficking, and may also be involved in assembly/disassembly of protein complexes and regulation of protein activity. For example, in *Drosophila*, the CyP NinaA is required for correct localization of rhodopsins, while a mammalian CyP (Cyp40) is part of the Hsp90/Hsc70 complex that binds steroid receptors. The mammalian CypA has been shown to bind the gag protein from human immunodeficiency virus 1 (HIV-1), an interaction that can be inhibited by cyclosporin. Since cyclosporin has potent anti-HIV-1 activity, CypA may play an essential function in HIV-1 replication. Finally, Cyp40 has been shown to bind and inactivate the transcription factor c-Myb, an effect that is reversed by cyclosporin. This effect implicates CyPs in the regulation of transcription, transformation, and differentiation (Bergsma, D.J. et al (1991) J. Biol. Chem. 266:23204-23214; Hunter, T. (1998) Cell 92:141-143; and Leverson, J.D. and S.A. Ness, (1998) Mol. Cell. 1:203-211).

Gamma-carboxyglutamic acid (Gla) proteins rich in proline (PRGPs) are members of a family of vitamin K-dependent single-pass integral membrane proteins. These proteins are characterized by an extracellular amino terminal domain of approximately 45 amino acids rich in Gla. The intracellular carboxyl terminal region contains one or two copies of the sequence PPXY, a motif present in a variety of proteins involved in such diverse cellular functions as signal transduction, cell cycle progression, and protein turnover (Kulman, J.D. et al. (2001) Proc. Natl. Acad. Sci. USA 98:1370-1375). The process of post-translational modification of glutamic residues to form Gla is Vitamin K-dependent carboxylation. Proteins which contain Gla include plasma proteins involved in blood coagulation. These proteins are prothrombin, proteins C, S, and Z, and coagulation factors VII, IX, and X. Osteocalcin (bone-Gla protein, BGP) and matrix Gla-protein (MGP) also contain Gla (Friedman, P.A. and C.T. Przysiecki (1987) Int. J. Biochem. 19:1-7; Vermeer, C. (1990) Biochem. J. 266:625-636).

25 Immunoglobulins

Antigen recognition molecules are key players in the sophisticated and complex immune systems which all vertebrates have developed to provide protection from viral, bacterial, fungal, and parasitic infections. A key feature of the immune system is its ability to distinguish foreign molecules, or antigens, from "self" molecules. This ability is mediated primarily by secreted and transmembrane proteins expressed by leukocytes (white blood cells) such as lymphocytes, granulocytes, and monocytes. Most of these proteins belong to the immunoglobulin (Ig) superfamily, members of which contain one or more repeats of a conserved structural domain. This Ig domain is comprised of antiparallel β sheets joined by a disulfide bond in an arrangement called the Ig fold. The criteria for a

protein to be a member of the Ig superfamily is to have one or more Ig domains, which are regions of 70-110 amino acid residues in length homologous to either Ig variable-like (V) or Ig constant-like (C) domains. Members of the Ig superfamily include antibodies (Ab), T cell receptors (TCRs), class I and II major histocompatibility (MHC) proteins and immune cell-specific surface markers such as the "cluster of differentiation" or CD antigens, CD2, CD3, CD4, CD8, poly-Ig receptors, Fc receptors, neural cell-adhesion molecule (NCAM) and platelet-derived growth factor receptor (PDGFR).

lg domains (V and C) are regions of conserved amino acid residues that give a polypeptide a globular tertiary structure called an immunoglobulin (or antibody) fold, which consists of two approximately parallel layers of β -sheets. Conserved cysteine residues form an intrachain disulfide-bonded loop, 55-75 amino acid residues in length, which connects the two layers of β -sheets. Each β -sheet has three or four anti-parallel β -strands of 5-10 amino acid residues. Hydrophobic and hydrophilic interactions of amino acid residues within the β -strands stabilize the lg fold (hydrophobic on inward facing amino acid residues and hydrophilic on the amino acid residues in the outward facing portion of the strands). A V domain consists of a longer polypeptide than a C domain, with an additional pair of β -strands in the lg fold.

A consistent feature of Ig superfamily genes is that each sequence of an Ig domain is encoded by a single exon. It is possible that the superfamily evolved from a gene coding for a single Ig domain involved in mediating cell-cell interactions. New members of the superfamily then arose by exon and gene duplications. Modern Ig superfamily proteins contain different numbers of V and/or C domains. Another evolutionary feature of this superfamily is the ability to undergo DNA rearrangements, a unique feature retained by the antigen receptor members of the family.

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Many members of the Ig superfamily are integral plasma membrane proteins with extracellular Ig domains. The hydrophobic amino acid residues of their transmembrane domains and their cytoplasmic tails are very diverse, with little or no homology among Ig family members or to known signal-transducing structures. There are exceptions to this general superfamily description. For example, the cytoplasmic tail of PDGFR has tyrosine kinase activity. In addition Thy-1 is a glycoprotein found on thymocytes and T cells. This protein has no cytoplasmic tail, but is instead attached to the plasma membrane by a covalent glycophosphatidylinositol linkage.

Another common feature of many lg superfamily proteins is the interactions between lg domains which are essential for the function of these molecules. Interactions between lg domains of a multimeric protein can be either homophilic or heterophilic (i.e., between the same or different lg domains). Antibodies are multimeric proteins which have both homophilic and heterophilic interactions between lg domains. Pairing of constant regions of heavy chains forms the Fc region of an antibody

and pairing of variable regions of light and heavy chains form the antigen binding site of an antibody. Heterophilic interactions also occur between lg domains of different molecules. These interactions provide adhesion between cells for significant cell-cell interactions in the immune system and in the developing and mature nervous system. (Reviewed in Abbas, A.K. et al. (1991) Cellular and

Molecular Immunology, W.B. Saunders Company, Philadelphia, PA, pp. 142-145.)

Antibodies

MHC proteins are cell surface markers that bind to and present foreign antigens to T cells.

MHC molecules are classified as either class I or class II. Class I MHC molecules (MHC I) are expressed on the surface of almost all cells and are involved in the presentation of antigen to cytotoxic T cells. For example, a cell infected with virus will degrade intracellular viral proteins and express the protein fragments bound to MHC I molecules on the cell surface. The MHC I/antigen complex is recognized by cytotoxic T-cells which destroy the infected cell and the virus within. Class II MHC molecules are expressed primarily on specialized antigen-presenting cells of the immune system, such as B-cells and macrophages. These cells ingest foreign proteins from the extracellular fluid and express MHC II/antigen complex on the cell surface. This complex activates helper T-cells, which then secrete cytokines and other factors that stimulate the immune response. MHC molecules also play an important role in organ rejection following transplantation. Rejection occurs when the recipient's T-cells respond to foreign MHC molecules on the transplanted organ in the same way as to self MHC molecules bound to foreign antigen. (Reviewed in Alberts et al., supra, pp. 1229-1246.)

Antibodies are multimeric members of the Ig superfamily which are either expressed on the surface of B-cells or secreted by B-cells into the circulation. Antibodies bind and neutralize foreign antigens in the blood and other extracellular fluids. The prototypical antibody is a tetramer consisting of two identical heavy polypeptide chains (H-chains) and two identical light polypeptide chains (L-chains) interlinked by disulfide bonds. This arrangement confers the characteristic Y-shape to antibody molecules. Antibodies are classified based on their H-chain composition. The five antibody classes, IgA, IgD, IgE, IgG and IgM, are defined by the α , δ , ϵ , γ , and μ H-chain types. There are two types of L-chains, κ and λ , either of which may associate as a pair with any H-chain pair. IgG, the most common class of antibody found in the circulation, is tetrameric, while the other classes of antibodies are generally variants or multimers of this basic structure.

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H-chains and L-chains each contain an N-terminal variable region and a C-terminal constant region. The constant region consists of about 110 amino acids in L-chains and about 330 or 440 amino acids in H-chains. The amino acid sequence of the constant region is nearly identical among H- or L-chains of a particular class. The variable region consists of about 110 amino acids in both H- and L-

chains. However, the amino acid sequence of the variable region differs among H- or L-chains of a particular class. Within each H- or L-chain variable region are three hypervariable regions of extensive sequence diversity, each consisting of about 5 to 10 amino acids. In the antibody molecule, the H- and L-chain hypervariable regions come together to form the antigen recognition site.

5 (Reviewed in Alberts et al. *supra*, pp. 1206-1213; 1216-1217.)

Both H-chains and L-chains contain the repeated Ig domains of members of the Ig superfamily. For example, a typical H-chain contains four Ig domains, three of which occur within the constant region and one of which occurs within the variable region and contributes to the formation of the antigen recognition site. Likewise, a typical L-chain contains two Ig domains, one of which occurs within the constant region and one of which occurs within the variable region.

The immune system is capable of recognizing and responding to any foreign molecule that enters the body. Therefore, the immune system must be armed with a full repertoire of antibodies against all potential antigens. Such antibody diversity is generated by somatic rearrangement of gene segments encoding variable and constant regions. These gene segments are joined together by site-specific recombination which occurs between highly conserved DNA sequences that flank each gene segment. Because there are hundreds of different gene segments, millions of unique genes can be generated combinatorially. In addition, imprecise joining of these segments and an unusually high rate of somatic mutation within these segments further contribute to the generation of a diverse antibody population.

20 Expression profiling

Microarrays are analytical tools used in bioanalysis. A microarray has a plurality of molecules spatially distributed over, and stably associated with, the surface of a solid support. Microarrays of polypeptides, polynucleotides, and/or antibodies have been developed and find use in a variety of applications, such as gene sequencing, monitoring gene expression, gene mapping, bacterial identification, drug discovery, and combinatorial chemistry.

One area in particular in which microarrays find use is in gene expression analysis. Array technology can provide a simple way to explore the expression of a single polymorphic gene or the expression profile of a large number of related or unrelated genes. When the expression of a single gene is examined, arrays are employed to detect the expression of a specific gene or its variants. When an expression profile is examined, arrays provide a platform for identifying genes that are tissue specific, are affected by a substance being tested in a toxicology assay, are part of a signaling cascade, carry out housekeeping functions, or are specifically related to a particular genetic predisposition, condition, disease, or disorder.

Breast Cancer

There are more than 180,000 new cases of breast cancer diagnosed each year, and the mortality rate for breast cancer approaches 10% of all deaths in females between the ages of 45-54 (K. Gish (1999) AWIS Magazine 28:7-10). However the survival rate based on early diagnosis of localized breast cancer is extremely high (97%), compared with the advanced stage of the disease in which the tumor has spread beyond the breast (22%). Current procedures for clinical breast examination are lacking in sensitivity and specificity, and efforts are underway to develop comprehensive gene expression profiles for breast cancer that may be used in conjunction with conventional screening methods to improve diagnosis and prognosis of this disease (Perou CM et al. (2000) Nature 406:747-752).

Breast cancer is a genetic disease commonly caused by mutations in cellular disease.

Mutations in two genes, BRCA1 and BRCA2, are known to greatly predispose a woman to breast cancer and may be passed on from parents to children (Gish, <u>supra</u>). However, this type of hereditary breast cancer accounts for only about 5% to 9% of breast cancers, while the vast majority of breast cancer is due to noninherited mutations that occur in breast epithelial cells.

A good deal is already known about the expression of specific genes associated with breast cancer. For example, the relationship between expression of epidermal growth factor (EGF) and its receptor, EGFR, to human mammary carcinoma has been particularly well studied. (See Khazaie et al., supra, and references cited therein for a review of this area.) Overexpression of EGFR, particularly coupled with down-regulation of the estrogen receptor, is a marker of poor prognosis in breast cancer patients. In addition, EGFR expression in breast tumor metastases is frequently elevated relative to the primary tumor, suggesting that EGFR is involved in tumor progression and metastasis. This is supported by accumulating evidence that EGF has effects on cell functions related to metastatic potential, such as cell motility, chemotaxis, secretion and differentiation. Changes in 25 expression of other members of the erbB receptor family, of which EGFR is one, have also been implicated in breast cancer. The abundance of erbB receptors, such as HER-2/neu, HER-3, and HER-4, and their ligands in breast cancer points to their functional importance in the pathogenesis of the disease, and may therefore provide targets for therapy of the disease (Bacus, SS et al. (1994) Am J Clin Pathol 102:S13-S24). Other known markers of breast cancer include a human secreted frizzled protein mRNA that is downregulated in breast tumors; the matrix G1a protein which is overexpressed is human breast carcinoma cells; Drg1 or RTP, a gene whose expression is diminished in colon, breast, and prostate tumors; maspin, a tumor suppressor gene downregulated in invasive breast carcinomas; and CaN19, a member of the S100 protein family, all of which are down regulated in mammary

carcinoma cells relative to normal mammary epithelial cells (Zhou Z et al. (1998) Int J Cancer 78:95-99; Chen, L et al. (1990) Oncogene 5:1391-1395; Ulrix W et al (1999) FEBS Lett 455:23-26; Sager, R et al. (1996) Curr Top Microbiol Immunol 213:51-64; and Lee, SW et al. (1992) Proc Natl Acad Sci USA 89:2504-2508).

Cell lines derived from human mammary epithelial cells at various stages of breast cancer provide a useful model to study the process of malignant transformation and tumor progression as it has been shown that these cell lines retain many of the properties of their parental tumors for lengthy culture periods (Wistuba II et al. (1998) Clin Cancer Res 4:2931-2938). Such a model is particularly useful for comparing phenotypic and molecular characteristics of human mammary epithelial cells at various stages of malignant transformation.

Colon Cancer

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Colorectal cancer is the second leading cause of cancer deaths in the United States. Colon cancer is associated with aging, since 90% of the total cases occur in individuals over the age of 55.

A widely accepted hypothesis is that several contributing genetic mutations must accumulate over time in an individual who develops the disease. To understand the nature of genetic alterations in colorectal cancer, a number of studies have focused on the inherited syndromes. The first known inherited syndrome, Familial Adenomatous Polyposis (FAP), is caused by mutations in the Adenomatous Polyposis Coli gene (APC), resulting in truncated or inactive forms of the protein. This tumor suppressor gene has been mapped to chromosome 5q. The second known inherited syndrome is hereditary nonpolyposis colorectal cancer (HNPCC), which is caused by mutations in mismatch repair genes.

Although hereditary colon cancer syndromes occur in a small percentage of the population and most colorectal cancers are considered sporadic, knowledge from studies of the hereditary syndromes can be generally applied. For instance, somatic mutations in APC occur in at least 80% of indiscriminate colon tumors. APC mutations are thought to be the initiating event in the disease. Other mutations occur subsequently. Approximately 50% of colorectal cancers contain activating mutations in ras, while 85% contain inactivating mutations in p53. Changes in these genes lead to gene expression changes in colon cancer. Less is understood about downstream targets of these mutations and the role they may play in cancer development and progression.

30 Tangier Disease

Tangier disease (TD) is a genetic disorder characterized by near absence of circulating high density lipoprotein (HDL) and the accumulation of cholesterol esters in many tissues, including tonsils, lymph nodes, liver, spleen, thymus, and intestine. Low levels of HDL represent a clear predictor of

premature coronary artery disease and homozygous TD correlates with a four- to six-fold increase in cardiovascular disease compared to controls. HDL plays a cardio-protective role in reverse cholesterol transport, the flux of cholesterol from peripheral cells such as tissue macrophages through plasma lipoproteins to the liver. The HDL protein, apolipoprotein A-I, plays a major role in this process, interacting with the cell surface to remove excess cholesterol and phospholipids. This pathway is severely impaired in TD and the defect lies in a specific gene, the ABC1 transporter. This gene is a member of the family of ATP-binding cassette transporters, which utilize ATP hydrolysis to transport a variety of substrates across membranes.

Cell lines

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Cell lines derived from human mammary epithelial cells at various stages of breast cancer provide a useful model to study the process of malignant transformation and tumor progression as it has been shown that these cell lines retain many of the properties of their parental tumors for lengthy culture periods (Wistuba, I.I. et al. (1998) Clin. Cancer Res. 4:2931-2938). Such a model is particularly useful for comparing phenotypic and molecular characteristics of human mammary epithelial cells at various stages of malignant transformation.

The human C3A cell line is a clonal derivative of HepG2/C3 (hepatoma cell line, isolated from a 15-year-old male with liver tumor), which was selected for strong contact inhibition of growth. The use of a clonal population enhances the reproducibility of the cells. C3A cells have many characteristics of primary human hepatocytes in culture: i) expression of insulin receptor and insulin-like growth factor II receptor; ii) secretion of a high ratio of serum albumin compared with α-fetoprotein iii) conversion of ammonia to urea and glutamine; iv) metabolism of aromatic amino acids; and v) proliferation in glucose-free and insulin-free medium. The C3A cell line is now well established as an in vitro model of the mature human liver (Mickelson et al. (1995) Hepatology 22:866-875; Nagendra et al. (1997) Am J Physiol 272:G408-G416).

Gemfibrozil is a fibric acid antilipemic agent that lowers serum triglycerides and produces favorable changes in lipoproteins. Gemfibrozil is effective in reducing the risk of coronary heart disease in men (Frick, M.H., et al. (1987) New Engl. J. Med; 317:1237-1245). The compound can inhibit peripheral lipolysis and decrease hepatic extraction of free fatty acids, which, decreases hepatic triglyceride production. Gemfibrozil also inhibits the synthesis and increases the clearance of apolipoprotein B, a carrier molecule for VLDL. Gemfibrozil has variable effects on LDL cholesterol. Although it causes moderate reductions in patients with type IIa hyperlipoproteinemia, changes in patients with either type IIb or type IV hyperlipoproteinemia are unpredictable. In general, the HMG-CoA reductase inhibitors are more effective than gemfibrozil in reducing LDL cholesterol. At the

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molecular level gemfibozil may function as a peroxisome proliferator-activated receptor (PPAR) agonist. Gemfibrozil is rapidly and completely absorbed from the GI tract and undergoes enterohepatic recirculation. Gemfibrozil is metabolized by the liver and excreted by the kidneys, mainly as metabolites, one of which possesses pharmacologic activity. Gemfibozil causes peroxisome proliferation and hepatocarcinogenesis in rats, which is a cause for concern generally for fibric acid derivative drugs. In humans, fibric acid derivatives are known to increase the risk of gall bladder disease although gemfibrozil is better tolerated than other fibrates. The relative safety of gemfibrozil in humans compared to rodent species including rats may be attributed to differences in metabolism and clearance of the compound in different species (Dix, K.J., et al., (1999) Drug Metab. Distrib. 27 (1) 138-146; Thomas, B.F., et al., (1999) Drug Metab. Distrib. 27 (1) 147-157). Lung cancer

Lung cancer is the leading cause of cancer death for men and the second leading cause of cancer death for women in the U.S. Lung cancers are divided into four histopathologically distinct groups. Three groups (squamous cell carcinoma, adenocarcinoma, and large cell carcinoma) are 15 classified as non-small cell lung cancers (NSCLCs). The fourth group of cancers is referred to as small cell lung cancer (SCLC). Deletions on chromosome 3 are common in this disease and are thought to indicate the presence of a tumor suppressor gene in this region. Activating mutations in Kras are commonly found in lung cancer and are the basis of one of the mouse models for the disease.

In nearly 80% of patients diagnosed with lung cancer, metastasis has already occurred. Most commonly lung cancers metastasize to pleura, brain, bone, pericardium, and liver. This adversely affects the overall five-year survival rate which is 37% for squamous carcinoma, 27% for adenocarcinoma and large cell carcinoma, and less than 1% for small cell carcinomas. Earlier diagnosis and an systematic approach to identification, staging, and treatment could positively affect patient outcome (DeVita et al. (1997) Cancer: Principles and Practice of Oncology, Lippincott-25 Raven, Philadelphia PA) and Fauci et al. (1998) Harrison's Principals of Internal Medicine, McGraw Hill, New York, NY).

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Lung cancers progress through a series of morphologically distinct stages from hyperplasia to invasive carcinoma. Malignant lung cancers are divided into two groups comprising four histopathological classes. The nonsmall cell lung carcinoma (NSCLC) group includes squamous cell carcinomas, adenocarcinomas, and large cell carcinomas and accounts for about 70% of all lung cancer cases. Adenocarcinomas typically arise in the peripheral airways and often form mucin secreting glands. Squamous cell carcinomas typically arise in proximal airways. The histogenesis of squamous cell carcinomas may be related to chronic inflammation and injury to the bronchial

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epithelium, leading to squamous metaplasia. The small cell lung carcinoma (SCLC) group accounts for about 20% of lung cancer cases. SCLCs typically arise in proximal airways and exhibit a number of paraneoplastic syndromes including inappropriate production of adrenocorticotropin and anti-diuretic hormone.

Lung cancer cells accumulate numerous genetic lesions, many of which are associated with cytologically visible chromosomal aberrations. The high frequency of chromosomal deletions associated with lung cancer may reflect the role of multiple tumor suppressor loci in the etiology of this disease. Several studies report deletions of regions of chromosome 11 in NSCLC (Bepler, G. and Garcia-Blanco, M.A. (1994) Proc. Natl. Acad. Sci. USA 91:5513-7; lizuka, M., et al. (1995) Genes, Chromosomes & Cancer 13:40-46; Rasio, D. (1995) Cancer Research 55:3988-91). Deletions in other chromosome arms such as 3p, 9p and 17p are also common. Other frequently observed genetic lesions include overexpression of telomerase, activation of oncogenes such as K-ras and c-myc, and inactivation of tumor suppressor genes such as RB, p53 and p16 (Toomey, D. et al.(2001) Cancer 92:2648-57; Zajac-Kaye M. (2001) Lung Cancer 34:S43-6; Wright, G. et al. (2000) Current Opinion in 15 Oncology 12:143-8; Kohno, T. and Yokota, J. (1999) Carcinogenesis 20:1403-10).

Ovarian cancer is the leading cause of death from a gynecologic cancer. The majority of ovarian cancers are derived from epithelial cells, and 70% of patients with epithelial ovarian cancers present with late-stage disease. As a result, the long-term survival rates for this disease is very low. Identification of early-stage markers for ovarian cancer would significantly increase the survival rate. The molecular events that lead to ovarian cancer are poorly understood. Some of the known aberrations include mutation of p53 and microsatellite instability. Since gene expression patterns are likely to vary when normal ovary is compared to ovarian tumors, examination of gene expression in these tissues to identify possible markers for ovarian cancer is particularly relevant to improving diagnosis, prognosis, and treatment of this disease.

Prostate Cancer

Ovarian Cancer

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Prostate cancer is a common malignancy in men over the age of 50, and the incidence increases with age. In the US, there are approximately 132,000 newly diagnosed cases of prostate cancer and more than 33,000 deaths from the disorder each year.

Once cancer cells arise in the prostate, they are stimulated by testosterone to a more rapid growth. Thus, removal of the testes can indirectly reduce both rapid growth and metastasis of the cancer. Over 95 percent of prostatic cancers are adenocarcinomas which originate in the prostatic

acini. The remaining 5 percent are divided between squamous cell and transitional cell carcinomas, both of which arise in the prostatic ducts or other parts of the prostate gland.

As with most cancers, prostate cancer develops through a multistage progression ultimately resulting in an aggressive, metastatic phenotype. The initial step in tumor progression involves the hyperproliferation of normal luminal and/or basal epithelial cells that become hyperplastic and evolve into early-stage tumors. The early-stage tumors are localized in the prostate but eventually may metastasize, particularly to the bone, brain or lung. About 80% of these tumors remain responsive to androgen treatment, an important hormone controlling the growth of prostate epithelial cells. However, in its most advanced state, cancer growth becomes androgen-independent and there is currently no known treatment for this condition.

A primary diagnostic marker for prostate cancer is prostate specific antigen (PSA). PSA is a tissue-specific serine protease almost exclusively produced by prostatic epithelial cells. The quantity of PSA correlates with the number and volume of the prostatic epithelial cells, and consequently, the levels of PSA are an excellent indicator of abnormal prostate growth. Men with prostate cancer exhibit an early linear increase in PSA levels followed by an exponential increase prior to diagnosis. However, since PSA levels are also influenced by factors such as inflammation, androgen and other growth factors, some scientists maintain that changes in PSA levels are not useful in detecting individual cases of prostate cancer.

Current areas of cancer research provide additional prospects for markers as well as potential
therapeutic targets for prostate cancer. Several growth factors have been shown to play a critical role
in tumor development, growth, and progression. The growth factors Epidermal Growth Factor (EGF),
Fibroblast Growth Factor (FGF), and Tumor Growth Factor alpha (TGFα) are important in the growth
of normal as well as hyperproliferative prostate epithelial cells, particularly at early stages of tumor
development and progression, and affect signaling pathways in these cells in various ways (Lin J et al.
(1999) Cancer Res. 59:2891-2897; Putz T et al. (1999) Cancer Res 59:227-233). The TGF-β family
of growth factors are generally expressed at increased levels in human cancers and the high
expression levels in many cases correlates with advanced stages of malignancy and poor survival
(Gold LI (1999) Crit Rev Oncog 10:303-360). Finally, there are human cell lines representing both the
androgen-dependent stage of prostate cancer (LNCap) as well as the androgen-independent, hormone
refractory stage of the disease (PC3 and DU-145) that have proved useful in studying gene
expression patterns associated with the progression of prostate cancer, and the effects of cell
treatments on these expressed genes (Chung TD (1999) Prostate 15:199-207).
Adipocyte Differentiation

The potential application of gene expression profiling is relevant to improving diagnosis, prognosis, and treatment of disease. For example, both the levels and sequences expressed in tissues from subjects with obesity or type II diabetes may be compared with the levels and sequences expressed in normal tissue.

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The primary function of adipose tissue is the ability to store and release fat during periods of feeding and fasting. White adipose tissue is the major energy reserve in periods of fasting, and its reserve is mobilized during energy deprivation. Adipose tissue is one of the primary target tissues for insulin, and adipogenesis and insulin resistance are linked in type II diabetes, non-insulin dependent diabetes mellitus (NIDDM). Cytologically the conversion of a preadipocytes into mature adipocytes is characterized by deposition of fat droplets around the nuclei. The conversion process in vivo can be induced by thiazolidinediones (TZDs) and other PPARy agonists (Adams et al. (1997) J. Clin. Invest. 100:3149-3153) which also lead to increased sensitivity to insulin and reduced plasma glucose and blood pressure.

Thiazolidinediones (TZDs) act as agonists for the peroxisome-proliferator-activated receptor gamma (PPARy), a member of the nuclear hormone receptor superfamily. TZDs reduce hyperglycemia, hyperinsulinemia, and hypertension, in part by promoting glucose metabolism and inhibiting gluconeogenesis. Roles for PPARy and its agonists have been demonstrated in a wide range of pathological conditions including diabetes, obesity, hypertension, atherosclerosis, polycystic ovarian syndrome, and cancers such as breast, prostate, liposarcoma, and colon cancer.

The mechanism by which TZDs and other PPAR\(\gamma\) agonists enhance insulin sensitivity is not fully understood, but may involve the ability of PPAR\(\gamma\) to promote adipogenesis. When ectopically expressed in cultured preadipocytes, PPAR\(\gamma\) is a potent inducer of adipocyte differentiation. TZDs, in combination with insulin and other factors, can also enhance differentiation of human preadipocytes in culture (Adams et al. (1997) J. Clin. Invest. 100:3149-3153). The relative potency of different TZDs in promoting adipogenesis in vitro is proportional to both their insulin sensitizing effects in vivo, and their ability to bind and activate PPAR\(\gamma\) in vitro. Interestingly, adipocytes derived from omental adipose depots are refractory to the effects of TZDs. It has therefore been suggested that the insulin sensitizing effects of TZDs may result from their ability to promote adipogenesis in subcutaneous adipose depots (Adams et al., ibid). Further, dominant negative mutations in the PPAR\(\gamma\) gene have been identified in two non-obese subjects with severe insulin resistance, hypertension, and overt non-insulin dependent diabetes mellitus (NIDDM) (Barroso et al. (1998) Nature 402:880-883).

NIDDM is the most common form of diabetes mellitus, a chronic metabolic disease that affects 143 million people worldwide. NIDDM is characterized by abnormal glucose and lipid

metabolism that result from a combination of peripheral insulin resistance and defective insulin secretion. NIDDM has a complex, progressive etiology and a high degree of heritability. Numerous complications of diabetes including heart disease, stroke, renal failure, retinopathy, and peripheral neuropathy contribute to the high rate of morbidity and mortality.

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At the molecular level, PPARy functions as a ligand activated transcription factor. In the presence of ligand, PPARy forms a heterodimer with the retinoid X receptor (RXR) which then activates transcription of target genes containing one or more copies of a PPARy response element (PPRE). Many genes important in lipid storage and metabolism contain PPREs and have been identified as PPARy targets, including PEPCK, aP2, LPL, ACS, and FAT-P (Auwerx, J. (1999) Diabetologia 42:1033-1049). Multiple ligands for PPARy have been identified. These include a variety of fatty acid metabolites; synthetic drugs belonging to the TZD class, such as Pioglitazone and Rosiglitazone (BRLA9653); and certain non-glitazone tyrosine analogs such as GI262570 and GW1929. The prostaglandin derivative 15-dPGJ2 is a potent endogenous ligand for PPARy.

Expression of PPARy is very high in adipose but barely detectable in skeletal muscle, the primary site for insulin stimulated glucose disposal in the body. PPARy is also moderately expressed in large intestine, kidney, liver, vascular smooth muscle, hematopoietic cells, and macrophages. The high expression of PPARy in adipose suggests that the insulin sensitizing effects of TZDs may result from alterations in the expression of one or more PPARy regulated genes in adipose tissue. Identification of PPARy target genes will contribute to better drug design and the development of novel therapeutic strategies for diabetes, obesity, and other conditions.

Systematic attempts to identify PPAR γ target genes have been made in several rodent models of obesity and diabetes (Suzuki et al. (2000) Jpn. J. Pharmacol. 84:113-123; Way et al. (2001) Endocrinology 142:1269-1277). However, a serious drawback of the rodent gene expression studies is that significant differences exist between human and rodent models of adipogenesis, diabetes, and obesity (Taylor (1999) Cell 97:9-12; Gregoire et al. (1998) Physiol. Reviews 78:783-809). Therefore, an unbiased approach to identifying TZD regulated genes in primary cultures of human tissues is necessary to fully elucidate the molecular basis for diseases associated with PPAR γ activity.

The majority of research in adipocyte biology to date has been done using transformed mouse preadipocyte cell lines. The culture condition, which stimulates mouse preadipocyte differentiation is different from that for inducing human primary preadipocyte differentiation. In addition, primary cells are diploid and may therefore reflect the <u>in vivo</u> context better than aneuploid cell lines.

Understanding the gene expression profile during adipogenesis in human will lead to understanding the fundamental mechanism of adiposity regulation. Furthermore, through comparing the gene expression

profiles of adipogenesis between donor with normal weight and donor with obesity, identification of crucial genes, potential drug targets for obesity and type 11 diabetes, will be possible.

Dendritic Cells

Dendritic cells (DC) are antigen presenting cells (APC) that play a key role in the primary immune response because of their unique ability to present antigens to naive T cells. In addition, DC differentiate into separate subsets that sustain and regulate immune responses following initial contact with antigen. DC subsets include those that preferentially induce particular T helper 1 (Th1) or T helper 2 (Th2) responses and those that regulate B cell responses. Moreover, DC are increasingly being used to manipulate immune responses, either to downregulate an aberrant autoimmune response or to enhance vaccination or a tumor-specific response.

DC are functionally specialized in correlation with their particular differentiation state. CD34+ myeloid cells found in the bone marrow mature in response to as yet unclear signals into CD14+ CD11c+ monocytes. An innate or antigen non-specific response takes place initially when monocytes circulate to nonlymphoid tissues and respond to lipopolysaccharide (LPS), a bacterially-derived mitogen, and viruses. Such direct encounter with antigen causes secretion of pro-inflammatory cytokines that attract and regulate natural killer cells, macrophages, and eosinophils in the first line of defense against invading pathogens. Monocytes then mature into DC, which capture antigen highly efficiently through endocytosis and antigen receptor uptake. Antigen processing and presentation trigger activation and differentiation into mature DC that express MHC class II molecules on the cell surface and efficiently activate T cells, initiating antigen-specific T cell and B cell responses. In turn, T cells activate DC through CD40 ligand - CD40 interactions, which stimulate expression of the costimulatory molecules CD80 and CD86, the latter most potent in amplifying T cell responses. DC interaction via CD40 with T cells also stimulates the production of inflammatory cytokines such as TNF alpha and IL-1. Engagement of RANK, a member of the TNF receptor family by its ligand, TRANCE, which is expressed on activated T cells, enhances the survival of DC through inhibition of apoptosis, thereby enhancing T cell activation. The maturation and differentiation of monocytes into mature DC links the antigen non-specific innate immune response to the antigen-specific adaptive immune response.

The process by which monocytes differentiate into immature dendritic cells in vivo has not been fully elucidated. Incubation of monocytes with granulocyte-macrophage colony stimulating factor (GM-CSF) and interleukin (IL) -4 in vitro yields cells that exhibit functional and morphological characteristics equivalent to immature dendritic cells found in vivo. Moreover, incubation in vitro of immature dendritic cells with tumor necrosis factor alpha (TNF-α), CD40 ligand, LPS, or monocyte-

conditioned medium yields mature dendritic cells that are potent activators of naive T cells.

The ability to manipulate DC in vitro and their capacity to mount an effective immune response with small numbers of DC and little antigen has led to potential immunotherapies for diseases such as cancer, AIDS, and infectious diseases; and enhancing vaccine efficacy. Spontaneous remissions of particular cancers such as renal cell carcinomas and melanomas indicate that the immune system can respond to tumor antigens and eliminate tumors. However, tumors escape immune surveillance through a number of means including secretion of IL-10, macrophage colony stimulating factor, IL-6, and vascular endothelial growth factor, all of which inhibit DC activity and promote tolerance of tumor tissue. Delivery of tumor antigen-loaded DC to tumors can induce tumor-specific rejection in animal models. Similarly, pathogens can escape immune surveillance by altering antigen processing and presentation pathways or interfering with maturation of antigen presenting cells. Rather than providing resistance, DC can complicate infection by hosting latent viruses such as Kaposi's virus and cytomegalovirus, complicating infection. HIV-1 and measles virus particles are efficiently produced in DC. Vaccines against tumors or infectious pathogens could be improved by systemic or local administration of DC loaded with tumor antigens or attenuated viral particles or components, respectively.

The expression of killer-inhibitor regulatory molecules, chemokines, chemokine receptors, and proteinases have been identified in DC through sequencing of ESTs. Continuing this search may reveal new lymphocyte-binding and antigen-processing molecules, transmembrane and secretory products, and transcription factors that may help to explain the specialized features of DC and allow manipulation of the immune system.

Alzheimer's Disease

Characterization of region-specific gene expression in the human brain provides a context and background for molecular neurobiology on a variety of neurological disorders. For example, Alzheimer's disease (AD) is a progressive, neurodestructive process of the human neocortex, characterized by the deterioration of memory and higher cognitive function. A progressive and irreversible brain disorder, AD is characterized by three major pathogenic episodes involving (a) an aberrant processing and deposition of beta-amyloid precursor protein (betaAPP) to form neurotoxic beta-amyloid (betaA) peptides and an aggregated insoluble polymer of betaA that forms the senile plaque, (b) the establishment of intraneuronal neuritic tau pathology yielding widespread deposits of agyrophilic neurofibrillary tangles (NFT) and (c) the initiation and proliferation of a brain-specific inflammatory response. These three seemingly disperse attributes of AD etiopathogenesis are linked by the fact that proinflammatory microglia, reactive astrocytes and their associated cytokines and

chemokines are associated with the biology of the microtubule associated protein tau, betaA speciation and aggregation. Missense mutations in the presentilin genes PS1 and PS2, implicated in early onset familial AD, cause abnormal betaAPP processing with resultant overproduction of betaA42 and related neurotoxic peptides. Specific betaA fragments such as betaA42 can further potentiate proinflammatory mechanisms. Expression of the inducible oxidoreductase cyclooxygenase-2 and cytosolic phospholipase A2 (cPLA2) is strongly activated during cerebral ischemia and trauma, epilepsy and AD, indicating the induction of proinflammatory gene pathways as a response to brain injury. Neurotoxic metals such as aluminum and zinc, both implicated in AD etiopathogenesis, and arachidonic acid, a major metabolite of brain cPLA2 activity, each polymerize hyperphosphorylated tau to form NFT-like bundles. Studies have identified a reduced risk for AD in patients aged over 70 years who were previously treated with non-steroidal anti-inflammatory drugs for non-CNS afflictions that include arthritis. (For a review of the interrelationships between the mechanisms of PS1, PS2 and betaAPP gene expression, tau and betaA deposition and the induction, regulation and proliferation in AD of the neuroinflammatory response, see Lukiw, W.J., and Bazan, N.G. (2000) Neurochem. Res. 2000 25:1173-1184).

There is a need in the art for new compositions, including nucleic acids and proteins, for the diagnosis, prevention, and treatment of cell proliferative, autoimmune/inflammatory, cardiovascular, neurological, and developmental disorders.

SUMMARY OF THE INVENTION

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Various embodiments of the invention provide purified polypeptides, secreted proteins, referred to collectively as 'SECP' and individually as 'SECP-1,' 'SECP-2,' 'SECP-3,' 'SECP-4,' 'SECP-5,' 'SECP-6,' 'SECP-7,' 'SECP-8,' 'SECP-9,' 'SECP-10,' 'SECP-11,' 'SECP-12,' 'SECP-13,' 'SECP-15,' 'SECP-16,' 'SECP-17,' 'SECP-18,' 'SECP-19,' 'SECP-20,' 'SECP-21,' 'SECP-22,' 'SECP-23,' 'SECP-24,' 'SECP-25,' 'SECP-26,' 'SECP-27,' 'SECP-28,' 'SECP-29,' 'SECP-30,' 'SECP-31,' 'SECP-32,' 'SECP-33,' 'SECP-34,' 'SECP-35,' 'SECP-36,' 'SECP-37,' 'SECP-38,' 'SECP-39,' 'SECP-40,' 'SECP-41,' 'SECP-42,' 'SECP-43,' 'SECP-44,' 'SECP-45,' 'SECP-46,' 'SECP-47,' 'SECP-48,' 'SECP-49,' 'SECP-50,' and 'SECP-51' and methods for using these proteins and their encoding polynucleotides for the detection, diagnosis, and treatment of diseases and medical conditions. Embodiments also provide methods for utilizing the purified secreted proteins and/or their encoding polynucleotides for facilitating the drug discovery process, including determination of efficacy, dosage, toxicity, and pharmacology. Related embodiments provide methods for utilizing the purified secreted proteins and/or their encoding polynucleotides for investigating the

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pathogenesis of diseases and medical conditions.

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An embodiment provides an isolated polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51. Another embodiment provides an isolated polypeptide comprising an amino acid sequence of SEQ ID NO:1-51.

Still another embodiment provides an isolated polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, c) a biologically active fragment of a polypeptide 15 having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51. In another embodiment, the polynucleotide encodes a polypeptide selected from the group consisting of SEQ ID NO:1-51. In an alternative embodiment, the polynucleotide is selected from the group consisting of SEQ ID NO:52-102.

Still another embodiment provides a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group 25 consisting of SEQ ID NO:1-51, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51. Another embodiment provides a cell transformed with the recombinant polynucleotide. Yet another embodiment provides a transgenic organism comprising the recombinant polynucleotide.

Another embodiment provides a method for producing a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, b) a polypeptide comprising a naturally occurring amino acid sequence at least

90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51. The method comprises a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding the polypeptide, and b) recovering the polypeptide so expressed.

Yet another embodiment provides an isolated antibody which specifically binds to a

polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence
selected from the group consisting of SEQ ID NO:1-51, b) a polypeptide comprising a naturally
occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid
sequence selected from the group consisting of SEQ ID NO:1-51, c) a biologically active fragment of
a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51,
and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the
group consisting of SEQ ID NO:1-51.

Still yet another embodiment provides an isolated polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:52-102, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:52-102, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). In other embodiments, the polynucleotide can comprise at least about 20, 30, 40, 60, 80, or 100 contiguous nucleotides.

Yet another embodiment provides a method for detecting a target polynucleotide in a sample, said target polynucleotide being selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:52-102, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:52-102, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence

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complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and b) detecting the presence or absence of said hybridization complex. In a related embodiment, the method can include detecting the amount of the hybridization complex. In still other embodiments, the probe can comprise at least about 20, 30, 40, 60, 80, or 100 contiguous nucleotides.

Still yet another embodiment provides a method for detecting a target polynucleotide in a sample, said target polynucleotide being selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:52-102, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:52-102, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof. In a related embodiment, the method can include detecting the amount of the amplified target polynucleotide or fragment thereof.

Another embodiment provides a composition comprising an effective amount of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, and a pharmaceutically acceptable excipient. In one embodiment, the composition can comprise an amino acid sequence selected from the group consisting of SEQ ID NO:1-51. Other embodiments provide a method of treating a disease or condition associated with decreased or abnormal expression of functional SECP, comprising administering to a patient in need of such treatment the composition.

Yet another embodiment provides a method for screening a compound for effectiveness as an agonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, b) a polypeptide comprising a

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naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting agonist activity in the sample. Another embodiment provides a composition comprising an agonist compound identified by the method and a pharmaceutically acceptable excipient. Yet another embodiment provides a method of treating a disease or condition associated with decreased expression of functional SECP, comprising administering to a patient in need of such treatment the composition.

Still yet another embodiment provides a method for screening a compound for effectiveness as an antagonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting antagonist activity in the sample. Another embodiment provides a composition comprising an antagonist compound identified by the method and a pharmaceutically acceptable excipient. Yet another embodiment provides a method of treating a disease or condition associated with overexpression of functional SECP, comprising administering to a patient in need of such treatment the composition.

Another embodiment provides a method of screening for a compound that specifically binds to
25 a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid
sequence selected from the group consisting of SEQ ID NO:1-51, b) a polypeptide comprising a
naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an
amino acid sequence selected from the group consisting of SEQ ID NO:1-51, c) a biologically active
fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ
30 ID NO:1-51, and d) an immunogenic fragment of a polypeptide having an amino acid sequence
selected from the group consisting of SEQ ID NO:1-51. The method comprises a) combining the
polypeptide with at least one test compound under suitable conditions, and b) detecting binding of the

polypeptide to the test compound, thereby identifying a compound that specifically binds to the polypeptide.

Yet another embodiment provides a method of screening for a compound that modulates the activity of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51. The method comprises a) combining the polypeptide with at least one test compound under conditions permissive for the activity of the polypeptide, b) assessing the activity of the polypeptide in the presence of the test compound, and c) comparing the activity of the polypeptide in the presence of the test compound with the activity of the polypeptide in the absence of the test compound, wherein a change in the activity of the polypeptide in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide.

Still yet another embodiment provides a method for screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a polynucleotide sequence selected from the group consisting of SEQ ID NO:52-102, the method comprising a) exposing a sample comprising the target polynucleotide to a compound, b) detecting altered expression of the target polynucleotide, and c) comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.

Another embodiment provides a method for assessing toxicity of a test compound, said

method comprising a) treating a biological sample containing nucleic acids with the test compound; b)

hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20

contiguous nucleotides of a polynucleotide selected from the group consisting of i) a polynucleotide

comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:52-102, ii) a

polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at

least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID

NO:52-102, iii) a polynucleotide having a sequence complementary to i), iv) a polynucleotide

complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Hybridization occurs

under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:52-102, ii) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:52-102, iii) a polynucleotide complementary to the polynucleotide of i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Alternatively, the target polynucleotide can comprise a fragment of a polynucleotide selected from the group consisting of i)-v) above; c) quantifying the amount of hybridization complex; and d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

BRIEF DESCRIPTION OF THE TABLES

Table 1 summarizes the nomenclature for full length polynucleotide and polypeptide embodiments of the invention.

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Table 2 shows the GenBank identification number and annotation of the nearest GenBank homolog, and the PROTEOME database identification numbers and annotations of PROTEOME database homologs, for polypeptides of the invention. The probability scores for the matches between each polypeptide and its homolog(s) are also shown.

Table 3 shows structural features of polypeptide embodiments, including predicted motifs and domains, along with the methods, algorithms, and searchable databases used for analysis of the polypeptides.

Table 4 lists the cDNA and/or genomic DNA fragments which were used to assemble polynucleotide embodiments, along with selected fragments of the polynucleotides.

Table 5 shows representative cDNA libraries for polynucleotide embodiments.

Table 6 provides an appendix which describes the tissues and vectors used for construction of the cDNA libraries shown in Table 5.

Table 7 shows the tools, programs, and algorithms used to analyze polynucleotides and polypeptides, along with applicable descriptions, references, and threshold parameters.

Table 8 shows single nucleotide polymorphisms found in polynucleotide sequences of the invention, along with allele frequencies in different human populations.

DESCRIPTION OF THE INVENTION

Before the present proteins, nucleic acids, and methods are described, it is understood that embodiments of the invention are not limited to the particular machines, instruments, materials, and methods described, as these may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the invention.

As used herein and in the appended claims, the singular forms "a," "an," and "the" include plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to "a host cell" includes a plurality of such host cells, and a reference to "an antibody" is a reference to one or more antibodies and equivalents thereof known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any machines, materials, and methods similar or equivalent to those described herein can be used to practice or test the present invention, the preferred machines, materials and methods are now described. All publications mentioned herein are cited for the purpose of describing and disclosing the cell lines, protocols, reagents and vectors which are reported in the publications and which might be used in connection with various embodiments of the invention. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention.

DEFINITIONS

"SECP" refers to the amino acid sequences of substantially purified SECP obtained from any species, particularly a mammalian species, including bovine, ovine, porcine, murine, equine, and human, and from any source, whether natural, synthetic, semi-synthetic, or recombinant.

The term "agonist" refers to a molecule which intensifies or mimics the biological activity of SECP. Agonists may include proteins, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of SECP either by directly interacting with SECP or by acting on components of the biological pathway in which SECP participates.

An "allelic variant" is an alternative form of the gene encoding SECP. Allelic variants may result from at least one mutation in the nucleic acid sequence and may result in altered mRNAs or in polypeptides whose structure or function may or may not be altered. A gene may have none, one, or many allelic variants of its naturally occurring form. Common mutational changes which give rise to allelic variants are generally ascribed to natural deletions, additions, or substitutions of nucleotides.

Each of these types of changes may occur alone, or in combination with the others, one or more times in a given sequence.

"Altered" nucleic acid sequences encoding SECP include those sequences with deletions, insertions, or substitutions of different nucleotides, resulting in a polypeptide the same as SECP or a polypeptide with at least one functional characteristic of SECP. Included within this definition are polymorphisms which may or may not be readily detectable using a particular oligonucleotide probe of the polynucleotide encoding SECP, and improper or unexpected hybridization to allelic variants, with a locus other than the normal chromosomal locus for the polynucleotide encoding SECP. The encoded protein may also be "altered," and may contain deletions, insertions, or substitutions of amino acid residues which produce a silent change and result in a functionally equivalent SECP. Deliberate amino acid substitutions may be made on the basis of one or more similarities in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues, as long as the biological or immunological activity of SECP is retained. For example, negatively charged amino acids may include aspartic acid and glutamic acid, and positively charged amino acids may include lysine and arginine. Amino acids with uncharged polar side chains having similar hydrophilicity values may include: asparagine and glutamine; and serine and threonine. Amino acids with uncharged side chains having similar hydrophilicity values may include: leucine, isoleucine, and valine; glycine and alanine; and phenylalanine and tyrosine.

The terms "amino acid" and "amino acid sequence" can refer to an oligopeptide, a peptide, a polypeptide, or a protein sequence, or a fragment of any of these, and to naturally occurring or synthetic molecules. Where "amino acid sequence" is recited to refer to a sequence of a naturally occurring protein molecule, "amino acid sequence" and like terms are not meant to limit the amino acid sequence to the complete native amino acid sequence associated with the recited protein molecule.

"Amplification" relates to the production of additional copies of a nucleic acid. Amplification may be carried out using polymerase chain reaction (PCR) technologies or other nucleic acid amplification technologies well known in the art.

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The term "antagonist" refers to a molecule which inhibits or attenuates the biological activity of SECP. Antagonists may include proteins such as antibodies, anticalins, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of SECP either by directly interacting with SECP or by acting on components of the biological pathway in which SECP participates.

The term "antibody" refers to intact immunoglobulin molecules as well as to fragments

thereof, such as Fab, F(ab')₂, and Fv fragments, which are capable of binding an epitopic determinant. Antibodies that bind SECP polypeptides can be prepared using intact polypeptides or using fragments containing small peptides of interest as the immunizing antigen. The polypeptide or oligopeptide used to immunize an animal (e.g., a mouse, a rat, or a rabbit) can be derived from the translation of RNA, or synthesized chemically, and can be conjugated to a carrier protein if desired. Commonly used carriers that are chemically coupled to peptides include bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin (KLH). The coupled peptide is then used to immunize the animal.

The term "antigenic determinant" refers to that region of a molecule (i.e., an epitope) that makes contact with a particular antibody. When a protein or a fragment of a protein is used to immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to antigenic determinants (particular regions or three-dimensional structures on the protein). An antigenic determinant may compete with the intact antigen (i.e., the immunogen used to elicit the immune response) for binding to an antibody.

The term "aptamer" refers to a nucleic acid or oligonucleotide molecule that binds to a

specific molecular target. Aptamers are derived from an *in vitro* evolutionary process (e.g., SELEX.

(Systematic Evolution of Ligands by EXponential Enrichment), described in U.S. Patent No.

5,270,163), which selects for target-specific aptamer sequences from large combinatorial libraries.

Aptamer compositions may be double-stranded or single-stranded, and may include deoxyribonucleotides, ribonucleotides, nucleotide derivatives, or other nucleotide-like molecules. The nucleotide components of an aptamer may have modified sugar groups (e.g., the 2'-OH group of a ribonucleotide may be replaced by 2'-F or 2'-NH₂), which may improve a desired property, e.g., resistance to nucleases or longer lifetime in blood. Aptamers may be conjugated to other molecules, e.g., a high molecular weight carrier to slow clearance of the aptamer from the circulatory system. Aptamers may be specifically cross-linked to their cognate ligands, e.g., by photo-activation of a cross-linker (Brody, E.N. and L. Gold (2000) J. Biotechnol. 74:5-13).

The term "intramer" refers to an aptamer which is expressed *in vivo*. For example, a vaccinia virus-based RNA expression system has been used to express specific RNA aptamers at high levels in the cytoplasm of leukocytes (Blind, M. et al. (1999) Proc. Natl. Acad. Sci. USA 96:3606-3610).

The term "spiegelmer" refers to an aptamer which includes L-DNA, L-RNA, or other lefthanded nucleotide derivatives or nucleotide-like molecules. Aptamers containing left-handed nucleotides are resistant to degradation by naturally occurring enzymes, which normally act on

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substrates containing right-handed nucleotides.

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The term "antisense" refers to any composition capable of base-pairing with the "sense"

(coding) strand of a polynucleotide having a specific nucleic acid sequence. Antisense compositions may include DNA; RNA; peptide nucleic acid (PNA); oligonucleotides having modified backbone

5 linkages such as phosphorothioates, methylphosphonates, or benzylphosphonates; oligonucleotides having modified sugar groups such as 2'-methoxyethyl sugars or 2'-methoxyethoxy sugars; or oligonucleotides having modified bases such as 5-methyl cytosine, 2'-deoxyuracil, or 7-deaza-2'-deoxyguanosine. Antisense molecules may be produced by any method including chemical synthesis or transcription. Once introduced into a cell, the complementary antisense molecule base-pairs with a naturally occurring nucleic acid sequence produced by the cell to form duplexes which block either transcription or translation. The designation "negative" or "minus" can refer to the antisense strand, and the designation "positive" or "plus" can refer to the sense strand of a reference DNA molecule.

The term "biologically active" refers to a protein having structural, regulatory, or biochemical functions of a naturally occurring molecule. Likewise, "immunologically active" or "immunogenic" refers to the capability of the natural, recombinant, or synthetic SECP, or of any oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

"Complementary" describes the relationship between two single-stranded nucleic acid sequences that anneal by base-pairing. For example, 5'-AGT-3' pairs with its complement, 3'-TCA-5'.

A "composition comprising a given polynucleotide" and a "composition comprising a given polypeptide" can refer to any composition containing the given polynucleotide or polypeptide. The composition may comprise a dry formulation or an aqueous solution. Compositions comprising polynucleotides encoding SECP or fragments of SECP may be employed as hybridization probes. The probes may be stored in freeze-dried form and may be associated with a stabilizing agent such as a carbohydrate. In hybridizations, the probe may be deployed in an aqueous solution containing salts (e.g., NaCl), detergents (e.g., sodium dodecyl sulfate; SDS), and other components (e.g., Denhardt's solution, dry milk, salmon sperm DNA, etc.).

"Consensus sequence" refers to a nucleic acid sequence which has been subjected to repeated DNA sequence analysis to resolve uncalled bases, extended using the XL-PCR kit (Applied Biosystems, Foster City CA) in the 5' and/or the 3' direction, and resequenced, or which has been assembled from one or more overlapping cDNA, EST, or genomic DNA fragments using a computer

program for fragment assembly, such as the GELVIEW fragment assembly system (Accelrys, Burlington MA) or Phrap (University of Washington, Seattle WA). Some sequences have been both extended and assembled to produce the consensus sequence.

"Conservative amino acid substitutions" are those substitutions that are predicted to least interfere with the properties of the original protein, i.e., the structure and especially the function of the protein is conserved and not significantly changed by such substitutions. The table below shows amino acids which may be substituted for an original amino acid in a protein and which are regarded as conservative amino acid substitutions.

	Original Residue	Conservative Substitution
10	Ala	Gly, Ser
	Arg	His, Lys
	Asn	Asp, Gln, His
	Asp	Asn, Glu
	Cys	Ala, Ser
15	Gin	Asn, Glu, His
	Glu	Asp, Gln, His
	Gly	Ala
	His	Asn, Arg, Gln, Glu
	Ile	Leu, Val
20	Leu	Ile, Val
	Lys	Arg, Gln, Glu
	Met	Leu, Ile
	Phe	His, Met, Leu, Trp, Tyr
	Ser .	Cys, Thr
25	Thr	Ser, Val
	Trp	Phe, Tyr
	Tyr	His, Phe, Trp
	Val	Ile, Leu, Thr

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Conservative amino acid substitutions generally maintain (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a beta sheet or alpha helical conformation, (b) the charge or hydrophobicity of the molecule at the site of the substitution, and/or (c) the bulk of the side chain.

A "deletion" refers to a change in the amino acid or nucleotide sequence that results in the absence of one or more amino acid residues or nucleotides.

The term "derivative" refers to a chemically modified polynucleotide or polypeptide.

Chemical modifications of a polynucleotide can include, for example, replacement of hydrogen by an alkyl, acyl, hydroxyl, or amino group. A derivative polynucleotide encodes a polypeptide which retains at least one biological or immunological function of the natural molecule. A derivative polypeptide is

one modified by glycosylation, pegylation, or any similar process that retains at least one biological or immunological function of the polypeptide from which it was derived.

A "detectable label" refers to a reporter molecule or enzyme that is capable of generating a measurable signal and is covalently or noncovalently joined to a polynucleotide or polypeptide.

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"Differential expression" refers to increased or upregulated; or decreased, downregulated, or absent gene or protein expression, determined by comparing at least two different samples. Such comparisons may be carried out between, for example, a treated and an untreated sample, or a diseased and a normal sample.

"Exon shuffling" refers to the recombination of different coding regions (exons). Since an exon may represent a structural or functional domain of the encoded protein, new proteins may be assembled through the novel reassortment of stable substructures, thus allowing acceleration of the evolution of new protein functions.

A "fragment" is a unique portion of SECP or a polynucleotide encoding SECP which can be identical in sequence to, but shorter in length than, the parent sequence. A fragment may comprise up to the entire length of the defined sequence, minus one nucleotide/amino acid residue. For example, a fragment may comprise from about 5 to about 1000 contiguous nucleotides or amino acid residues. A fragment used as a probe, primer, antigen, therapeutic molecule, or for other purposes, may be at least 5, 10, 15, 16, 20, 25, 30, 40, 50, 60, 75, 100, 150, 250 or at least 500 contiguous nucleotides or amino acid residues in length. Fragments may be preferentially selected from certain regions of a molecule.

For example, a polypeptide fragment may comprise a certain length of contiguous amino acids selected from the first 250 or 500 amino acids (or first 25% or 50%) of a polypeptide as shown in a certain defined sequence. Clearly these lengths are exemplary, and any length that is supported by the specification, including the Sequence Listing, tables, and figures, may be encompassed by the present embodiments.

A fragment of SEQ ID NO:52-102 can comprise a region of unique polynucleotide sequence that specifically identifies SEQ ID NO:52-102, for example, as distinct from any other sequence in the genome from which the fragment was obtained. A fragment of SEQ ID NO:52-102 can be employed in one or more embodiments of methods of the invention, for example, in hybridization and amplification technologies and in analogous methods that distinguish SEQ ID NO:52-102 from related polynucleotides. The precise length of a fragment of SEQ ID NO:52-102 and the region of SEQ ID NO:52-102 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A fragment of SEQ ID NO:1-51 is encoded by a fragment of SEQ ID NO:52-102. A fragment of SEQ ID NO:1-51 can comprise a region of unique amino acid sequence that specifically identifies SEQ ID NO:1-51. For example, a fragment of SEQ ID NO:1-51 can be used as an immunogenic peptide for the development of antibodies that specifically recognize SEQ ID NO:1-51. The precise length of a fragment of SEQ ID NO:1-51 and the region of SEQ ID NO:1-51 to which the fragment corresponds can be determined based on the intended purpose for the fragment using one or more analytical methods described herein or otherwise known in the art.

A "full length" polynucleotide is one containing at least a translation initiation codon (e.g., methionine) followed by an open reading frame and a translation termination codon. A "full length" polynucleotide sequence encodes a "full length" polypeptide sequence.

"Homology" refers to sequence similarity or, alternatively, sequence identity, between two or more polynucleotide sequences or two or more polypeptide sequences.

The terms "percent identity" and "% identity," as applied to polynucleotide sequences, refer to the percentage of identical nucleotide matches between at least two polynucleotide sequences aligned using a standardized algorithm. Such an algorithm may insert, in a standardized and reproducible way, gaps in the sequences being compared in order to optimize alignment between two sequences, and therefore achieve a more meaningful comparison of the two sequences.

Percent identity between polynucleotide sequences may be determined using one or more computer algorithms or programs known in the art or described herein. For example, percent identity can be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program. This program is part of the LASERGENE software package, a suite of molecular biological analysis programs (DNASTAR, Madison WI). CLUSTAL V is described in Higgins, D.G. and P.M. Sharp (1989; CABIOS 5:151-153) and in Higgins, D.G. et al. (1992; CABIOS 8:189-191). For pairwise alignments of polynucleotide sequences, the default parameters are set as follows: Ktuple=2, gap penalty=5, window=4, and "diagonals saved"=4. The "weighted" residue weight table is selected as the default.

Alternatively, a suite of commonly used and freely available sequence comparison algorithms which can be used is provided by the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search Tool (BLAST) (Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410), which is available from several sources, including the NCBI, Bethesda, MD, and on the Internet at http://www.ncbi.nlm.nih.gov/BLAST/. The BLAST software suite includes various sequence analysis programs including "blastn," that is used to align a known polynucleotide sequence with other

polynucleotide sequences from a variety of databases. Also available is a tool called "BLAST 2 Sequences" that is used for direct pairwise comparison of two nucleotide sequences. "BLAST 2 Sequences" can be accessed and used interactively at http://www.ncbi.nlm.nih.gov/gorf/b12.html. The "BLAST 2 Sequences" tool can be used for both blastn and blastp (discussed below). BLAST 5 programs are commonly used with gap and other parameters set to default settings. For example, to compare two nucleotide sequences, one may use blastn with the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Reward for match: 1

10 Penalty for mismatch: -2

Open Gap: 5 and Extension Gap: 2 penalties

Gap x drop-off: 50

Expect: 10

Word Size: 11

15 Filter: on

Percent identity may be measured over the length of an entire defined sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined sequence, for instance, a fragment of at least 20, at least 30, at least 40, at least 50, at least 70, at least 100, or at least 200 contiguous 20 nucleotides. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures, or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

Nucleic acid sequences that do not show a high degree of identity may nevertheless encode similar amino acid sequences due to the degeneracy of the genetic code. It is understood that changes in a nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid sequences that all encode substantially the same protein.

The phrases "percent identity" and "% identity," as applied to polypeptide sequences, refer to the percentage of identical residue matches between at least two polypeptide sequences aligned using a standardized algorithm. Methods of polypeptide sequence alignment are well-known. Some alignment methods take into account conservative amino acid substitutions. Such conservative substitutions, explained in more detail above, generally preserve the charge and hydrophobicity at the site of substitution, thus preserving the structure (and therefore function) of the polypeptide. The

phrases "percent similarity" and "% similarity," as applied to polypeptide sequences, refer to the percentage of residue matches, including identical residue matches and conservative substitutions, between at least two polypeptide sequences aligned using a standardized algorithm. In contrast, conservative substitutions are not included in the calculation of percent identity between polypeptide sequences.

Percent identity between polypeptide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program (described and referenced above). For pairwise alignments of polypeptide sequences using CLUSTAL V, the default parameters are set as follows: Ktuple=1, gap penalty=3, window=5, and "diagonals saved"=5. The PAM250 matrix is selected as the default residue weight table.

Alternatively the NCBI BLAST software suite may be used. For example, for a pairwise comparison of two polypeptide sequences, one may use the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) with blastp set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Open Gap: 11 and Extension Gap: 1 penalties

Gap x drop-off: 50

Expect: 10

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Percent identity may be measured over the length of an entire defined polypeptide sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined polypeptide sequence, for instance, a fragment of at least 15, at least 20, at least 30, at least 40, at least 50, at least 70 or at least 150 contiguous residues. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

"Human artificial chromosomes" (HACs) are linear microchromosomes which may contain DNA sequences of about 6 kb to 10 Mb in size and which contain all of the elements required for chromosome replication, segregation and maintenance.

The term "humanized antibody" refers to an antibody molecule in which the amino acid

sequence in the non-antigen binding regions has been altered so that the antibody more closely resembles a human antibody, and still retains its original binding ability.

"Hybridization" refers to the process by which a polynucleotide strand anneals with a complementary strand through base pairing under defined hybridization conditions. Specific hybridization is an indication that two nucleic acid sequences share a high degree of complementarity. Specific hybridization complexes form under permissive annealing conditions and remain hybridized after the "washing" step(s). The washing step(s) is particularly important in determining the stringency of the hybridization process, with more stringent conditions allowing less non-specific binding, i.e., binding between pairs of nucleic acid strands that are not perfectly matched. Permissive conditions for annealing of nucleic acid sequences are routinely determinable by one of ordinary skill in the art and may be consistent among hybridization experiments, whereas wash conditions may be varied among experiments to achieve the desired stringency, and therefore hybridization specificity. Permissive annealing conditions occur, for example, at 68°C in the presence of about 6 x SSC, about 1% (w/v) SDS, and about 100 μg/ml sheared, denatured salmon sperm DNA.

Generally, stringency of hybridization is expressed, in part, with reference to the temperature under which the wash step is carried out. Such wash temperatures are typically selected to be about 5°C to 20°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. An equation for calculating T_m and conditions for nucleic acid hybridization are well known and can be found in Sambrook, J. and D.W. Russell (2001; Molecular Cloning: A Laboratory Manual, 3rd ed., vol. 1-3, Cold Spring Harbor Press, Cold Spring Harbor NY, ch. 9).

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High stringency conditions for hybridization between polynucleotides of the present invention include wash conditions of 68°C in the presence of about 0.2 x SSC and about 0.1% SDS, for 1 hour. Alternatively, temperatures of about 65°C, 60°C, 55°C, or 42°C may be used. SSC concentration may be varied from about 0.1 to 2 x SSC, with SDS being present at about 0.1%. Typically, blocking reagents are used to block non-specific hybridization. Such blocking reagents include, for instance, sheared and denatured salmon sperm DNA at about 100-200 µg/ml. Organic solvent, such as formamide at a concentration of about 35-50% v/v, may also be used under particular circumstances, such as for RNA:DNA hybridizations. Useful variations on these wash conditions will be readily apparent to those of ordinary skill in the art. Hybridization, particularly under high stringency conditions, may be suggestive of evolutionary similarity between the nucleotides. Such similarity is

strongly indicative of a similar role for the nucleotides and their encoded polypeptides.

The term "hybridization complex" refers to a complex formed between two nucleic acids by virtue of the formation of hydrogen bonds between complementary bases. A hybridization complex may be formed in solution (e.g., Cot or Rot analysis) or formed between one nucleic acid present in solution and another nucleic acid immobilized on a solid support (e.g., paper, membranes, filters, chips, pins or glass slides, or any other appropriate substrate to which cells or their nucleic acids have been fixed).

The words "insertion" and "addition" refer to changes in an amino acid or polynucleotide sequence resulting in the addition of one or more amino acid residues or nucleotides, respectively.

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"Immune response" can refer to conditions associated with inflammation, trauma, immune disorders, or infectious or genetic disease, etc. These conditions can be characterized by expression of various factors, e.g., cytokines, chemokines, and other signaling molecules, which may affect cellular and systemic defense systems.

An "immunogenic fragment" is a polypeptide or oligopeptide fragment of SECP which is

capable of eliciting an immune response when introduced into a living organism, for example, a

mammal. The term "immunogenic fragment" also includes any polypeptide or oligopeptide fragment

of SECP which is useful in any of the antibody production methods disclosed herein or known in the

art.

The term "microarray" refers to an arrangement of a plurality of polynucleotides, polypeptides, antibodies, or other chemical compounds on a substrate.

The terms "element" and "array element" refer to a polynucleotide, polypeptide, antibody, or other chemical compound having a unique and defined position on a microarray.

The term "modulate" refers to a change in the activity of SECP. For example, modulation may cause an increase or a decrease in protein activity, binding characteristics, or any other biological, functional, or immunological properties of SECP.

The phrases "nucleic acid" and "nucleic acid sequence" refer to a nucleotide, oligonucleotide, polynucleotide, or any fragment thereof. These phrases also refer to DNA or RNA of genomic or synthetic origin which may be single-stranded or double-stranded and may represent the sense or the antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material.

"Operably linked" refers to the situation in which a first nucleic acid sequence is placed in a functional relationship with a second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding

sequence. Operably linked DNA sequences may be in close proximity or contiguous and, where necessary to join two protein coding regions, in the same reading frame.

"Peptide nucleic acid" (PNA) refers to an antisense molecule or anti-gene agent which comprises an oligonucleotide of at least about 5 nucleotides in length linked to a peptide backbone of amino acid residues ending in lysine. The terminal lysine confers solubility to the composition. PNAs preferentially bind complementary single stranded DNA or RNA and stop transcript elongation, and may be pegylated to extend their lifespan in the cell.

"Post-translational modification" of an SECP may involve lipidation, glycosylation, phosphorylation, acetylation, racemization, proteolytic cleavage, and other modifications known in the art. These processes may occur synthetically or biochemically. Biochemical modifications will vary by cell type depending on the enzymatic milieu of SECP.

"Probe" refers to nucleic acids encoding SECP, their complements, or fragments thereof, which are used to detect identical, allelic or related nucleic acids. Probes are isolated oligonucleotides or polynucleotides attached to a detectable label or reporter molecule. Typical labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes. "Primers" are short nucleic acids, usually DNA oligonucleotides, which may be annealed to a target polynucleotide by complementary base-pairing. The primer may then be extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification (and identification) of a nucleic acid, e.g., by the polymerase chain reaction (PCR).

Probes and primers as used in the present invention typically comprise at least 15 contiguous nucleotides of a known sequence. In order to enhance specificity, longer probes and primers may also be employed, such as probes and primers that comprise at least 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, or at least 150 consecutive nucleotides of the disclosed nucleic acid sequences. Probes and primers may be considerably longer than these examples, and it is understood that any length supported by the specification, including the tables, figures, and Sequence Listing, may be used.

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Methods for preparing and using probes and primers are described in, for example, Sambrook, J. and D.W. Russell (2001; Molecular Cloning: A Laboratory Manual, 3rd ed., vol. 1-3, Cold Spring Harbor Press, Cold Spring Harbor NY), Ausubel, F.M. et al. (1999; Short Protocols in Molecular Biology, 4th ed., John Wiley & Sons, New York NY), and Innis, M. et al. (1990; PCR Protocols, A Guide to Methods and Applications, Academic Press, San Diego CA). PCR primer pairs can be derived from a known sequence, for example, by using computer programs intended for that purpose such as Primer (Version 0.5, 1991, Whitehead Institute for Biomedical Research, Cambridge MA).

Oligonucleotides for use as primers are selected using software known in the art for such purpose. For example, OLIGO 4.06 software is useful for the selection of PCR primer pairs of up to 100 nucleotides each, and for the analysis of oligonucleotides and larger polynucleotides of up to 5,000 nucleotides from an input polynucleotide sequence of up to 32 kilobases. Similar primer selection programs have incorporated additional features for expanded capabilities. For example, the PrimOU primer selection program (available to the public from the Genome Center at University of Texas South West Medical Center, Dallas TX) is capable of choosing specific primers from megabase sequences and is thus useful for designing primers on a genome-wide scope. The Primer3 primer selection program (available to the public from the Whitehead Institute/MIT Center for Genome 10 Research, Cambridge MA) allows the user to input a "mispriming library," in which sequences to avoid as primer binding sites are user-specified. Primer3 is useful, in particular, for the selection of oligonucleotides for microarrays. (The source code for the latter two primer selection programs may also be obtained from their respective sources and modified to meet the user's specific needs.) The PrimeGen program (available to the public from the UK Human Genome Mapping Project Resource 15 Centre, Cambridge UK) designs primers based on multiple sequence alignments, thereby allowing selection of primers that hybridize to either the most conserved or least conserved regions of aligned nucleic acid sequences. Hence, this program is useful for identification of both unique and conserved oligonucleotides and polynucleotide fragments. The oligonucleotides and polynucleotide fragments identified by any of the above selection methods are useful in hybridization technologies, for example, as PCR or sequencing primers, microarray elements, or specific probes to identify fully or partially complementary polynucleotides in a sample of nucleic acids. Methods of oligonucleotide selection are not limited to those described above.

A "recombinant nucleic acid" is a nucleic acid that is not naturally occurring or has a sequence that is made by an artificial combination of two or more otherwise separated segments of sequence. This artificial combination is often accomplished by chemical synthesis or, more commonly, by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques such as those described in Sambrook and Russell (supra). The term recombinant includes nucleic acids that have been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently, a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter sequence. Such a recombinant nucleic acid may be part of a vector that is used, for example, to transform a cell.

Alternatively, such recombinant nucleic acids may be part of a viral vector, e.g., based on a

vaccinia virus, that could be use to vaccinate a mammal wherein the recombinant nucleic acid is expressed, inducing a protective immunological response in the mammal.

A "regulatory element" refers to a nucleic acid sequence usually derived from untranslated regions of a gene and includes enhancers, promoters, introns, and 5' and 3' untranslated regions

(UTRs). Regulatory elements interact with host or viral proteins which control transcription, translation, or RNA stability.

"Reporter molecules" are chemical or biochemical moieties used for labeling a nucleic acid, amino acid, or antibody. Reporter molecules include radionuclides; enzymes; fluorescent, chemiluminescent, or chromogenic agents; substrates; cofactors; inhibitors; magnetic particles; and other moieties known in the art.

An "RNA equivalent," in reference to a DNA molecule, is composed of the same linear sequence of nucleotides as the reference DNA molecule with the exception that all occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

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The term "sample" is used in its broadest sense. A sample suspected of containing SECP, nucleic acids encoding SECP, or fragments thereof may comprise a bodily fluid; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; a cell; genomic DNA, RNA, or cDNA, in solution or bound to a substrate; a tissue; a tissue print; etc.

The terms "specific binding" and "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, an antagonist, a small molecule, or any natural or synthetic binding composition. The interaction is dependent upon the presence of a particular structure of the protein, e.g., the antigenic determinant or epitope, recognized by the binding molecule. For example, if an antibody is specific for epitope "A," the presence of a polypeptide comprising the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the antibody will reduce the amount of labeled A that binds to the antibody.

The term "substantially purified" refers to nucleic acid or amino acid sequences that are removed from their natural environment and are isolated or separated, and are at least about 60% free, preferably at least about 75% free, and most preferably at least about 90% free from other components with which they are naturally associated.

A "substitution" refers to the replacement of one or more amino acid residues or nucleotides by different amino acid residues or nucleotides, respectively.

"Substrate" refers to any suitable rigid or semi-rigid support including membranes, filters,

chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which polynucleotides or polypeptides are bound.

A "transcript image" or "expression profile" refers to the collective pattern of gene expression by a particular cell type or tissue under given conditions at a given time.

"Transformation" describes a process by which exogenous DNA is introduced into a recipient cell. Transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transformation is selected based 10 on the type of host cell being transformed and may include, but is not limited to, bacteriophage or viral infection, electroporation, heat shock, lipofection, and particle bombardment. The term "transformed cells" includes stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome, as well as transiently transformed cells which express the inserted DNA or RNA for limited periods of time.

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A "transgenic organism," as used herein, is any organism, including but not limited to animals and plants, in which one or more of the cells of the organism contains heterologous nucleic acid introduced by way of human intervention, such as by transgenic techniques well known in the art. The nucleic acid is introduced into the cell, directly or indirectly by introduction into a precursor of the cell, by way of deliberate genetic manipulation, such as by microinjection or by infection with a recombinant virus. In another embodiment, the nucleic acid can be introduced by infection with a recombinant viral vector, such as a lentiviral vector (Lois, C. et al. (2002) Science 295:868-872). The term genetic manipulation does not include classical cross-breeding, or in vitro fertilization, but rather is directed to the introduction of a recombinant DNA molecule. The transgenic organisms contemplated in accordance with the present invention include bacteria, cyanobacteria, fungi, plants 25 and animals. The isolated DNA of the present invention can be introduced into the host by methods known in the art, for example infection, transfection, transformation or transconjugation. Techniques for transferring the DNA of the present invention into such organisms are widely known and provided in references such as Sambrook and Russell (supra).

A "variant" of a particular nucleic acid sequence is defined as a nucleic acid sequence having at least 40% sequence identity to the particular nucleic acid sequence over a certain length of one of the nucleic acid sequences using blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of nucleic acids may show, for example, at least 50%, at

least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a certain defined length. A variant may be described as, for example, an "allelic" (as defined above), "splice," "species," or "polymorphic" variant. A splice variant may have significant identity to a reference molecule, but will generally have a greater or lesser number of polynucleotides due to alternate splicing during mRNA processing. The corresponding polypeptide may possess additional functional domains or lack domains that are present in the reference molecule. Species variants are polynucleotides that vary from one species to another. The resulting polypeptides will generally have significant amino acid identity relative to each other. A polymorphic variant is a variation in the polynucleotide sequence of a particular gene between individuals of a given species. Polymorphic variants also may encompass "single nucleotide polymorphisms" (SNPs) in which the polynucleotide sequence varies by one nucleotide base. The presence of SNPs may be indicative of, for example, a certain population, a disease state, or a propensity for a disease state.

A "variant" of a particular polypeptide sequence is defined as a polypeptide sequence having at least 40% sequence identity or sequence similarity to the particular polypeptide sequence over a certain length of one of the polypeptide sequences using blastp with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of polypeptides may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity or sequence similarity over a certain defined length of one of the polypeptides.

THE INVENTION

Various embodiments of the invention include new human secreted proteins (SECP), the polynucleotides encoding SECP, and the use of these compositions for the diagnosis, treatment, or prevention of cell proliferative, autoimmune/inflammatory, cardiovascular, neurological, and developmental disorders.

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide embodiments of the invention. Each polynucleotide and its corresponding polypeptide are correlated to a single Incyte project identification number (Incyte Project ID). Each polypeptide sequence is denoted by both a polypeptide sequence identification number (Polypeptide SEQ ID NO:) and an Incyte polypeptide sequence number (Incyte Polypeptide ID) as shown. Each polynucleotide

sequence is denoted by both a polynucleotide sequence identification number (Polynucleotide SEQ ID NO:) and an Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) as shown. Column 6 shows the Incyte ID numbers of physical, full length clones corresponding to the polypeptide and polynucleotide sequences of the invention. The full length clones encode polypeptides which have at least 95% sequence identity to the polypeptide sequences shown in column 3.

Table 2 shows sequences with homology to polypeptide embodiments of the invention as identified by BLAST analysis against the GenBank protein (genpept) database and the PROTEOME database. Columns 1 and 2 show the polypeptide sequence identification number (Polypeptide SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for polypeptides of the invention. Column 3 shows the GenBank identification number (GenBank ID NO:) of the nearest GenBank homolog and the PROTEOME database identification numbers (PROTEOME ID NO:) of the nearest PROTEOME database homologs. Column 4 shows the probability scores for the matches between each polypeptide and its homolog(s). Column 5 shows the annotation of the GenBank and PROTEOME database homolog(s) along with relevant citations where applicable, all of which are expressly incorporated by reference herein.

Table 3 shows various structural features of the polypeptides of the invention. Columns 1 and 2 show the polypeptide sequence identification number (SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for each polypeptide of the invention. Column 3 shows the number of amino acid residues in each polypeptide. Column 4 shows potential phosphorylation sites and glycosylation sites, as determined by the MOTIFS program of the GCG sequence analysis software package (Accelrys, Burlington MA), as well as amino acid residues comprising signature sequences, domains, and motifs, including the locations of signal peptides (as indicated by "Signal Peptide" and/or "signal_cleavage"). Column 5 shows analytical methods for protein structure/function analysis and in some cases, searchable databases to which the analytical methods were applied.

Together, Tables 2 and 3 summarize the properties of polypeptides of the invention, and these properties establish that the claimed polypeptides are secreted proteins. For example, SEQ ID NO:24 is 100% identical, from residue M1 to residue R110, to human MYG1 (probable metal-dependent hydrolase) homolog (GenBank ID g10444289) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 3.7e-58, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. Data from BLAST, analysis of the PRODOM database provides further corroborative evidence that SEQ ID NO:24 is a

secreted hydrolase.

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As another example, SEQ ID NO:46 is 70% identical, from residue G45 to residue P1317 and is 39% identical, from residue W13 to residue E351, to rat MEGF6 (GenBank ID g3449294) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST 5 probability scores are both 0.0, which indicate the probabilities of obtaining the observed polypeptide sequence alignments by chance. As determined by BLAST analysis using the PROTEOME database, SEQ ID NO:46 has homology to rat MEGF6 (multiple epidermal growth factor (EGF)-like domains 6) which contains 30 epidermal growth factor-like motifs, is a putative secreted protein, and is predicted to bind calcium (PROTEOME ID: 662841 [Egfl3). SEQ ID NO:46 also has homology to a 10 human protein which contains five laminin epidermal growth factor (EGF)-like and 15 epidermal growth factor (EGF)-like domains (PROTEOME ID: 716683 MEGF11). SEQ ID NO:46 also contains EGF-like domains and laminin-type EGF-like domains as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM and SMART databases of conserved protein families/domains. (See Table 3.) Data from BLIMPS, MOTIFS, and 15 additional BLAST analyses against PRODOM and DOMO databases provide further corroborative evidence that SEQ ID NO:46 is a protein containing EGF-like domains.

SEQ ID NO:1-23, SEQ ID NO:25-45, and SEQ ID NO:47-51 were analyzed and annotated in a similar manner. The algorithms and parameters for the analysis of SEQ ID NO:1-51 are described in Table 7.

As shown in Table 4, the full length polynucleotide embodiments were assembled using cDNA sequences or coding (exon) sequences derived from genomic DNA, or any combination of these two types of sequences. Column 1 lists the polynucleotide sequence identification number (Polynucleotide SEQ ID NO:), the corresponding Incyte polynucleotide consensus sequence number (Incyte ID) for each polynucleotide of the invention, and the length of each polynucleotide sequence in basepairs. 25 Column 2 shows the nucleotide start (5') and stop (3') positions of the cDNA and/or genomic sequences used to assemble the full length polynucleotide embodiments, and of fragments of the polynucleotides which are useful, for example, in hybridization or amplification technologies that identify SEQ ID NO:52-102 or that distinguish between SEQ ID NO:52-102 and related polynucleotides.

The polynucleotide fragments described in Column 2 of Table 4 may refer specifically, for example, to Incyte cDNAs derived from tissue-specific cDNA libraries or from pooled cDNA libraries. Alternatively, the polynucleotide fragments described in column 2 may refer to GenBank

cDNAs or ESTs which contributed to the assembly of the full length polynucleotides. In addition, the polynucleotide fragments described in column 2 may identify sequences derived from the ENSEMBL (The Sanger Centre, Cambridge, UK) database (i.e., those sequences including the designation "ENST"). Alternatively, the polynucleotide fragments described in column 2 may be derived from the 5 NCBI RefSeq Nucleotide Sequence Records Database (i.e., those sequences including the designation "NM" or "NT") or the NCBI RefSeq Protein Sequence Records (i.e., those sequences including the designation "NP"). Alternatively, the polynucleotide fragments described in column 2 may refer to assemblages of both cDNA and Genscan-predicted exons brought together by an "exon stitching" algorithm. For example, a polynucleotide sequence identified as 10 FL_XXXXXX_N, N, YYYYY_N, N, represents a "stitched" sequence in which XXXXXX is the identification number of the cluster of sequences to which the algorithm was applied, and YYYYY is the number of the prediction generated by the algorithm, and $N_{1,2,3,-}$, if present, represent specific exons that may have been manually edited during analysis (See Example V). Alternatively, the polynucleotide fragments in column 2 may refer to assemblages of exons brought together by an "exon-stretching" algorithm. For example, a polynucleotide sequence identified as FLXXXXXX_gAAAAA_gBBBBB_1_N is a "stretched" sequence, with XXXXXX being the Incyte project identification number, gAAAAA being the GenBank identification number of the human genomic sequence to which the "exon-stretching" algorithm was applied, gBBBBB being the GenBank identification number or NCBI RefSeq identification number of the nearest GenBank protein homolog, and N referring to specific exons (See Example V). In instances where a RefSeq sequence was used

Alternatively, a prefix identifies component sequences that were hand-edited, predicted from genomic DNA sequences, or derived from a combination of sequence analysis methods. The

following Table lists examples of component sequence prefixes and corresponding sequence analysis methods associated with the prefixes (see Example IV and Example V).

as a protein homolog for the "exon-stretching" algorithm, a RefSeq identifier (denoted by "NM,"

"NP," or "NT") may be used in place of the GenBank identifier (i.e., gBBBBB).

Prefix	Type of analysis and/or examples of programs
GNN, GFG,	Exon prediction from genomic sequences using, for example,
ENST	GENSCAN (Stanford University, CA, USA) or FGENES
	(Computer Genomics Group, The Sanger Centre, Cambridge, UK)
GBI	Hand-edited analysis of genomic sequences.
FL	Stitched or stretched genomic sequences (see Example V).

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INCY	Full length transcript and exon prediction from mapping of EST
	sequences to the genome. Genomic location and EST composition
	data are combined to predict the exons and resulting transcript.

In some cases, Incyte cDNA coverage redundant with the sequence coverage shown in Table 4 was obtained to confirm the final consensus polynucleotide sequence, but the relevant Incyte cDNA identification numbers are not shown.

Table 5 shows the representative cDNA libraries for those full length polynucleotides which were assembled using Incyte cDNA sequences. The representative cDNA library is the Incyte cDNA library which is most frequently represented by the Incyte cDNA sequences which were used to assemble and confirm the above polynucleotides. The tissues and vectors which were used to construct the cDNA libraries shown in Table 5 are described in Table 6.

Table 8 shows single nucleotide polymorphisms (SNPs) found in polynucleotide sequences of the invention, along with allele frequencies in different human populations. Columns 1 and 2 show the polynucleotide sequence identification number (SEQ ID NO:) and the corresponding Incyte project identification number (PID) for polynucleotides of the invention. Column 3 shows the Incyte identification number for the EST in which the SNP was detected (EST ID), and column 4 shows the identification number for the SNP (SNP ID). Column 5 shows the position within the EST sequence at which the SNP is located (EST SNP), and column 6 shows the position of the SNP within the full-length polynucleotide sequence (CB1 SNP). Column 7 shows the allele found in the EST sequence. Columns 8 and 9 show the two alleles found at the SNP site. Column 10 shows the amino acid encoded by the codon including the SNP site, based upon the allele found in the EST. Columns 11-14 show the frequency of allele 1 in four different human populations. An entry of n/d (not detected) indicates that the frequency of allele 1 in the population was too low to be detected, while n/a (not available) indicates that the allele frequency was not determined for the population.

The invention also encompasses SECP variants. Various embodiments of SECP variants can have at least about 80%, at least about 90%, or at least about 95% amino acid sequence identity to the SECP amino acid sequence, and can contain at least one functional or structural characteristic of SECP.

Various embodiments also encompass polynucleotides which encode SECP. In a particular embodiment, the invention encompasses a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:52-102, which encodes SECP. The polynucleotide

sequences of SEQ ID NO:52-102, as presented in the Sequence Listing, embrace the equivalent RNA sequences, wherein occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The invention also encompasses variants of a polynucleotide encoding SECP. In particular, such a variant polynucleotide will have at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a polynucleotide encoding SECP. A particular aspect of the invention encompasses a variant of a polynucleotide comprising a sequence selected from the group consisting of SEQ ID NO:52-102 which has at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a nucleic acid 10 sequence selected from the group consisting of SEQ ID NO:52-102. Any one of the polynucleotide variants described above can encode a polypeptide which contains at least one functional or structural characteristic of SECP.

In addition, or in the alternative, a polynucleotide variant of the invention is a splice variant of a polynucleotide encoding SECP. A splice variant may have portions which have significant sequence 15 identity to a polynucleotide encoding SECP, but will generally have a greater or lesser number of polynucleotides due to additions or deletions of blocks of sequence arising from alternate splicing during mRNA processing. A splice variant may have less than about 70%, or alternatively less than about 60%, or alternatively less than about 50% polynucleotide sequence identity to a polynucleotide encoding SECP over its entire length; however, portions of the splice variant will have at least about 70%, or alternatively at least about 85%, or alternatively at least about 95%, or alternatively 100% polynucleotide sequence identity to portions of the polynucleotide encoding SECP. For example, a polynucleotide comprising a sequence of SEQ ID NO:52 and a polynucleotide comprising a sequence of SEQ ID NO:53 are splice variants of each other; a polynucleotide comprising a sequence of SEQ ID NO:54, a polynucleotide comprising a sequence of SEQ ID NO:55, a polynucleotide comprising a sequence of SEQ ID NO:56, and a polynucleotide comprising a sequence of SEQ ID NO:83 are splice variants of each other; a polynucleotide comprising a sequence of SEQ ID NO:63 and a polynucleotide comprising a sequence of SEQ ID NO:64 are splice variants of each other; a polynucleotide comprising a sequence of SEQ ID NO:66 and a polynucleotide comprising a sequence of SEQ ID NO:67 are splice variants of each other; and a polynucleotide comprising a sequence of SEQ ID NO:74 and a polynucleotide comprising a sequence of SEQ ID NO:75 are splice variants of each other; and a polynucleotide comprising a sequence of SEQ ID NO:96 and a polynucleotide comprising a sequence of SEQ ID NO:97 are splice variants of each other. Any one of the splice

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variants described above can encode a polypeptide which contains at least one functional or structural characteristic of SECP.

It will be appreciated by those skilled in the art that as a result of the degeneracy of the genetic code, a multitude of polynucleotide sequences encoding SECP, some bearing minimal similarity to the polynucleotide sequences of any known and naturally occurring gene, may be produced. Thus, the invention contemplates each and every possible variation of polynucleotide sequence that could be made by selecting combinations based on possible codon choices. These combinations are made in accordance with the standard triplet genetic code as applied to the polynucleotide sequence of naturally occurring SECP, and all such variations are to be considered as being specifically disclosed.

Although polynucleotides which encode SECP and its variants are generally capable of hybridizing to polynucleotides encoding naturally occurring SECP under appropriately selected conditions of stringency, it may be advantageous to produce polynucleotides encoding SECP or its derivatives possessing a substantially different codon usage, e.g., inclusion of non-naturally occurring codons. Codons may be selected to increase the rate at which expression of the peptide occurs in a particular prokaryotic or eukaryotic host in accordance with the frequency with which particular codons are utilized by the host. Other reasons for substantially altering the nucleotide sequence encoding SECP and its derivatives without altering the encoded amino acid sequences include the production of RNA transcripts having more desirable properties, such as a greater half-life, than transcripts produced from the naturally occurring sequence.

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The invention also encompasses production of polynucleotides which encode SECP and SECP derivatives, or fragments thereof, entirely by synthetic chemistry. After production, the synthetic polynucleotide may be inserted into any of the many available expression vectors and cell systems using reagents well known in the art. Moreover, synthetic chemistry may be used to introduce mutations into a polynucleotide encoding SECP or any fragment thereof.

Embodiments of the invention can also include polynucleotides that are capable of hybridizing to the claimed polynucleotides, and, in particular, to those having the sequences shown in SEQ ID NO:52-102 and fragments thereof, under various conditions of stringency (Wahl, G.M. and S.L. Berger (1987) Methods Enzymol. 152:399-407; Kimmel, A.R. (1987) Methods Enzymol. 152:507-511) Hybridization conditions, including annealing and wash conditions, are described in "Definitions."

Methods for DNA sequencing are well known in the art and may be used to practice any of the embodiments of the invention. The methods may employ such enzymes as the Klenow fragment of DNA polymerase I, SEQUENASE (US Biochemical, Cleveland OH), Taq polymerase (Applied

Biosystems), thermostable T7 polymerase (Amersham Biosciences, Piscataway NJ), or combinations of polymerases and proofreading exonucleases such as those found in the ELONGASE amplification system (Invitrogen, Carlsbad CA). Preferably, sequence preparation is automated with machines such as the MICROLAB 2200 liquid transfer system (Hamilton, Reno NV), PTC200 thermal cycler (MJ Research, Watertown MA) and ABI CATALYST 800 thermal cycler (Applied Biosystems). Sequencing is then carried out using either the ABI 373 or 377 DNA sequencing system (Applied Biosystems), the MEGABACE 1000 DNA sequencing system (Amersham Biosciences), or other systems known in the art. The resulting sequences are analyzed using a variety of algorithms which are well known in the art (Ausubel et al., supra, ch. 7; Meyers, R.A. (1995) Molecular Biology and Biotechnology, Wiley VCH, New York NY, pp. 856-853).

The nucleic acids encoding SECP may be extended utilizing a partial nucleotide sequence and employing various PCR-based methods known in the art to detect upstream sequences, such as promoters and regulatory elements. For example, one method which may be employed, restriction-site PCR, uses universal and nested primers to amplify unknown sequence from genomic DNA within a 15 cloning vector (Sarkar, G. (1993) PCR Methods Applic. 2:318-322). Another method, inverse PCR, uses primers that extend in divergent directions to amplify unknown sequence from a circularized template. The template is derived from restriction fragments comprising a known genomic locus and surrounding sequences (Triglia, T. et al. (1988) Nucleic Acids Res. 16:8186). A third method, capture PCR, involves PCR amplification of DNA fragments adjacent to known sequences in human and yeast artificial chromosome DNA (Lagerstrom, M. et al. (1991) PCR Methods Applic. 1:111-119). In this method, multiple restriction enzyme digestions and ligations may be used to insert an engineered double-stranded sequence into a region of unknown sequence before performing PCR. Other methods which may be used to retrieve unknown sequences are known in the art (Parker, J.D. et al. (1991) Nucleic Acids Res. 19:3055-3060). Additionally, one may use PCR, nested primers, and 25 PROMOTERFINDER libraries (Clontech, Palo Alto CA) to walk genomic DNA. This procedure avoids the need to screen libraries and is useful in finding intron/exon junctions. For all PCR-based methods, primers may be designed using commercially available software, such as OLIGO 4.06 primer analysis software (National Biosciences, Plymouth MN) or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the 30 template at temperatures of about 68°C to 72°C.

When screening for full length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. In addition, random-primed libraries, which often include

sequences containing the 5' regions of genes, are preferable for situations in which an oligo d(T) library does not yield a full-length cDNA. Genomic libraries may be useful for extension of sequence into 5' non-transcribed regulatory regions.

Capillary electrophoresis systems which are commercially available may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products. In particular, capillary sequencing may employ flowable polymers for electrophoretic separation, four different nucleotide-specific, laser-stimulated fluorescent dyes, and a charge coupled device camera for detection of the emitted wavelengths. Output/light intensity may be converted to electrical signal using appropriate software (e.g., GENOTYPER and SEQUENCE NAVIGATOR, Applied Biosystems), and the entire process from loading of samples to computer analysis and electronic data display may be computer controlled. Capillary electrophoresis is especially preferable for sequencing small DNA fragments which may be present in limited amounts in a particular sample.

In another embodiment of the invention, polynucleotides or fragments thereof which encode SECP may be cloned in recombinant DNA molecules that direct expression of SECP, or fragments or functional equivalents thereof, in appropriate host cells. Due to the inherent degeneracy of the genetic code, other polynucleotides which encode substantially the same or a functionally equivalent polypeptides may be produced and used to express SECP.

The polynucleotides of the invention can be engineered using methods generally known in the art in order to alter SECP-encoding sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, oligonucleotide-mediated site-directed mutagenesis may be used to introduce mutations that create new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, and so forth.

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The nucleotides of the present invention may be subjected to DNA shuffling techniques such as MOLECULARBREEDING (Maxygen Inc., Santa Clara CA; described in U.S. Patent No. 5,837,458; Chang, C.-C. et al. (1999) Nat. Biotechnol. 17:793-797; Christians, F.C. et al. (1999) Nat. Biotechnol. 17:259-264; and Crameri, A. et al. (1996) Nat. Biotechnol. 14:315-319) to alter or improve the biological properties of SECP, such as its biological or enzymatic activity or its ability to bind to other molecules or compounds. DNA shuffling is a process by which a library of gene variants is produced using PCR-mediated recombination of gene fragments. The library is then subjected to selection or screening procedures that identify those gene variants with the desired properties. These

preferred variants may then be pooled and further subjected to recursive rounds of DNA shuffling and selection/screening. Thus, genetic diversity is created through "artificial" breeding and rapid molecular evolution. For example, fragments of a single gene containing random point mutations may be recombined, screened, and then reshuffled until the desired properties are optimized. Alternatively, fragments of a given gene may be recombined with fragments of homologous genes in the same gene family, either from the same or different species, thereby maximizing the genetic diversity of multiple naturally occurring genes in a directed and controllable manner.

In another embodiment, polynucleotides encoding SECP may be synthesized, in whole or in part, using one or more chemical methods well known in the art (Caruthers, M.H. et al. (1980)

Nucleic Acids Symp. Ser. 7:215-223; Horn, T. et al. (1980) Nucleic Acids Symp. Ser. 7:225-232).

Alternatively, SECP itself or a fragment thereof may be synthesized using chemical methods known in the art. For example, peptide synthesis can be performed using various solution-phase or solid-phase techniques (Creighton, T. (1984) Proteins, Structures and Molecular Properties, WH Freeman, New York NY, pp. 55-60; Roberge, J.Y. et al. (1995) Science 269:202-204). Automated synthesis may be achieved using the ABI 431A peptide synthesizer (Applied Biosystems). Additionally, the amino acid sequence of SECP, or any part thereof, may be altered during direct synthesis and/or combined with sequences from other proteins, or any part thereof, to produce a variant polypeptide or a polypeptide having a sequence of a naturally occurring polypeptide.

The peptide may be substantially purified by preparative high performance liquid chromatography (Chiez, R.M. and F.Z. Regnier (1990) Methods Enzymol. 182:392-421). The composition of the synthetic peptides may be confirmed by amino acid analysis or by sequencing (Creighton, *supra*, pp. 28-53).

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In order to express a biologically active SECP, the polynucleotides encoding SECP or derivatives thereof may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for transcriptional and translational control of the inserted coding sequence in a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and inducible promoters, and 5' and 3' untranslated regions in the vector and in polynucleotides encoding SECP. Such elements may vary in their strength and specificity. Specific initiation signals may also be used to achieve more efficient translation of polynucleotides encoding SECP. Such signals include the ATG initiation codon and adjacent sequences, e.g. the Kozak sequence. In cases where a polynucleotide sequence encoding SECP and its initiation codon and upstream regulatory sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control

signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals including an in-frame ATG initiation codon should be provided by the vector. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate for the particular host cell system used (Scharf, D. et al. (1994) Results Probl. Cell Differ. 20:125-162).

Methods which are well known to those skilled in the art may be used to construct expression vectors containing polynucleotides encoding SECP and appropriate transcriptional and translational control elements. These methods include *in vitro* recombinant DNA techniques, synthetic techniques, and *in vivo* genetic recombination (Sambrook and Russell, *supra*, ch. 1-4, and 8; Ausubel et al., *supra*, ch. 1, 3, and 15).

A variety of expression vector/host systems may be utilized to contain and express polynucleotides encoding SECP. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; 15 yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (e.g., baculovirus); plant cell systems transformed with viral expression vectors (e.g., cauliflower mosaic virus, CaMV, or tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems (Sambrook and Russell, supra; Ausubel et al., supra; Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509; Engelhard, E.K. et al. 20 (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945; Takamatsu, N. (1987) EMBO J. 6:307-311; The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196; Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659; Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355). Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from 25 various bacterial plasmids, may be used for delivery of polynucleotides to the targeted organ, tissue, or cell population (Di Nicola, M. et al. (1998) Cancer Gen. Ther. 5:350-356; Yu, M. et al. (1993) Proc. Natl. Acad. Sci. USA 90:6340-6344; Buller, R.M. et al. (1985) Nature 317:813-815; McGregor, D.P. et al. (1994) Mol. Immunol. 31:219-226; Verma, I.M. and N. Somia (1997) Nature 389:239-242). The invention is not limited by the host cell employed.

In bacterial systems, a number of cloning and expression vectors may be selected depending upon the use intended for polynucleotides encoding SECP. For example, routine cloning, subcloning, and propagation of polynucleotides encoding SECP can be achieved using a multifunctional *E. coli*

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vector such as PBLUESCRIPT (Stratagene, La Jolla CA) or PSPORT1 plasmid (Invitrogen). Ligation of polynucleotides encoding SECP into the vector's multiple cloning site disrupts the lacZ gene, allowing a colorimetric screening procedure for identification of transformed bacteria containing recombinant molecules. In addition, these vectors may be useful for in vitro transcription, dideoxy 5 sequencing, single strand rescue with helper phage, and creation of nested deletions in the cloned sequence (Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509). When large quantities of SECP are needed, e.g. for the production of antibodies, vectors which direct high level expression of SECP may be used. For example, vectors containing the strong, inducible SP6 or T7 bacteriophage promoter may be used.

Yeast expression systems may be used for production of SECP. A number of vectors containing constitutive or inducible promoters, such as alpha factor, alcohol oxidase, and PGH promoters, may be used in the yeast Saccharomyces cerevisiae or Pichia pastoris. In addition, such vectors direct either the secretion or intracellular retention of expressed proteins and enable integration of foreign polynucleotide sequences into the host genome for stable propagation (Ausubel et al., 15 supra; Bitter, G.A. et al. (1987) Methods Enzymol. 153:516-544; Scorer, C.A. et al. (1994) Bio/Technology 12:181-184).

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Plant systems may also be used for expression of SECP. Transcription of polynucleotides encoding SECP may be driven by viral promoters, e.g., the 35S and 19S promoters of CaMV used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) EMBO J. 6:307-311). Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used (Coruzzi, G. et al. (1984) EMBO J. 3:1671-1680; Broglie, R. et al. (1984) Science 224:838-843; Winter, J. et al. (1991) Results Probl. Cell Differ. 17:85-105). These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection (The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196).

In mammalian cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, polynucleotides encoding SECP may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain infective virus which expresses SECP in host cells (Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659). In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells. SV40 or EBV-based vectors

may also be used for high-level protein expression.

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Human artificial chromosomes (HACs) may also be employed to deliver larger fragments of DNA than can be contained in and expressed from a plasmid. HACs of about 6 kb to 10 Mb are constructed and delivered via conventional delivery methods (liposomes, polycationic amino polymers, or vesicles) for therapeutic purposes (Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355).

For long term production of recombinant proteins in mammalian systems, stable expression of SECP in cell lines is preferred. For example, polynucleotides encoding SECP can be transformed into cell lines using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for about 1 to 2 days in enriched media before being switched to selective media. The purpose of the selectable marker is to confer resistance to a selective agent, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be propagated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase and adenine phosphoribosyltransferase genes, for use in the and apr cells, respectively (Wigler, M. et al. (1977) Cell 11:223-232; Lowy, I. et al. (1980) Cell 22:817-823). Also, antimetabolite, antibiotic, or herbicide resistance can be used as the basis for selection. For example, dhfr confers resistance to methotrexate; neo confers resistance to the aminoglycosides neomycin and G-418; and als and pat confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively (Wigler, M. et al. (1980) Proc. Natl. Acad. Sci. USA 77:3567-3570; Colbere-Garapin, F. et al. (1981) J. Mol. Biol. 150:1-14). Additional selectable genes have been described, e.g., trpB and hisD, which alter cellular requirements for metabolites (Hartman, S.C. and R.C. Mulligan (1988) Proc. Natl. Acad. Sci. USA 85:8047-8051). Visible markers, e.g., anthocyanins, green fluorescent proteins (GFP; Clontech), β-glucuronidase and its substrate β-glucuronide, or luciferase and its substrate luciferin may be used. These markers can be used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system (Rhodes, C.A. (1995) Methods Mol. Biol. 55:121-131).

Although the presence/absence of marker gene expression suggests that the gene of interest is also present, the presence and expression of the gene may need to be confirmed. For example, if the sequence encoding SECP is inserted within a marker gene sequence, transformed cells containing

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polynucleotides encoding SECP can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a sequence encoding SECP under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

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In general, host cells that contain the polynucleotide encoding SECP and that express SECP may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein sequences.

Immunological methods for detecting and measuring the expression of SECP using either specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on SECP is preferred, but a competitive 15 binding assay may be employed. These and other assays are well known in the art (Hampton, R. et al. (1990) Serological Methods, a Laboratory Manual, APS Press, St. Paul MN, Sect. IV; Coligan, J.E. et al. (1997) Current Protocols in Immunology, Greene Pub. Associates and Wiley-Interscience, New York NY; Pound, J.D. (1998) Immunochemical Protocols, Humana Press, Totowa NJ).

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides encoding SECP include oligolabeling, nick translation, end-labeling, or PCR amplification using a labeled nucleotide. Alternatively, polynucleotides encoding SECP, or any fragments thereof, may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits, such as those provided by Amersham Biosciences, Promega (Madison WI), and US Biochemical. Suitable reporter molecules or labels which may be used for ease of detection include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents, as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with polynucleotides encoding SECP may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a

transformed cell may be secreted or retained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides which encode SECP may be designed to contain signal sequences which direct secretion of SECP through a prokaryotic or eukaryotic cell membrane.

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In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted polynucleotides or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" or "pro" form of the protein may also be used to specify protein targeting, folding, and/or activity. 10 Different host cells which have specific cellular machinery and characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38) are available from the American Type Culture Collection (ATCC, Manassas VA) and may be chosen to ensure the correct modification and processing of the foreign protein.

In another embodiment of the invention, natural, modified, or recombinant polynucleotides encoding SECP may be ligated to a heterologous sequence resulting in translation of a fusion protein in any of the aforementioned host systems. For example, a chimeric SECP protein containing a heterologous moiety that can be recognized by a commercially available antibody may facilitate the screening of peptide libraries for inhibitors of SECP activity. Heterologous protein and peptide moieties may also facilitate purification of fusion proteins using commercially available affinity matrices. Such moieties include, but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, FLAG, c-myc, and hemagglutinin (HA). GST, MBP, Trx, CBP, and 6-His enable purification of their cognate fusion proteins on immobilized glutathione, maltose, phenylarsine oxide, calmodulin, and metal-chelate resins, respectively. FLAG, c-myc, and hemagglutinin (HA) enable immunoaffinity purification of fusion proteins using commercially available monoclonal and polyclonal antibodies that specifically recognize these epitope tags. A fusion protein may also be engineered to contain a proteolytic cleavage site located between the SECP encoding sequence and the heterologous protein sequence, so that SECP may be cleaved away from the heterologous moiety following purification. Methods for fusion protein expression and purification are discussed in Ausubel et al. (supra, ch. 10 and 16). A variety of commercially available kits may also be used to facilitate expression and purification of fusion proteins.

In another embodiment, synthesis of radiolabeled SECP may be achieved in vitro using the TNT rabbit reticulocyte lysate or wheat germ extract system (Promega). These systems couple

transcription and translation of protein-coding sequences operably associated with the T7, T3, or SP6 promoters. Translation takes place in the presence of a radiolabeled amino acid precursor, for example, ³⁵S-methionine.

SECP, fragments of SECP, or variants of SECP may be used to screen for compounds that specifically bind to SECP. One or more test compounds may be screened for specific binding to SECP. In various embodiments, 1, 2, 3, 4, 5, 10, 20, 50, 100, or 200 test compounds can be screened for specific binding to SECP. Examples of test compounds can include antibodies, anticalins, oligonucleotides, proteins (e.g., ligands or receptors), or small molecules.

In related embodiments, variants of SECP can be used to screen for binding of test compounds, such as antibodies, to SECP, a variant of SECP, or a combination of SECP and/or one or more variants SECP. In an embodiment, a variant of SECP can be used to screen for compounds that bind to a variant of SECP, but not to SECP having the exact sequence of a sequence of SEQ ID NO:1-51. SECP variants used to perform such screening can have a range of about 50% to about 99% sequence identity to SECP, with various embodiments having 60%, 70%, 75%, 80%, 85%, 90%, and 95% sequence identity.

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In an embodiment, a compound identified in a screen for specific binding to SECP can be closely related to the natural ligand of SECP, e.g., a ligand or fragment thereof, a natural substrate, a structural or functional mimetic, or a natural binding partner (Coligan, J.E. et al. (1991) <u>Current Protocols in Immunology</u> 1(2):Chapter 5). In another embodiment, the compound thus identified can be a natural ligand of a receptor SECP (Howard, A.D. et al. (2001) Trends Pharmacol. Sci.22:132-140; Wise, A. et al. (2002) Drug Discovery Today 7:235-246).

In other embodiments, a compound identified in a screen for specific binding to SECP can be closely related to the natural receptor to which SECP binds, at least a fragment of the receptor, or a fragment of the receptor including all or a portion of the ligand binding site or binding pocket. For example, the compound may be a receptor for SECP which is capable of propagating a signal, or a decoy receptor for SECP which is not capable of propagating a signal (Ashkenazi, A. and V.M. Divit (1999) Curr. Opin. Cell Biol. 11:255-260; Mantovani, A. et al. (2001) Trends Immunol. 22:328-336). The compound can be rationally designed using known techniques. Examples of such techniques include those used to construct the compound etanercept (ENBREL; Amgen Inc., Thousand Oaks CA), which is efficacious for treating rheumatoid arthritis in humans. Etanercept is an engineered p75 tumor necrosis factor (TNF) receptor dimer linked to the Fc portion of human IgG₁ (Taylor, P.C. et al. (2001) Curr. Opin. Immunol. 13:611-616).

In one embodiment, two or more antibodies having similar or, alternatively, different specificities can be screened for specific binding to SECP, fragments of SECP, or variants of SECP. The binding specificity of the antibodies thus screened can thereby be selected to identify particular fragments or variants of SECP. In one embodiment, an antibody can be selected such that its binding specificity allows for preferential identification of specific fragments or variants of SECP. In another embodiment, an antibody can be selected such that its binding specificity allows for preferential diagnosis of a specific disease or condition having increased, decreased, or otherwise abnormal production of SECP.

In an embodiment, anticalins can be screened for specific binding to SECP, fragments of

SECP, or variants of SECP. Anticalins are ligand-binding proteins that have been constructed based
on a lipocalin scaffold (Weiss, G.A. and H.B. Lowman (2000) Chem. Biol. 7:R177-R184; Skerra, A.

(2001) J. Biotechnol. 74:257-275). The protein architecture of lipocalins can include a beta-barrel
having eight antiparallel beta-strands, which supports four loops at its open end. These loops form the
natural ligand-binding site of the lipocalins, a site which can be re-engineered in vitro by amino acid
substitutions to impart novel binding specificities. The amino acid substitutions can be made using
methods known in the art or described herein, and can include conservative substitutions (e.g.,
substitutions that do not alter binding specificity) or substitutions that modestly, moderately, or
significantly alter binding specificity.

In one embodiment, screening for compounds which specifically bind to, stimulate, or inhibit SECP involves producing appropriate cells which express SECP, either as a secreted protein or on the cell membrane. Preferred cells can include cells from mammals, yeast, *Drosophila*, or *E. coli*. Cells expressing SECP or cell membrane fractions which contain SECP are then contacted with a test compound and binding, stimulation, or inhibition of activity of either SECP or the compound is analyzed.

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An assay may simply test binding of a test compound to the polypeptide, wherein binding is detected by a fluorophore, radioisotope, enzyme conjugate, or other detectable label. For example, the assay may comprise the steps of combining at least one test compound with SECP, either in solution or affixed to a solid support, and detecting the binding of SECP to the compound. Alternatively, the assay may detect or measure binding of a test compound in the presence of a labeled competitor. Additionally, the assay may be carried out using cell-free preparations, chemical libraries, or natural product mixtures, and the test compound(s) may be free in solution or affixed to a solid support.

An assay can be used to assess the ability of a compound to bind to its natural ligand and/or to

inhibit the binding of its natural ligand to its natural receptors. Examples of such assays include radiolabeling assays such as those described in U.S. Patent No. 5,914,236 and U.S. Patent No. 6,372,724. In a related embodiment, one or more amino acid substitutions can be introduced into a polypeptide compound (such as a receptor) to improve or alter its ability to bind to its natural ligands (Matthews, 5 D.J. and J.A. Wells. (1994) Chem. Biol. 1:25-30). In another related embodiment, one or more amino acid substitutions can be introduced into a polypeptide compound (such as a ligand) to improve or alter its ability to bind to its natural receptors (Cunningham, B.C. and J.A. Wells (1991) Proc. Natl. Acad. Sci. USA 88:3407-3411; Lowman, H.B. et al. (1991) J. Biol. Chem. 266:10982-10988).

SECP, fragments of SECP, or variants of SECP may be used to screen for compounds that modulate the activity of SECP. Such compounds may include agonists, antagonists, or partial or inverse agonists. In one embodiment, an assay is performed under conditions permissive for SECP activity, wherein SECP is combined with at least one test compound, and the activity of SECP in the presence of a test compound is compared with the activity of SECP in the absence of the test compound. A change in the activity of SECP in the presence of the test compound is indicative of a 15 compound that modulates the activity of SECP. Alternatively, a test compound is combined with an in vitro or cell-free system comprising SECP under conditions suitable for SECP activity, and the assay is performed. In either of these assays, a test compound which modulates the activity of SECP may do so indirectly and need not come in direct contact with the test compound. At least one and up to a plurality of test compounds may be screened.

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In another embodiment, polynucleotides encoding SECP or their mammalian homologs may be "knocked out" in an animal model system using homologous recombination in embryonic stem (ES) cells. Such techniques are well known in the art and are useful for the generation of animal models of human disease (see, e.g., U.S. Patent No. 5,175,383 and U.S. Patent No. 5,767,337). For example, mouse ES cells, such as the mouse 129/SvJ cell line, are derived from the early mouse embryo and 25 grown in culture. The ES cells are transformed with a vector containing the gene of interest disrupted by a marker gene, e.g., the neomycin phosphotransferase gene (neo; Capecchi, M.R. (1989) Science 244:1288-1292). The vector integrates into the corresponding region of the host genome by homologous recombination. Alternatively, homologous recombination takes place using the Cre-loxP system to knockout a gene of interest in a tissue- or developmental stage-specific manner (Marth, J.D. 30 (1996) Clin. Invest. 97:1999-2002; Wagner, K.U. et al. (1997) Nucleic Acids Res. 25:4323-4330). Transformed ES cells are identified and microinjected into mouse cell blastocysts such as those from the C57BL/6 mouse strain. The blastocysts are surgically transferred to pseudopregnant dams, and

the resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous strains. Transgenic animals thus generated may be tested with potential therapeutic or toxic agents.

Polynucleotides encoding SECP may also be manipulated *in vitro* in ES cells derived from human blastocysts. Human ES cells have the potential to differentiate into at least eight separate cell lineages including endoderm, mesoderm, and ectodermal cell types. These cell lineages differentiate into, for example, neural cells, hematopoietic lineages, and cardiomyocytes (Thomson, J.A. et al. (1998) Science 282:1145-1147).

Polynucleotides encoding SECP can also be used to create "knockin" humanized animals (pigs) or transgenic animals (mice or rats) to model human disease. With knockin technology, a region of a polynucleotide encoding SECP is injected into animal ES cells, and the injected sequence integrates into the animal cell genome. Transformed cells are injected into blastulae, and the blastulae are implanted as described above. Transgenic progeny or inbred lines are studied and treated with potential pharmaceutical agents to obtain information on treatment of a human disease. Alternatively, a mammal inbred to overexpress SECP, e.g., by secreting SECP in its milk, may also serve as a convenient source of that protein (Janne, J. et al. (1998) Biotechnol. Annu. Rev. 4:55-74).

THERAPEUTICS

Chemical and structural similarity, e.g., in the context of sequences and motifs, exists between regions of SECP and secreted proteins. In addition, examples of tissues expressing SECP promonocyte line treated with PMA, peripheral blood mononuclear cells treated with interleukin 10, breast carcinoma and primary mammary epithelial cells, and normal and tumorous colon tissues, cancerous and normal breast, colon, lung, ovarian, prostate tissues, adipocytes, peripheral blood mononuclear cells, dendritic cells, monocytes, C3A hepatoma cell line, Raji B lymphoblast cells, and human umbilical vein endothelial cells. Further examples of tissues expressing SECP can be found in Table 6 and can also be found in Example XI. Therefore, SECP appears to play a role in cell proliferative, autoimmune/inflammatory, cardiovascular, neurological, and developmental disorders. In the treatment of disorders associated with increased SECP expression or activity, it is desirable to decrease the expression or activity of SECP. In the treatment of disorders associated with decreased SECP expression or activity, it is desirable to increase the expression or activity of SECP.

Therefore, in one embodiment, SECP or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of SECP. Examples of such disorders include, but are not limited to, a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease

(MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, a cancer of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, colon, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, hung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus; an autoimmune/inflammatory disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis. 10 cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, 15 Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; a cardiovascular disorder such as congestive heart failure, ischemic heart disease, angina pectoris, myocardial infarction, hypertensive heart disease, degenerative valvular heart disease, calcific aortic valve stenosis, congenitally bicuspid aortic valve, mitral annular calcification, mitral valve prolapse, rheumatic fever and rheumatic heart disease, infective endocarditis, nonbacterial thrombotic endocarditis, endocarditis of systemic hupus erythematosus, carcinoid heart disease, cardiomyopathy, myocarditis, pericarditis, neoplastic heart disease, congenital heart disease, complications of cardiac transplantation, arteriovenous fistula, atherosclerosis, hypertension, vasculitis, Raynaud's disease, aneurysms, arterial dissections, varicose veins, thrombophlebitis and phlebothrombosis, vascular tumors, and complications of thrombolysis, balloon angioplasty, vascular replacement, and coronary artery bypass graft surgery; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural

abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal 5 syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia gravis, periodic paralysis, mental disorders including 10 mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; and a developmental disorder such as renal tubular acidosis, anemia, Cushing's syndrome, achondroplastic dwarfism, Duchenne and Becker muscular dystrophy, epilepsy, 15 gonadal dysgenesis, WAGR syndrome (Wilms' tumor, aniridia, genitourinary abnormalities, and mental retardation), Smith-Magenis syndrome, myelodysplastic syndrome, hereditary mucoepithelial dysplasia, hereditary keratodermas, hereditary neuropathies such as Charcot-Marie-Tooth disease and neurofibromatosis, hypothyroidism, hydrocephalus, seizure disorders such as Syndenham's chorea and cerebral palsy, spina bifida, anencephaly, craniorachischisis, congenital glaucoma, cataract, and sensorineural hearing loss.

In another embodiment, a vector capable of expressing SECP or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of SECP including, but not limited to, those described above.

In a further embodiment, a composition comprising a substantially purified SECP in conjunction with a suitable pharmaceutical carrier may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of SECP including, but not limited to, those provided above.

In still another embodiment, an agonist which modulates the activity of SECP may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of SECP including, but not limited to, those listed above.

In a further embodiment, an antagonist of SECP may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of SECP. Examples of such

disorders include, but are not limited to, those cell proliferative, autoimmune/inflammatory, cardiovascular, neurological, and developmental disorders described above. In one aspect, an antibody which specifically binds SECP may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissues which express SECP.

In an additional embodiment, a vector expressing the complement of the polynucleotide encoding SECP may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of SECP including, but not limited to, those described above.

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In other embodiments, any protein, agonist, antagonist, antibody, complementary sequence, or vector embodiments may be administered in combination with other appropriate therapeutic agents.

Selection of the appropriate agents for use in combination therapy may be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents may act synergistically to effect the treatment or prevention of the various disorders described above. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects.

An antagonist of SECP may be produced using methods which are generally known in the art. In particular, purified SECP may be used to produce antibodies or to screen libraries of pharmaceutical agents to identify those which specifically bind SECP. Antibodies to SECP may also be generated using methods that are well known in the art. Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments, and fragments produced by a Fab expression library. In an embodiment, neutralizing antibodies (i.e., those which inhibit dimer formation) can be used therapeutically. Single chain antibodies (e.g., from camels or llamas) may be potent enzyme inhibitors and may have application in the design of peptide mimetics, and in the development of immuno-adsorbents and biosensors (Muyldermans, S. (2001) J. Biotechnol. 74:277-302).

For the production of antibodies, various hosts including goats, rabbits, rats, mice, camels, dromedaries, llamas, humans, and others may be immunized by injection with SECP or with any fragment or oligopeptide thereof which has immunogenic properties. Depending on the host species, various adjuvants may be used to increase immunological response. Such adjuvants include, but are not limited to, Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, KLH, and dinitrophenol. Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and *Corynebacterium parvum* are especially preferable.

It is preferred that the oligopeptides, peptides, or fragments used to induce antibodies to SECP have an amino acid sequence consisting of at least about 5 amino acids, and generally will consist of at least about 10 amino acids. It is also preferable that these oligopeptides, peptides, or fragments are substantially identical to a portion of the amino acid sequence of the natural protein. Short stretches of SECP amino acids may be fused with those of another protein, such as KLH, and antibodies to the chimeric molecule may be produced.

Monoclonal antibodies to SECP may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique (Kohler, G. et al. (1975) Nature 256:495-497; Kozbor, D. et al. (1985) J. Immunol. Methods 81:31-42; Cote, R.J. et al. (1983) Proc. Natl. Acad. Sci. USA 80:2026-2030; Cole, S.P. et al. (1984) Mol. Cell Biol. 62:109-120).

In addition, techniques developed for the production of "chimeric antibodies," such as the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used (Morrison, S.L. et al. (1984) Proc. Natl. Acad. Sci. USA 81:6851-6855; Neuberger, M.S. et al. (1984) Nature 312:604-608; Takeda, S. et al. (1985) Nature 314:452-454). Alternatively, techniques described for the production of single chain antibodies may be adapted, using methods known in the art, to produce SECP-specific single chain antibodies. Antibodies with related specificity, but of distinct idiotypic composition, may be generated by chain shuffling from random combinatorial immunoglobulin libraries (Burton, D.R. (1991) Proc. Natl. Acad. Sci. USA 88:10134-10137).

Antibodies may also be produced by inducing *in vivo* production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature (Orlandi, R. et al. (1989) Proc. Natl. Acad. Sci. USA 86:3833-3837; Winter, G. et al. (1991) Nature 349:293-299).

Antibody fragments which contain specific binding sites for SECP may also be generated. For example, such fragments include, but are not limited to, F(ab')₂ fragments produced by pepsin digestion of the antibody molecule and Fab fragments generated by reducing the disulfide bridges of the F(ab')2 fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity (Huse, W.D. et al. (1989) Science 246:1275-1281).

Various immunoassays may be used for screening to identify antibodies having the desired

specificity. Numerous protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art. Such immunoassays typically involve the measurement of complex formation between SECP and its specific antibody. A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering SECP epitopes is generally used, but a competitive binding assay may also be employed (Pound, *supra*).

Various methods such as Scatchard analysis in conjunction with radioimmunoassay techniques may be used to assess the affinity of antibodies for SECP. Affinity is expressed as an association constant, K_a, which is defined as the molar concentration of SECP-antibody complex divided by the molar concentrations of free antigen and free antibody under equilibrium conditions. The K_a determined for a preparation of polyclonal antibodies, which are heterogeneous in their affinities for multiple SECP epitopes, represents the average affinity, or avidity, of the antibodies for SECP. The K_a determined for a preparation of monoclonal antibodies, which are monospecific for a particular SECP epitope, represents a true measure of affinity. High-affinity antibody preparations with K_a ranging from about 10° to 10¹² L/mole are preferred for use in immunoassays in which the SECP-antibody complex must withstand rigorous manipulations. Low-affinity antibody preparations with K_a ranging from about 10° to 10¹ L/mole are preferred for use in immunopurification and similar procedures which ultimately require dissociation of SECP, preferably in active form, from the antibody (Catty, D. (1988) Antibodies, Volume I: A Practical Approach, IRL Press, Washington DC; Liddell, J.E. and A. Cryer (1991) A Practical Guide to Monoclonal Antibodies, John Wiley & Sons, New York NY).

The titer and avidity of polyclonal antibody preparations may be further evaluated to determine the quality and suitability of such preparations for certain downstream applications. For example, a polyclonal antibody preparation containing at least 1-2 mg specific antibody/ml, preferably 5-10 mg specific antibody/ml, is generally employed in procedures requiring precipitation of SECP-antibody complexes. Procedures for evaluating antibody specificity, titer, and avidity, and guidelines for antibody quality and usage in various applications, are generally available (Catty, supra; Coligan et al., supra).

In another embodiment of the invention, polynucleotides encoding SECP, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, modifications of gene expression can be achieved by designing complementary sequences or antisense molecules (DNA, RNA, PNA, or modified oligonucleotides) to the coding or regulatory regions of the gene encoding

SECP. Such technology is well known in the art, and antisense oligonucleotides or larger fragments can be designed from various locations along the coding or control regions of sequences encoding SECP (Agrawal, S., ed. (1996) Antisense Therapeutics, Humana Press, Totawa NJ).

In therapeutic use, any gene delivery system suitable for introduction of the antisense sequences into appropriate target cells can be used. Antisense sequences can be delivered intracellularly in the form of an expression plasmid which, upon transcription, produces a sequence complementary to at least a portion of the cellular sequence encoding the target protein (Slater, J.E. et al. (1998) J. Allergy Clin. Immunol. 102:469-475; Scanlon, K.J. et al. (1995) 9:1288-1296). Antisense sequences can also be introduced intracellularly through the use of viral vectors, such as retrovirus and adeno-associated virus vectors (Miller, A.D. (1990) Blood 76:271; Ausubel et al., supra; Uckert, W. and W. Walther (1994) Pharmacol. Ther. 63:323-347). Other gene delivery mechanisms include liposome-derived systems, artificial viral envelopes, and other systems known in the art (Rossi, J.J. (1995) Br. Med. Bull. 51:217-225; Boado, R.J. et al. (1998) J. Pharm. Sci. 87:1308-1315; Morris, M.C. et al. (1997) Nucleic Acids Res. 25:2730-2736).

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In another embodiment of the invention, polynucleotides encoding SECP may be used for somatic or germline gene therapy. Gene therapy may be performed to (i) correct a genetic deficiency (e.g., in the cases of severe combined immunodeficiency (SCID)-X1 disease characterized by Xlinked inheritance (Cavazzana-Calvo, M. et al. (2000) Science 288:669-672), severe combined immunodeficiency syndrome associated with an inherited adenosine deaminase (ADA) deficiency (Blaese, R.M. et al. (1995) Science 270:475-480; Bordignon, C. et al. (1995) Science 270:470-475), cystic fibrosis (Zabner, J. et al. (1993) Cell 75:207-216; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:643-666; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:667-703), thalassamias, familial hypercholesterolemia, and hemophilia resulting from Factor VIII or Factor IX deficiencies (Crystal, R.G. (1995) Science 270:404-410; Verma, I.M. and N. Somia (1997) Nature 389:239-242)), (ii) 25 express a conditionally lethal gene product (e.g., in the case of cancers which result from unregulated cell proliferation), or (iii) express a protein which affords protection against intracellular parasites (e.g., against human retroviruses, such as human immunodeficiency virus (HIV) (Baltimore, D. (1988) Nature 335:395-396; Poeschla, E. et al. (1996) Proc. Natl. Acad. Sci. USA 93:11395-11399), hepatitis B or C virus (HBV, HCV); fungal parasites, such as Candida albicans and Paracoccidioides brasiliensis; and protozoan parasites such as Plasmodium falciparum and Trypanosoma cruzi). In the case where a genetic deficiency in SECP expression or regulation causes disease, the expression of SECP from an appropriate population of transduced cells may alleviate the clinical manifestations

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caused by the genetic deficiency.

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In a further embodiment of the invention, diseases or disorders caused by deficiencies in SECP are treated by constructing mammalian expression vectors encoding SECP and introducing these vectors by mechanical means into SECP-deficient cells. Mechanical transfer technologies for 5 use with cells in vivo or ex vitro include (i) direct DNA microinjection into individual cells, (ii) ballistic gold particle delivery, (iii) liposome-mediated transfection, (iv) receptor-mediated gene transfer, and (v) the use of DNA transposons (Morgan, R.A. and W.F. Anderson (1993) Annu. Rev. Biochem. 62:191-217; Ivics, Z. (1997) Cell 91:501-510; Boulay, J.-L. and H. Récipon (1998) Curr. Opin. Biotechnol. 9:445-450).

Expression vectors that may be effective for the expression of SECP include, but are not limited to, the PCDNA 3.1, EPITAG, PRCCMV2, PREP, PVAX, PCR2-TOPOTA vectors (Invitrogen, Carlsbad CA), PCMV-SCRIPT, PCMV-TAG, PEGSH/PERV (Stratagene, La Jolla CA), and PTET-OFF, PTET-ON, PTRE2, PTRE2-LUC, PTK-HYG (Clontech, Palo Alto CA). SECP may be expressed using (i) a constitutively active promoter, (e.g., from cytomegalovirus (CMV), Rous 15 sarcoma virus (RSV), SV40 virus, thymidine kinase (TK), or β-actin genes), (ii) an inducible promoter (e.g., the tetracycline-regulated promoter (Gossen, M. and H. Bujard (1992) Proc. Natl. Acad. Sci. USA 89:5547-5551; Gossen, M. et al. (1995) Science 268:1766-1769; Rossi, F.M.V. and H.M. Blau (1998) Curr. Opin. Biotechnol. 9:451-456), commercially available in the T-REX plasmid (Invitrogen)); the ecdysone-inducible promoter (available in the plasmids PVGRXR and PIND; Invitrogen); the 20 FK506/rapamycin inducible promoter; or the RU486/mifepristone inducible promoter (Rossi, F.M.V. and H.M. Blau, supra)), or (iii) a tissue-specific promoter or the native promoter of the endogenous gene encoding SECP from a normal individual.

Commercially available liposome transformation kits (e.g., the PERFECT LIPID TRANSFECTION KIT, available from Invitrogen) allow one with ordinary skill in the art to deliver 25 polynucleotides to target cells in culture and require minimal effort to optimize experimental parameters. In the alternative, transformation is performed using the calcium phosphate method (Graham, F.L. and A.J. Eb (1973) Virology 52:456-467), or by electroporation (Neumann, E. et al. (1982) EMBO J. 1:841-845). The introduction of DNA to primary cells requires modification of these standardized mammalian transfection protocols.

In another embodiment of the invention, diseases or disorders caused by genetic defects with respect to SECP expression are treated by constructing a retrovirus vector consisting of (i) the polynucleotide encoding SECP under the control of an independent promoter or the retrovirus long

terminal repeat (LTR) promoter, (ii) appropriate RNA packaging signals, and (iii) a Rev-responsive element (RRE) along with additional retrovirus cis-acting RNA sequences and coding sequences required for efficient vector propagation. Retrovirus vectors (e.g., PFB and PFBNEO) are commercially available (Stratagene) and are based on published data (Riviere, I. et al. (1995) Proc. 5 Natl. Acad. Sci. USA 92:6733-6737), incorporated by reference herein. The vector is propagated in an appropriate vector producing cell line (VPCL) that expresses an envelope gene with a tropism for receptors on the target cells or a promiscuous envelope protein such as VSVg (Armentano, D. et al. (1987) J. Virol. 61:1647-1650; Bender, M.A. et al. (1987) J. Virol. 61:1639-1646; Adam, M.A. and A.D. Miller (1988) J. Virol. 62:3802-3806; Dull, T. et al. (1998) J. Virol. 72:8463-8471; Zufferey, R. et 10 al. (1998) J. Virol. 72:9873-9880). U.S. Patent No. 5,910,434 to Rigg ("Method for obtaining retrovirus packaging cell lines producing high transducing efficiency retroviral supernatant") discloses a method for obtaining retrovirus packaging cell lines and is hereby incorporated by reference. Propagation of retrovirus vectors, transduction of a population of cells (e.g., CD4+ T-cells), and the return of transduced cells to a patient are procedures well known to persons skilled in the art of gene therapy and have been well documented (Ranga, U. et al. (1997) J. Virol. 71:7020-7029; Bauer, G. et al. (1997) Blood 89:2259-2267; Bonyhadi, M.L. (1997) J. Virol. 71:4707-4716; Ranga, U. et al. (1998) Proc. Natl. Acad. Sci. USA 95:1201-1206; Su, L. (1997) Blood 89:2283-2290).

In an embodiment, an adenovirus-based gene therapy delivery system is used to deliver polynucleotides encoding SECP to cells which have one or more genetic abnormalities with respect to the expression of SECP. The construction and packaging of adenovirus-based vectors are well known to those with ordinary skill in the art. Replication defective adenovirus vectors have proven to be versatile for importing genes encoding immunoregulatory proteins into intact islets in the pancreas (Csete, M.E. et al. (1995) Transplantation 27:263-268). Potentially useful adenoviral vectors are described in U.S. Patent No. 5,707,618 to Armentano ("Adenovirus vectors for gene therapy"), hereby incorporated by reference. For adenoviral vectors, see also Antinozzi, P.A. et al. (1999; Annu. Rev. Nutr. 19:511-544) and Verma, I.M. and N. Somia (1997; Nature 18:389:239-242).

In another embodiment, a herpes-based, gene therapy delivery system is used to deliver polynucleotides encoding SECP to target cells which have one or more genetic abnormalities with respect to the expression of SECP. The use of herpes simplex virus (HSV)-based vectors may be especially valuable for introducing SECP to cells of the central nervous system, for which HSV has a tropism. The construction and packaging of herpes-based vectors are well known to those with ordinary skill in the art. A replication-competent herpes simplex virus (HSV) type 1-based vector has

been used to deliver a reporter gene to the eyes of primates (Liu, X. et al. (1999) Exp. Eye Res. 169:385-395). The construction of a HSV-1 virus vector has also been disclosed in detail in U.S. Patent No. 5,804,413 to DeLuca ("Herpes simplex virus strains for gene transfer"), which is hereby incorporated by reference. U.S. Patent No. 5,804,413 teaches the use of recombinant HSV d92 which consists of a genome containing at least one exogenous gene to be transferred to a cell under the control of the appropriate promoter for purposes including human gene therapy. Also taught by this patent are the construction and use of recombinant HSV strains deleted for ICP4, ICP27 and ICP22. For HSV vectors, see also Goins, W.F. et al. (1999; J. Virol. 73:519-532) and Xu, H. et al. (1994; Dev. Biol. 163:152-161). The manipulation of cloned herpesvirus sequences, the generation of recombinant virus following the transfection of multiple plasmids containing different segments of the large herpesvirus genomes, the growth and propagation of herpesvirus, and the infection of cells with herpesvirus are techniques well known to those of ordinary skill in the art.

In another embodiment, an alphavirus (positive, single-stranded RNA virus) vector is used to deliver polynucleotides encoding SECP to target cells. The biology of the prototypic alphavirus, Semliki Forest Virus (SFV), has been studied extensively and gene transfer vectors have been based on the SFV genome (Garoff, H. and K.-J. Li (1998) Curr. Opin. Biotechnol. 9:464-469). During alphavirus RNA replication, a subgenomic RNA is generated that normally encodes the viral capsid proteins. This subgenomic RNA replicates to higher levels than the full length genomic RNA, resulting in the overproduction of capsid proteins relative to the viral proteins with enzymatic activity (e.g., protease and polymerase). Similarly, inserting the coding sequence for SECP into the alphavirus genome in place of the capsid-coding region results in the production of a large number of SECPcoding RNAs and the synthesis of high levels of SECP in vector transduced cells. While alphavirus infection is typically associated with cell lysis within a few days, the ability to establish a persistent infection in hamster normal kidney cells (BHK-21) with a variant of Sindbis virus (SIN) indicates that the lytic replication of alphaviruses can be altered to suit the needs of the gene therapy application (Dryga, S.A. et al. (1997) Virology 228:74-83). The wide host range of alphaviruses will allow the introduction of SECP into a variety of cell types. The specific transduction of a subset of cells in a population may require the sorting of cells prior to transduction. The methods of manipulating infectious cDNA clones of alphaviruses, performing alphavirus cDNA and RNA transfections, and performing alphavirus infections, are well known to those with ordinary skill in the art.

Oligonucleotides derived from the transcription initiation site, e.g., between about positions -10 and +10 from the start site, may also be employed to inhibit gene expression. Similarly, inhibition can

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be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances using triplex DNA have been described in the literature (Gee, J.E. et al. (1994) in Huber, B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177). A complementary sequence or antisense molecule may also be designed to block translation of mRNA by preventing the transcript from binding to ribosomes.

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. For example, engineered hammerhead motif ribozyme molecules may specifically and efficiently catalyze endonucleolytic cleavage of RNA molecules encoding SECP.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the target molecule for ribozyme cleavage sites, including the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides, corresponding to the region of the target gene containing the cleavage site, may be evaluated for secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays.

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Complementary ribonucleic acid molecules and ribozymes may be prepared by any method known in the art for the synthesis of nucleic acid molecules. These include techniques for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by *in vitro* and *in vivo* transcription of DNA molecules encoding SECP. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA polymerase promoters such as T7 or SP6. Alternatively, these cDNA constructs that synthesize complementary RNA, constitutively or inducibly, can be introduced into cell lines, cells, or tissues.

RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends of the molecule, or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. This concept is inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine,

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guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

In other embodiments of the invention, the expression of one or more selected polynucleotides of the present invention can be altered, inhibited, decreased, or silenced using RNA interference (RNAi) or post-transcriptional gene silencing (PTGS) methods known in the art. RNAi is a post-5 transcriptional mode of gene silencing in which double-stranded RNA (dsRNA) introduced into a targeted cell specifically suppresses the expression of the homologous gene (i.e., the gene bearing the sequence complementary to the dsRNA). This effectively knocks out or substantially reduces the expression of the targeted gene. PTGS can also be accomplished by use of DNA or DNA fragments as well. RNAi methods are described by Fire, A. et al. (1998; Nature 391:806-811) and Gura, T. 10 (2000; Nature 404:804-808). PTGS can also be initiated by introduction of a complementary segment of DNA into the selected tissue using gene delivery and/or viral vector delivery methods described herein or known in the art.

RNAi can be induced in mammalian cells by the use of small interfering RNA also known as siRNA. SiRNA are shorter segments of dsRNA (typically about 21 to 23 nucleotides in length) that 15 result in vivo from cleavage of introduced dsRNA by the action of an endogenous ribonuclease. SiRNA appear to be the mediators of the RNAi effect in mammals. The most effective siRNAs appear to be 21 nucleotide dsRNAs with 2 nucleotide 3' overhangs. The use of siRNA for inducing RNAi in mammalian cells is described by Elbashir, S.M. et al. (2001; Nature 411:494-498).

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SiRNA can either be generated indirectly by introduction of dsRNA into the targeted cell, or directly by mammalian transfection methods and agents described herein or known in the art (such as liposome-mediated transfection, viral vector methods, or other polynucleotide delivery/introductory methods). Suitable SiRNAs can be selected by examining a transcript of the target polynucleotide (e.g., mRNA) for nucleotide sequences downstream from the AUG start codon and recording the occurrence of each nucleotide and the 3' adjacent 19 to 23 nucleotides as potential siRNA target sites, 25 with sequences having a 21 nucleotide length being preferred. Regions to be avoided for target siRNA sites include the 5' and 3' untranslated regions (UTRs) and regions near the start codon (within 75 bases), as these may be richer in regulatory protein binding sites. UTR-binding proteins and/or translation initiation complexes may interfere with binding of the siRNP endonuclease complex. The selected target sites for siRNA can then be compared to the appropriate genome database (e.g., 30 human, etc.) using BLAST or other sequence comparison algorithms known in the art. Target sequences with significant homology to other coding sequences can be eliminated from consideration. The selected SiRNAs can be produced by chemical synthesis methods known in the art or by in vitro

transcription using commercially available methods and kits such as the SILENCER siRNA construction kit (Ambion, Austin TX).

In alternative embodiments, long-term gene silencing and/or RNAi effects can be induced in selected tissue using expression vectors that continuously express siRNA. This can be accomplished using expression vectors that are engineered to express hairpin RNAs (shRNAs) using methods known in the art (see, e.g., Brummelkamp, T.R. et al. (2002) Science 296:550-553; and Paddison, P.J. et al. (2002) Genes Dev. 16:948-958). In these and related embodiments, shRNAs can be delivered to target cells using expression vectors known in the art. An example of a suitable expression vector for delivery of siRNA is the PSILENCER1.0-U6 (circular) plasmid (Ambion). Once delivered to the target tissue, shRNAs are processed *in vivo* into siRNA-like molecules capable of carrying out genespecific silencing.

In various embodiments, the expression levels of genes targeted by RNAi or PTGS methods can be determined by assays for mRNA and/or protein analysis. Expression levels of the mRNA of a targeted gene, can be determined by northern analysis methods using, for example, the

NORTHERNMAX-GLY kit (Ambion); by microarray methods; by PCR methods; by real time PCR methods; and by other RNA/polynucleotide assays known in the art or described herein. Expression levels of the protein encoded by the targeted gene can be determined by Western analysis using standard techniques known in the art.

An additional embodiment of the invention encompasses a method for screening for a compound which is effective in altering expression of a polynucleotide encoding SECP. Compounds which may be effective in altering expression of a specific polynucleotide may include, but are not limited to, oligonucleotides, antisense oligonucleotides, triple helix-forming oligonucleotides, transcription factors and other polypeptide transcriptional regulators, and non-macromolecular chemical entities which are capable of interacting with specific polynucleotide sequences. Effective compounds may alter polynucleotide expression by acting as either inhibitors or promoters of polynucleotide expression. Thus, in the treatment of disorders associated with increased SECP expression or activity, a compound which specifically inhibits expression of the polynucleotide encoding SECP may be therapeutically useful, and in the treatment of disorders associated with decreased SECP expression or activity, a compound which specifically promotes expression of the polynucleotide encoding SECP may be therapeutically useful.

In various embodiments, one or more test compounds may be screened for effectiveness in altering expression of a specific polynucleotide. A test compound may be obtained by any method

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commonly known in the art, including chemical modification of a compound known to be effective in altering polynucleotide expression; selection from an existing, commercially-available or proprietary library of naturally-occurring or non-natural chemical compounds; rational design of a compound based on chemical and/or structural properties of the target polynucleotide; and selection from a library of chemical compounds created combinatorially or randomly. A sample comprising a polynucleotide encoding SECP is exposed to at least one test compound thus obtained. The sample may comprise, for example, an intact or permeabilized cell, or an in vitro cell-free or reconstituted biochemical system. Alterations in the expression of a polynucleotide encoding SECP are assayed by any method commonly known in the art. Typically, the expression of a specific nucleotide is detected 10 by hybridization with a probe having a nucleotide sequence complementary to the sequence of the polynucleotide encoding SECP. The amount of hybridization may be quantified, thus forming the basis for a comparison of the expression of the polynucleotide both with and without exposure to one or more test compounds. Detection of a change in the expression of a polynucleotide exposed to a test compound indicates that the test compound is effective in altering the expression of the 15 polynucleotide. A screen for a compound effective in altering expression of a specific polynucleotide can be carried out, for example, using a Schizosaccharomyces pombe gene expression system (Atkins, D. et al. (1999) U.S. Patent No. 5,932,435; Arndt, G.M. et al. (2000) Nucleic Acids Res. 28:E15) or a human cell line such as HeLa cell (Clarke, M.L. et al. (2000) Biochem. Biophys. Res. Commun. 268:8-13). A particular embodiment of the present invention involves screening a combinatorial library of oligonucleotides (such as deoxyribonucleotides, ribonucleotides, peptide nucleic acids, and modified oligonucleotides) for antisense activity against a specific polynucleotide sequence (Bruice, T.W. et al. (1997) U.S. Patent No. 5,686,242; Bruice, T.W. et al. (2000) U.S. Patent No. 6,022,691).

Many methods for introducing vectors into cells or tissues are available and equally suitable for use in vivo, in vitro, and ex vivo. For ex vivo therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back into that same patient. Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved using methods which are well known in the art (Goldman, C.K. et al. (1997) Nat. Biotechnol. 15:462-466).

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Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as humans, dogs, cats, cows, horses, rabbits, and monkeys.

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An additional embodiment of the invention relates to the administration of a composition which generally comprises an active ingredient formulated with a pharmaceutically acceptable excipient. Excipients may include, for example, sugars, starches, celluloses, gums, and proteins. Various formulations are commonly known and are thoroughly discussed in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing, Easton PA). Such compositions may consist of SECP, antibodies to SECP, and mimetics, agonists, antagonists, or inhibitors of SECP.

In various embodiments, the compositions described herein, such as pharmaceutical compositions, may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, pulmonary, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual, or rectal means.

Compositions for pulmonary administration may be prepared in liquid or dry powder form.

These compositions are generally aerosolized immediately prior to inhalation by the patient. In the case of small molecules (e.g. traditional low molecular weight organic drugs), aerosol delivery of fast-acting formulations is well-known in the art. In the case of macromolecules (e.g. larger peptides and proteins), recent developments in the field of pulmonary delivery via the alveolar region of the lung have enabled the practical delivery of drugs such as insulin to blood circulation (see, e.g., Patton, J.S. et al., U.S. Patent No. 5,997,848). Pulmonary delivery allows administration without needle injection, and obviates the need for potentially toxic penetration enhancers.

Compositions suitable for use in the invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose. The determination of an effective dose is well within the capability of those skilled in the art.

Specialized forms of compositions may be prepared for direct intracellular delivery of macromolecules comprising SECP or fragments thereof. For example, liposome preparations containing a cell-impermeable macromolecule may promote cell fusion and intracellular delivery of the macromolecule. Alternatively, SECP or a fragment thereof may be joined to a short cationic N-terminal portion from the HIV Tat-1 protein. Fusion proteins thus generated have been found to transduce into the cells of all tissues, including the brain, in a mouse model system (Schwarze, S.R. et al. (1999) Science 285:1569-1572).

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells, or in animal models such as mice, rats, rabbits, dogs, monkeys, or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for

administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for example SECP or fragments thereof, antibodies of SECP, and agonists, antagonists or inhibitors of SECP, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be determined by standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the ED₅₀ (the dose therapeutically effective in 50% of the population) or LD₅₀ (the dose lethal to 50% of the population) statistics. The dose ratio of toxic to therapeutic effects is the therapeutic index, which can be expressed as the LD₅₀/ED₅₀ ratio. Compositions which exhibit large therapeutic indices are preferred. The data obtained from cell culture assays and animal studies are used to formulate a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that includes the ED₅₀ with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, the sensitivity of the patient, and the route of administration.

The exact dosage will be determined by the practitioner, in light of factors related to the subject requiring treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include the severity of the disease state, the general health of the subject, the age, weight, and gender of the subject, time and frequency of administration, drug combination(s), reaction sensitivities, and response to therapy. Long-acting compositions may be administered every 3 to 4 days, every week, or biweekly depending on the half-life and clearance rate of the particular formulation.

Normal dosage amounts may vary from about 0.1 μ g to 100,000 μ g, up to a total dose of about 1 gram, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

DIAGNOSTICS

In another embodiment, antibodies which specifically bind SECP may be used for the diagnosis of disorders characterized by expression of SECP, or in assays to monitor patients being treated with SECP or agonists, antagonists, or inhibitors of SECP. Antibodies useful for diagnostic purposes may be prepared in the same manner as described above for therapeutics. Diagnostic assays for SECP include methods which utilize the antibody and a label to detect SECP in human body

fluids or in extracts of cells or tissues. The antibodies may be used with or without modification, and may be labeled by covalent or non-covalent attachment of a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

A variety of protocols for measuring SECP, including ELISAs, RIAs, and FACS, are known in the art and provide a basis for diagnosing altered or abnormal levels of SECP expression. Normal or standard values for SECP expression are established by combining body fluids or cell extracts taken from normal mammalian subjects, for example, human subjects, with antibodies to SECP under conditions suitable for complex formation. The amount of standard complex formation may be quantitated by various methods, such as photometric means. Quantities of SECP expressed in subject, control, and disease samples from biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing disease.

In another embodiment of the invention, polynucleotides encoding SECP may be used for diagnostic purposes. The polynucleotides which may be used include oligonucleotides, complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantify gene expression in biopsied tissues in which expression of SECP may be correlated with disease. The diagnostic assay may be used to determine absence, presence, and excess expression of SECP, and to monitor regulation of SECP levels during therapeutic intervention.

In one aspect, hybridization with PCR probes which are capable of detecting polynucleotides, including genomic sequences, encoding SECP or closely related molecules may be used to identify nucleic acid sequences which encode SECP. The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification will determine whether the probe identifies only naturally occurring sequences encoding SECP, allelic variants, or related sequences.

Probes may also be used for the detection of related sequences, and may have at least 50% sequence identity to any of the SECP encoding sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NO:52-102 or from genomic sequences including promoters, enhancers, and introns of the SECP gene.

Means for producing specific hybridization probes for polynucleotides encoding SECP include the cloning of polynucleotides encoding SECP or SECP derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes *in vitro* by means of the addition of the appropriate RNA polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter

groups, for example, by radionuclides such as ³²P or ³⁵S, or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems, and the like.

Polynucleotides encoding SECP may be used for the diagnosis of disorders associated with expression of SECP. Examples of such disorders include, but are not limited to, a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, a cancer of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, colon, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus; an autoimmune/inflammatory disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy 15 (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; a cardiovascular disorder such as congestive heart failure, ischemic heart disease, angina pectoris, myocardial infarction, hypertensive heart disease, degenerative valvular heart disease, calcific aortic valve stenosis, congenitally bicuspid aortic valve, mitral annular calcification, mitral valve prolapse, rheumatic fever and rheumatic heart disease, infective endocarditis, nonbacterial thrombotic endocarditis, endocarditis of systemic lupus erythematosus, carcinoid heart disease, cardiomyopathy, myocarditis, pericarditis, neoplastic heart disease, congenital heart disease, 30 complications of cardiac transplantation, arteriovenous fistula, atherosclerosis, hypertension, vasculitis, Raynaud's disease, aneurysms, arterial dissections, varicose veins, thrombophlebitis and phlebothrombosis, vascular tumors, and complications of thrombolysis, balloon angioplasty, vascular

replacement, and coronary artery bypass graft surgery; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous 10 sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorders of the central nervous system including Down syndrome, cerebral palsy, neuroskeletal disorders, autonomic nervous system disorders, cranial nerve disorders, spinal cord diseases, muscular dystrophy and other neuromuscular disorders, peripheral nervous system disorders, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathies, myasthenia 15 gravis, periodic paralysis, mental disorders including mood, anxiety, and schizophrenic disorders, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, Tourette's disorder, progressive supranuclear palsy, corticobasal degeneration, and familial frontotemporal dementia; and a developmental disorder such as renal tubular acidosis, anemia, Cushing's syndrome, achondroplastic dwarfism, Duchenne and Becker muscular dystrophy, epilepsy, gonadal dysgenesis, WAGR syndrome (Wilms' tumor, aniridia, genitourinary abnormalities, and mental retardation), Smith-Magenis syndrome, myelodysplastic syndrome, hereditary mucoepithelial dysplasia, hereditary keratodermas, hereditary neuropathies such as Charcot-Marie-Tooth disease and neurofibromatosis, hypothyroidism, hydrocephalus, seizure disorders such as Syndenham's chorea and cerebral palsy, spina bifida, anencephaly, craniorachischisis, congenital glaucoma, cataract, and sensorineural hearing loss. Polynucleotides encoding SECP may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR technologies; in dipstick, pin, and multiformat ELISA-like assays; and in microarrays utilizing fluids or tissues from patients to detect altered SECP expression. Such qualitative or quantitative methods are well known in the art.

In a particular embodiment, polynucleotides encoding SECP may be used in assays that detect the presence of associated disorders, particularly those mentioned above. Polynucleotides complementary to sequences encoding SECP may be labeled by standard methods and added to a

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fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantified and compared with a standard value. If the amount of signal in the patient sample is significantly altered in comparison to a control sample then the presence of altered levels of polynucleotides encoding SECP in the sample indicates the presence of the associated disorder. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

In order to provide a basis for the diagnosis of a disorder associated with expression of SECP, a normal or standard profile for expression is established. This may be accomplished by combining body fluids or cell extracts taken from normal subjects, either animal or human, with a sequence, or a fragment thereof, encoding SECP, under conditions suitable for hybridization or amplification. Standard hybridization may be quantified by comparing the values obtained from normal subjects with values from an experiment in which a known amount of a substantially purified polynucleotide is used. Standard values obtained in this manner may be compared with values obtained from samples from patients who are symptomatic for a disorder. Deviation from standard values is used to establish the presence of a disorder.

Once the presence of a disorder is established and a treatment protocol is initiated, hybridization assays may be repeated on a regular basis to determine if the level of expression in the patient begins to approximate that which is observed in the normal subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

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With respect to cancer, the presence of an abnormal amount of transcript (either under- or overexpressed) in biopsied tissue from an individual may indicate a predisposition for the development of the disease, or may provide a means for detecting the disease prior to the appearance of actual clinical symptoms. A more definitive diagnosis of this type may allow health professionals to employ preventative measures or aggressive treatment earlier, thereby preventing the development or further progression of the cancer.

Additional diagnostic uses for oligonucleotides designed from the sequences encoding SECP may involve the use of PCR. These oligomers may be chemically synthesized, generated enzymatically, or produced *in vitro*. Oligomers will preferably contain a fragment of a polynucleotide encoding SECP, or a fragment of a polynucleotide complementary to the polynucleotide encoding SECP, and will be employed under optimized conditions for identification of a specific gene or

condition. Oligomers may also be employed under less stringent conditions for detection or quantification of closely related DNA or RNA sequences.

In a particular aspect, oligonucleotide primers derived from polynucleotides encoding SECP may be used to detect single nucleotide polymorphisms (SNPs). SNPs are substitutions, insertions and 5 deletions that are a frequent cause of inherited or acquired genetic disease in humans. Methods of SNP detection include, but are not limited to, single-stranded conformation polymorphism (SSCP) and fluorescent SSCP (fSSCP) methods. In SSCP, oligonucleotide primers derived from polynucleotides encoding SECP are used to amplify DNA using the polymerase chain reaction (PCR). The DNA may be derived, for example, from diseased or normal tissue, biopsy samples, bodily fluids, and the 10 like. SNPs in the DNA cause differences in the secondary and tertiary structures of PCR products in single-stranded form, and these differences are detectable using gel electrophoresis in non-denaturing gels. In fSCCP, the oligonucleotide primers are fluorescently labeled, which allows detection of the amplimers in high-throughput equipment such as DNA sequencing machines. Additionally, sequence database analysis methods, termed in silico SNP (isSNP), are capable of identifying polymorphisms by comparing the sequence of individual overlapping DNA fragments which assemble into a common consensus sequence. These computer-based methods filter out sequence variations due to laboratory preparation of DNA and sequencing errors using statistical models and automated analyses of DNA sequence chromatograms. In the alternative, SNPs may be detected and characterized by mass spectrometry using, for example, the high throughput MASSARRAY system (Sequenom, Inc., San Diego CA).

SNPs may be used to study the genetic basis of human disease. For example, at least 16 common SNPs have been associated with non-insulin-dependent diabetes mellitus. SNPs are also useful for examining differences in disease outcomes in monogenic disorders, such as cystic fibrosis, sickle cell anemia, or chronic granulomatous disease. For example, variants in the mannose-binding 25 lectin, MBL2, have been shown to be correlated with deleterious pulmonary outcomes in cystic fibrosis. SNPs also have utility in pharmacogenomics, the identification of genetic variants that influence a patient's response to a drug, such as life-threatening toxicity. For example, a variation in N-acetyl transferase is associated with a high incidence of peripheral neuropathy in response to the anti-tuberculosis drug isoniazid, while a variation in the core promoter of the ALOX5 gene results in diminished clinical response to treatment with an anti-asthma drug that targets the 5-lipoxygenase pathway. Analysis of the distribution of SNPs in different populations is useful for investigating genetic drift, mutation, recombination, and selection, as well as for tracing the origins of populations

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and their migrations (Taylor, J.G. et al. (2001) Trends Mol. Med. 7:507-512; Kwok, P.-Y. and Z. Gu (1999) Mol. Med. Today 5:538-543; Nowotny, P. et al. (2001) Curr. Opin. Neurobiol. 11:637-641).

Methods which may also be used to quantify the expression of SECP include radiolabeling or biotinylating nucleotides, coamplification of a control nucleic acid, and interpolating results from standard curves (Melby, P.C. et al. (1993) J. Immunol. Methods 159:235-244; Duplaa, C. et al. (1993) Anal. Biochem. 212:229-236). The speed of quantitation of multiple samples may be accelerated by running the assay in a high-throughput format where the oligomer or polynucleotide of interest is presented in various dilutions and a spectrophotometric or colorimetric response gives rapid quantitation.

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In further embodiments, oligonucleotides or longer fragments derived from any of the polynucleotides described herein may be used as elements on a microarray. The microarray can be used in transcript imaging techniques which monitor the relative expression levels of large numbers of genes simultaneously as described below. The microarray may also be used to identify genetic variants, mutations, and polymorphisms. This information may be used to determine gene function, to 15 understand the genetic basis of a disorder, to diagnose a disorder, to monitor progression/regression of disease as a function of gene expression, and to develop and monitor the activities of therapeutic agents in the treatment of disease. In particular, this information may be used to develop a pharmacogenomic profile of a patient in order to select the most appropriate and effective treatment regimen for that patient. For example, therapeutic agents which are highly effective and display the fewest side effects may be selected for a patient based on his/her pharmacogenomic profile.

In another embodiment, SECP, fragments of SECP, or antibodies specific for SECP may be used as elements on a microarray. The microarray may be used to monitor or measure protein-protein interactions, drug-target interactions, and gene expression profiles, as described above.

A particular embodiment relates to the use of the polynucleotides of the present invention to generate a transcript image of a tissue or cell type. A transcript image represents the global pattern of gene expression by a particular tissue or cell type. Global gene expression patterns are analyzed by quantifying the number of expressed genes and their relative abundance under given conditions and at a given time (Seilhamer et al., "Comparative Gene Transcript Analysis," U.S. Patent No. 5,840,484; hereby expressly incorporated by reference herein). Thus a transcript image may be generated by 30 hybridizing the polynucleotides of the present invention or their complements to the totality of transcripts or reverse transcripts of a particular tissue or cell type. In one embodiment, the hybridization takes place in high-throughput format, wherein the polynucleotides of the present

invention or their complements comprise a subset of a plurality of elements on a microarray. The resultant transcript image would provide a profile of gene activity.

Transcript images may be generated using transcripts isolated from tissues, cell lines, biopsies, or other biological samples. The transcript image may thus reflect gene expression in vivo, as in the case of a tissue or biopsy sample, or in vitro, as in the case of a cell line.

Transcript images which profile the expression of the polynucleotides of the present invention may also be used in conjunction with in vitro model systems and preclinical evaluation of pharmaceuticals, as well as toxicological testing of industrial and naturally-occurring environmental compounds. All compounds induce characteristic gene expression patterns, frequently termed 10 molecular fingerprints or toxicant signatures, which are indicative of mechanisms of action and toxicity (Nuwaysir, E.F. et al. (1999) Mol. Carcinog. 24:153-159; Steiner, S. and N.L. Anderson (2000) Toxicol. Lett. 112-113:467-471). If a test compound has a signature similar to that of a compound with known toxicity, it is likely to share those toxic properties. These fingerprints or signatures are most useful and refined when they contain expression information from a large number of genes and 15 gene families. Ideally, a genome-wide measurement of expression provides the highest quality signature. Even genes whose expression is not altered by any tested compounds are important as well, as the levels of expression of these genes are used to normalize the rest of the expression data. The normalization procedure is useful for comparison of expression data after treatment with different compounds. While the assignment of gene function to elements of a toxicant signature aids in interpretation of toxicity mechanisms, knowledge of gene function is not necessary for the statistical matching of signatures which leads to prediction of toxicity (see, for example, Press Release 00-02 from the National Institute of Environmental Health Sciences, released February 29, 2000, available at http://www.niehs.nih.gov/oc/news/toxchip.htm). Therefore, it is important and desirable in toxicological screening using toxicant signatures to include all expressed gene sequences.

In an embodiment, the toxicity of a test compound can be assessed by treating a biological sample containing nucleic acids with the test compound. Nucleic acids that are expressed in the treated biological sample are hybridized with one or more probes specific to the polynucleotides of the present invention, so that transcript levels corresponding to the polynucleotides of the present invention may be quantified. The transcript levels in the treated biological sample are compared with levels in an untreated biological sample. Differences in the transcript levels between the two samples are indicative of a toxic response caused by the test compound in the treated sample.

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Another embodiment relates to the use of the polypeptides disclosed herein to analyze the

proteome of a tissue or cell type. The term proteome refers to the global pattern of protein expression in a particular tissue or cell type. Each protein component of a proteome can be subjected individually to further analysis. Proteome expression patterns, or profiles, are analyzed by quantifying the number of expressed proteins and their relative abundance under given conditions and at a given time. A profile of a cell's proteome may thus be generated by separating and analyzing the polypeptides of a particular tissue or cell type. In one embodiment, the separation is achieved using two-dimensional gel electrophoresis, in which proteins from a sample are separated by isoelectric focusing in the first dimension, and then according to molecular weight by sodium dodecyl sulfate slab gel electrophoresis in the second dimension (Steiner and Anderson, supra). The proteins are visualized in the gel as 10 discrete and uniquely positioned spots, typically by staining the gel with an agent such as Coomassie Blue or silver or fluorescent stains. The optical density of each protein spot is generally proportional to the level of the protein in the sample. The optical densities of equivalently positioned protein spots from different samples, for example, from biological samples either treated or untreated with a test compound or therapeutic agent, are compared to identify any changes in protein spot density related to 15 the treatment. The proteins in the spots are partially sequenced using, for example, standard methods employing chemical or enzymatic cleavage followed by mass spectrometry. The identity of the protein in a spot may be determined by comparing its partial sequence, preferably of at least 5 contiguous amino acid residues, to the polypeptide sequences of interest. In some cases, further sequence data may be obtained for definitive protein identification.

A proteomic profile may also be generated using antibodies specific for SECP to quantify the levels of SECP expression. In one embodiment, the antibodies are used as elements on a microarray, and protein expression levels are quantified by exposing the microarray to the sample and detecting the levels of protein bound to each array element (Lucking, A. et al. (1999) Anal. Biochem. 270:103-111; Mendoze, L.G. et al. (1999) Biotechniques 27:778-788). Detection may be performed by a 25 variety of methods known in the art, for example, by reacting the proteins in the sample with a thiol- or amino-reactive fluorescent compound and detecting the amount of fluorescence bound at each array element.

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Toxicant signatures at the proteome level are also useful for toxicological screening, and should be analyzed in parallel with toxicant signatures at the transcript level. There is a poor 30 correlation between transcript and protein abundances for some proteins in some tissues (Anderson, N.L. and J. Seilhamer (1997) Electrophoresis 18:533-537), so proteome toxicant signatures may be useful in the analysis of compounds which do not significantly affect the transcript image, but which

alter the proteomic profile. In addition, the analysis of transcripts in body fluids is difficult, due to rapid degradation of mRNA, so proteomic profiling may be more reliable and informative in such cases.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins that are expressed in the treated biological sample are separated so that the amount of each protein can be quantified. The amount of each protein is compared to the amount of the corresponding protein in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample. Individual proteins are identified by sequencing the amino acid residues of the individual proteins and comparing these partial sequences to the polypeptides of the present invention.

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In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins from the biological sample are incubated with antibodies specific to the polypeptides of the present invention. The amount of protein recognized by the antibodies is quantified. The amount of protein in the treated biological sample is compared with the amount in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample.

Microarrays may be prepared, used, and analyzed using methods known in the art (Brennan, T.M. et al. (1995) U.S. Patent No. 5,474,796; Schena, M. et al. (1996) Proc. Natl. Acad. Sci. USA 93:10614-10619; Baldeschweiler et al. (1995) PCT application WO95/25116; Shalon, D. et al. (1995) PCT application WO95/35505; Heller, R.A. et al. (1997) Proc. Natl. Acad. Sci. USA 94:2150-2155; Heller, M.J. et al. (1997) U.S. Patent No. 5,605,662). Various types of microarrays are well known and thoroughly described in Schena, M., ed. (1999; <u>DNA Microarrays: A Practical Approach</u>, Oxford University Press, London).

In another embodiment of the invention, nucleic acid sequences encoding SECP may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. Either coding or noncoding sequences may be used, and in some instances, noncoding sequences may be preferable over coding sequences. For example, conservation of a coding sequence among members of a multi-gene family may potentially cause undesired cross hybridization during chromosomal mapping. The sequences may be mapped to a particular chromosome, to a specific region of a chromosome, or to artificial chromosome constructions, e.g., human artificial chromosomes (HACs), yeast artificial chromosomes (YACs), bacterial artificial chromosomes (BACs), bacterial P1 constructions, or single chromosome cDNA libraries (Harrington, J.J. et al. (1997) Nat. Genet. 15:345-

355; Price, C.M. (1993) Blood Rev. 7:127-134; Trask, B.J. (1991) Trends Genet. 7:149-154). Once mapped, the nucleic acid sequences may be used to develop genetic linkage maps, for example, which correlate the inheritance of a disease state with the inheritance of a particular chromosome region or restriction fragment length polymorphism (RFLP) (Lander, E.S. and D. Botstein (1986) Proc. Natl. Acad. Sci. USA 83:7353-7357).

Fluorescent in situ hybridization (FISH) may be correlated with other physical and genetic map data (Heinz-Ulrich, et al. (1995) in Meyers, supra, pp. 965-968). Examples of genetic map data can be found in various scientific journals or at the Online Mendelian Inheritance in Man (OMIM) World Wide Web site. Correlation between the location of the gene encoding SECP on a physical map and a specific disorder, or a predisposition to a specific disorder, may help define the region of DNA associated with that disorder and thus may further positional cloning efforts.

In situ hybridization of chromosomal preparations and physical mapping techniques, such as linkage analysis using established chromosomal markers, may be used for extending genetic maps.

Often the placement of a gene on the chromosome of another mammalian species, such as mouse, may reveal associated markers even if the exact chromosomal locus is not known. This information is valuable to investigators searching for disease genes using positional cloning or other gene discovery techniques. Once the gene or genes responsible for a disease or syndrome have been crudely localized by genetic linkage to a particular genomic region, e.g., ataxia-telangiectasia to 11q22-23, any sequences mapping to that area may represent associated or regulatory genes for further investigation

(Gatti, R.A. et al. (1988) Nature 336:577-580). The nucleotide sequence of the instant invention may also be used to detect differences in the chromosomal location due to translocation, inversion, etc., among normal, carrier, or affected individuals.

In another embodiment of the invention, SECP, its catalytic or immunogenic fragments, or oligopeptides thereof can be used for screening libraries of compounds in any of a variety of drug screening techniques. The fragment employed in such screening may be free in solution, affixed to a solid support, borne on a cell surface, or located intracellularly. The formation of binding complexes between SECP and the agent being tested may be measured.

Another technique for drug screening provides for high throughput screening of compounds having suitable binding affinity to the protein of interest (Geysen, et al. (1984) PCT application

WO84/03564). In this method, large numbers of different small test compounds are synthesized on a solid substrate. The test compounds are reacted with SECP, or fragments thereof, and washed.

Bound SECP is then detected by methods well known in the art. Purified SECP can also be coated

directly onto plates for use in the aforementioned drug screening techniques. Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a solid support.

In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding SECP specifically compete with a test compound for binding SECP. In this manner, antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with SECP.

In additional embodiments, the nucleotide sequences which encode SECP may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

The disclosures of all patents, applications, and publications mentioned above and below, including U.S. Ser. No. 60/357,002, U.S. Ser. No. 60/362,439, and U.S. Ser. No. 60/366,041, are hereby expressly incorporated by reference.

EXAMPLES

20 I. Construction of cDNA Libraries

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Incyte cDNAs were derived from cDNA libraries described in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA). Some tissues were homogenized and lysed in guanidinium isothiocyanate, while others were homogenized and lysed in phenol or in a suitable mixture of denaturants, such as TRIZOL (Invitrogen), a monophasic solution of phenol and guanidine isothiocyanate. The resulting lysates were centrifuged over CsCl cushions or extracted with chloroform. RNA was precipitated from the lysates with either isopropanol or sodium acetate and ethanol, or by other routine methods.

Phenol extraction and precipitation of RNA were repeated as necessary to increase RNA purity. In some cases, RNA was treated with DNase. For most libraries, poly(A)+ RNA was isolated using oligo d(T)-coupled paramagnetic particles (Promega), OLIGOTEX latex particles (QIAGEN, Chatsworth CA), or an OLIGOTEX mRNA purification kit (QIAGEN). Alternatively, RNA was isolated directly from tissue lysates using other RNA isolation kits, e.g., the

POLY(A)PURE mRNA purification kit (Ambion, Austin TX).

In some cases, Stratagene was provided with RNA and constructed the corresponding cDNA libraries. Otherwise, cDNA was synthesized and cDNA libraries were constructed with the UNIZAP vector system (Stratagene) or SUPERSCRIPT plasmid system (Invitrogen), using the recommended procedures or similar methods known in the art (Ausubel et al., supra, ch. 5). Reverse transcription was initiated using oligo d(T) or random primers. Synthetic oligonucleotide adapters were ligated to double stranded cDNA, and the cDNA was digested with the appropriate restriction enzyme or enzymes. For most libraries, the cDNA was size-selected (300-1000 bp) using SEPHACRYL S1000, SEPHAROSE CL2B, or SEPHAROSE CL4B column chromatography (Amersham 10 Biosciences) or preparative agarose gel electrophoresis. cDNAs were ligated into compatible restriction enzyme sites of the polylinker of a suitable plasmid, e.g., PBLUESCRIPT plasmid (Stratagene), PSPORT1 plasmid (Invitrogen, Carlsbad CA), PCDNA2.1 plasmid (Invitrogen), PBK-CMV plasmid (Stratagene), PCR2-TOPOTA plasmid (Invitrogen), PCMV-ICIS plasmid (Stratagene), pIGEN (Incyte Genomics, Palo Alto CA), pRARE (Incyte Genomics), or pINCY (Incyte Genomics), 15 or derivatives thereof. Recombinant plasmids were transformed into competent E. coli cells including XL1-Blue, XL1-BlueMRF, or SOLR from Stratagene or DH5a, DH10B, or ElectroMAX DH10B from Invitrogen.

II. Isolation of cDNA Clones

Plasmids obtained as described in Example I were recovered from host cells by in vivo excision using the UNIZAP vector system (Stratagene) or by cell lysis. Plasmids were purified using at least one of the following: a Magic or WIZARD Minipreps DNA purification system (Promega); an AGTC Miniprep purification kit (Edge Biosystems, Gaithersburg MD); and QIAWELL 8 Plasmid, QIAWELL 8 Plasmid, QIAWELL 8 Ultra Plasmid purification systems or the R.E.A.L. PREP 96 plasmid purification kit from QIAGEN. Following precipitation, plasmids were resuspended in 0.1 ml of distilled water and stored, with or without lyophilization, at 4°C.

Alternatively, plasmid DNA was amplified from host cell lysates using direct link PCR in a high-throughput format (Rao, V.B. (1994) Anal. Biochem. 216:1-14). Host cell lysis and thermal cycling steps were carried out in a single reaction mixture. Samples were processed and stored in 384-well plates, and the concentration of amplified plasmid DNA was quantified fluorometrically using PICOGREEN dye (Molecular Probes, Eugene OR) and a FLUOROSKAN II fluorescence scanner (Labsystems Oy, Helsinki, Finland).

III. Sequencing and Analysis

Incyte cDNA recovered in plasmids as described in Example II were sequenced as follows. Sequencing reactions were processed using standard methods or high-throughput instrumentation such as the ABI CATALYST 800 (Applied Biosystems) thermal cycler or the PTC-200 thermal cycler (MJ Research) in conjunction with the HYDRA microdispenser (Robbins Scientific) or the 5 MICROLAB 2200 (Hamilton) liquid transfer system. cDNA sequencing reactions were prepared using reagents provided by Amersham Biosciences or supplied in ABI sequencing kits such as the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems). Electrophoretic separation of cDNA sequencing reactions and detection of labeled polynucleotides were carried out using the MEGABACE 1000 DNA sequencing system (Amersham Biosciences); 10 the ABI PRISM 373 or 377 sequencing system (Applied Biosystems) in conjunction with standard ABI protocols and base calling software; or other sequence analysis systems known in the art. Reading frames within the cDNA sequences were identified using standard methods (Ausubel et al., supra, ch. 7). Some of the cDNA sequences were selected for extension using the techniques disclosed in Example VIII.

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The polynucleotide sequences derived from Incyte cDNAs were validated by removing vector, linker, and poly(A) sequences and by masking ambiguous bases, using algorithms and programs based on BLAST, dynamic programming, and dinucleotide nearest neighbor analysis. The Incyte cDNA sequences or translations thereof were then queried against a selection of public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases, and BLOCKS, PRINTS, DOMO, PRODOM; PROTEOME databases with sequences from Homo sapiens, Rattus norvegicus, Mus musculus, Caenorhabditis elegans, Saccharomyces cerevisiae, Schizosaccharomyces pombe, and Candida albicans (Incyte Genomics, Palo Alto CA); hidden Markov model (HMM)-based protein family databases such as PFAM, INCY, and TIGRFAM (Haft, D.H. et al. (2001) Nucleic Acids Res. 29:41-43); and HMM-based protein domain databases such as SMART (Schultz, J. et al. (1998) Proc. Natl. Acad. Sci. USA 95:5857-5864; Letunic, I. et al. (2002) Nucleic Acids Res. 30:242-244). (HMM is a probabilistic approach which analyzes consensus primary structures of gene families; see, for example, Eddy, S.R. (1996) Curr. Opin. Struct. Biol. 6:361-365.) The queries were performed using programs based on BLAST, FASTA, BLIMPS, and HMMER. The Incyte cDNA sequences were assembled to produce full length polynucleotide 30 sequences. Alternatively, GenBank cDNAs, GenBank ESTs, stitched sequences, stretched sequences, or Genscan-predicted coding sequences (see Examples IV and V) were used to extend Incyte cDNA assemblages to full length. Assembly was performed using programs based on Phred,

Phrap, and Consed, and cDNA assemblages were screened for open reading frames using programs based on GeneMark, BLAST, and FASTA. The full length polynucleotide sequences were translated to derive the corresponding full length polypeptide sequences. Alternatively, a polypeptide may begin at any of the methionine residues of the full length translated polypeptide. Full length polypeptide sequences were subsequently analyzed by querying against databases such as the GenBank protein databases (genpept), SwissProt, the PROTEOME databases, BLOCKS, PRINTS, DOMO, PRODOM, Prosite, hidden Markov model (HMM)-based protein family databases such as PFAM, INCY, and TIGRFAM; and HMM-based protein domain databases such as SMART. Full length polynucleotide sequences are also analyzed using MACDNASIS PRO software (MiraiBio, Alameda CA) and LASERGENE software (DNASTAR). Polynucleotide and polypeptide sequence alignments are generated using default parameters specified by the CLUSTAL algorithm as incorporated into the MEGALIGN multisequence alignment program (DNASTAR), which also calculates the percent identity between aligned sequences.

Table 7 summarizes the tools, programs, and algorithms used for the analysis and assembly of

Incyte cDNA and full length sequences and provides applicable descriptions, references, and threshold
parameters. The first column of Table 7 shows the tools, programs, and algorithms used, the second
column provides brief descriptions thereof, the third column presents appropriate references, all of
which are incorporated by reference herein in their entirety, and the fourth column presents, where
applicable, the scores, probability values, and other parameters used to evaluate the strength of a

match between two sequences (the higher the score or the lower the probability value, the greater the
identity between two sequences).

The programs described above for the assembly and analysis of full length polynucleotide and polypeptide sequences were also used to identify polynucleotide sequence fragments from SEQ ID NO:52-102. Fragments from about 20 to about 4000 nucleotides which are useful in hybridization and amplification technologies are described in Table 4, column 2.

IV. Identification and Editing of Coding Sequences from Genomic DNA

Putative secreted proteins were initially identified by running the Genscan gene identification program against public genomic sequence databases (e.g., gbpri and gbhtg). Genscan is a general-purpose gene identification program which analyzes genomic DNA sequences from a variety of organisms (Burge, C. and S. Karlin (1997) J. Mol. Biol. 268:78-94; Burge, C. and S. Karlin (1998) Curr. Opin. Struct. Biol. 8:346-354). The program concatenates predicted exons to form an assembled cDNA sequence extending from a methionine to a stop codon. The output of Genscan is a

FASTA database of polynucleotide and polypeptide sequences. The maximum range of sequence for Genscan to analyze at once was set to 30 kb. To determine which of these Genscan predicted cDNA sequences encode secreted proteins, the encoded polypeptides were analyzed by querying against PFAM models for secreted proteins. Potential secreted proteins were also identified by homology to Incyte cDNA sequences that had been annotated as secreted proteins. These selected Genscan-predicted sequences were then compared by BLAST analysis to the genpept and gbpri public databases. Where necessary, the Genscan-predicted sequences were then edited by comparison to the top BLAST hit from genpept to correct errors in the sequence predicted by Genscan, such as extra or omitted exons. BLAST analysis was also used to find any Incyte cDNA or public cDNA coverage of the Genscan-predicted sequences, thus providing evidence for transcription. When Incyte cDNA coverage was available, this information was used to correct or confirm the Genscan-predicted sequence. Full length polynucleotide sequences were obtained by assembling Genscan-predicted coding sequences with Incyte cDNA sequences and/or public cDNA sequences using the assembly process described in Example III. Alternatively, full length polynucleotide sequences were derived entirely from edited or unedited Genscan-predicted coding sequences.

V. Assembly of Genomic Sequence Data with cDNA Sequence Data "Stitched" Sequences

Partial cDNA sequences were extended with exons predicted by the Genscan gene identification program described in Example IV. Partial cDNAs assembled as described in Example III were mapped to genomic DNA and parsed into clusters containing related cDNAs and Genscan exon predictions from one or more genomic sequences. Each cluster was analyzed using an algorithm based on graph theory and dynamic programming to integrate cDNA and genomic information, generating possible splice variants that were subsequently confirmed, edited, or extended to create a full length sequence. Sequence intervals in which the entire length of the interval was present on more than one sequence in the cluster were identified, and intervals thus identified were considered to be equivalent by transitivity. For example, if an interval was present on a cDNA and two genomic sequences, then all three intervals were considered to be equivalent. This process allows unrelated but consecutive genomic sequences to be brought together, bridged by cDNA sequence. Intervals thus identified were then "stitched" together by the stitching algorithm in the order that they appear along their parent sequences to generate the longest possible sequence, as well as sequence variants. Linkages between intervals which proceed along one type of parent sequence (cDNA to cDNA or genomic sequence to genomic sequence) were given preference over linkages which change parent

type (cDNA to genomic sequence). The resultant stitched sequences were translated and compared by BLAST analysis to the genpept and gbpri public databases. Incorrect exons predicted by Genscan were corrected by comparison to the top BLAST hit from genpept. Sequences were further extended with additional cDNA sequences, or by inspection of genomic DNA, when necessary.

"Stretched" Sequences

Partial DNA sequences were extended to full length with an algorithm based on BLAST analysis. First, partial cDNAs assembled as described in Example III were queried against public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases using the BLAST program. The nearest GenBank protein homolog was then compared by BLAST analysis to either Incyte cDNA sequences or GenScan exon predicted sequences described in Example IV. A chimeric protein was generated by using the resultant high-scoring segment pairs (HSPs) to map the translated sequences onto the GenBank protein homolog. Insertions or deletions may occur in the chimeric protein with respect to the original GenBank protein homolog. The GenBank protein homolog, the chimeric protein, or both were used as probes to search for homologous genomic sequences from the public human genome databases. Partial DNA sequences were therefore "stretched" or extended by the addition of homologous genomic sequences. The resultant stretched sequences were examined to determine whether it contained a complete gene.

VI. Chromosomal Mapping of SECP Encoding Polynucleotides

The sequences which were used to assemble SEQ ID NO:52-102 were compared with sequences from the Incyte LIFESEQ database and public domain databases using BLAST and other implementations of the Smith-Waterman algorithm. Sequences from these databases that matched SEQ ID NO:52-102 were assembled into clusters of contiguous and overlapping sequences using assembly algorithms such as Phrap (Table 7). Radiation hybrid and genetic mapping data available from public resources such as the Stanford Human Genome Center (SHGC), Whitehead Institute for Genome Research (WIGR), and Généthon were used to determine if any of the clustered sequences had been previously mapped. Inclusion of a mapped sequence in a cluster resulted in the assignment of all sequences of that cluster, including its particular SEQ ID NO:, to that map location.

Map locations are represented by ranges, or intervals, of human chromosomes. The map position of an interval, in centiMorgans, is measured relative to the terminus of the chromosome's parm. (The centiMorgan (cM) is a unit of measurement based on recombination frequencies between chromosomal markers. On average, 1 cM is roughly equivalent to 1 megabase (Mb) of DNA in humans, although this can vary widely due to hot and cold spots of recombination.) The cM distances

are based on genetic markers mapped by Généthon which provide boundaries for radiation hybrid markers whose sequences were included in each of the clusters. Human genome maps and other resources available to the public, such as the NCBI "GeneMap'99" World Wide Web site (http://www.ncbi.nlm.nih.gov/genemap/), can be employed to determine if previously identified disease genes map within or in proximity to the intervals indicated above.

Association of SECP polynucleotides with Lung Cancer

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Heritable forms of lung carcinoma have not been reported and thus, identification of relevant disease-associated genes through conventional linkage analysis is not possible. However, several studies of sporadic nonsmall cell lung carcinoma (NSCLC) tumors have reported loss of 10 heterozygosity (LOH) in regions of chromosome 11 suggesting the presence of one or more tumor suppressor genes (Bepler, G. and Garcia-Blanco, M.A. (1994) Proc. Natl. Acad. Sci. 91:5513-5517; lizuka, M. (1995) Genes, Chromosomes & Cancer 13:40-46; Rasio, D. (1995) Cancer Research 55:3988-91). In a study of 79 patients with lung cancer, lizuka and coworkers found that 11q14-11q24.2 was deleted in many of the lung tumors studied. Mapping of this region with additional markers showed that the region of chromosome 11q bounded by markers D11S939 and D11S938 was commonly deleted (lizuka, et al., supra). In another study it was shown that human A549 NSCLC cells transformed with a human-derived YAC clone containing a region of chromosome 11q within the region bounded by D11S939 and D11S938, exhibited little or no increase in cell number (versus control cells whose number increased 5-10-fold in the same 5 day period). Further studies of these hybrid cells showed a decrease in tumorigenicity and an increase in latency following injection into athymic, nude mice, as compared with mice injected with control A549 cells. These studies suggest the presence of a tumor suppressor gene within this region of chromosome 11q and support the association of LOH in this region with NSCLC.

Restriction fragment length polymorphism (RFLP) markers shown to be near regions of DNA known as sequence-tagged sites (STS), have been mapped to NT_Contigs generated by the Human Genome Project using ePCR (Schuler, G.D. (1997) Genome Research 7: 541-550, and (1998) Trends Biotechnol. 16(11):456-459). Contigs containing regions of DNA with known disease-associated markers are therefore used to identify SECP sequences that map to disease-associated regions of the genome.

Polynucleotides encoding SECP were mapped to NT_Contigs. Contigs longer than 1Mb were broken into subcontigs of 1Mb length with overlapping sections of 100kb. A preliminary step used an algorithm, similar to MEGABLAST, to define the mRNA sequence /masked genomic DNA contig

pairings. The cDNA/genomic pairings identified by the first algorithm were confirmed, and the SECP polynucleotides mapped to DNA contigs, using SIM4 (Florea, L. et al. (1998) Genome Res. 8:967-74, version May 2000) which had been optimized for high throughput processing and strand assignment confidence). The SIM4 output of the mRNA sequence/genomic contig pairs was further processed to determine the correct location of the SECP polynucleotides on the genomic contig, as well as their strand identity.

SEQ ID NO:63 was mapped to NT_Contig NT_009151_021.8 from Genbank, version 128, covering a 5.5 Mb region of the genome that also contains lung cancer-associated genetic markers D11S939 and D11S938. The maximum distance between SEQ ID NO:63 and markers D11S939 and D11S938, therefore, is 5.5 Mb. Thus, SEQ ID NO:63 is in proximity with genetic markers shown to consistently associate with lung cancer. Therefore, in various embodiments, SEQ ID NO:63 can be used for one or more of the following: i) determination of LOH in persons with lung cancer in the lung cancer disease region at 11q12-24.2, ii) diagnostic assays for lung cancer, and iii) developing therapeutics and/or other treatments for lung cancer.

SEQ ID NO:64 was mapped to NT_Contig NT_009151_021.8 from Genbank, version 128, covering a 5.5 Mb region of the genome that also contains lung cancer-associated genetic markers D11S939 and D11S938. The maximum distance between SEQ ID NO:64 and markers D11S939 and D11S938, therefore, is 5.5 Mb. Thus, SEQ ID NO:64 is in proximity with genetic markers shown to consistently associate with lung cancer. Therefore, in various embodiments, SEQ ID NO:64 can be used for one or more of the following: i) determination of LOH in persons with lung cancer in the lung cancer disease region at 11q12-24.2, ii) diagnostic assays for lung cancer, and iii) developing therapeutics and/or other treatments for lung cancer.

Association of SECP polynucleotides with Alzheimer's Disease

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SECP polynucleotides were mapped to NT_Contigs, available from NCBI, using the following procedures. Contigs longer than 1Mb were broken into subcontigs of 1Mb in length with overlapping sections of 100kb. A preliminary step used an algorithm, similar to MEGABLAST, to define the mRNA sequence/masked genomic DNA contig pairings. The cDNA/genomic pairings identified by the first algorithm were run through Sim4 (Florea, L. et al. (1998) Genome Res. 8:967-74, version May 2000) that had been optimized in house for high throughput and strand assignment confidence). The Sim4 output of the mRNA sequence/genomic contig pairs was further processed to determine the correct location of the SECP polynucleotides on the genomic contig, and also their strand identity.

Loci on chromosomes that map to regions associated with particular diseases can be used as

markers for these particular diseases. These markers then can be used to develop diagnostic and therapeutic tools for these diseases. For example, loci on chromosome 10 are associated with or linked to Alzheimer's disease (AD), a progressive neurodegenerative disease that represents the most common form of dementia (Ait-Ghezala, G. et al. (2002) Neurosci Lett. 325:87-90). AD can be inherited as an autosomal dominant trait. Further, genetic studies have focused on identification of genes that are potential targets for new treatments or improved diagnostics. The deposition and aggregation of β -amyloid in specific regions of the brain are key neuropathological hallmarks of AD. Insulin-degrading enzyme (IDE) can degrade β-amyloid Abraham, R. et al. (2001) Hum. Genet. 109:646-652). The IDE gene has been mapped near an AD-associated locus, 10q23-q25 (Espinosa R. 3rd et al. (1991) Cytogenet. Cell Genet. 57:184-186). Linkage analysis using IDE gene markers was performed on 1426 subjects from 435 families in which at least two family members were affected with AD.

A logarithm of the odds ratio for linkage (lod) score of over 3 indicates a probability of 1 in 1000 that a particular marker was found solely by chance in affected individuals. Significant linkage 15 (lod score of 3.3) was reported between the polymorphic marker D10S583, located at 115.3 cM on chromosome 10, and AD with age of onset ≥50 years (Betram, L. et al. (2000) Science 290:2302-2303). D10S583 maps 36 kb upstream of the IDE gene. Further analysis of this region, however, failed to show association of SNPs (single nucleotide polymorphisms) within the IDE gene and flanking regions with late-onset AD (LOAD), in a study of 134 Caucasian LOAD cases and 111 matched controls from the United Kingdom (Abraham, R. et al, supra). Thus, although the activity of IDE may not influence the susceptibility to LOAD, there is substantial linkage in the chromosomal region containing the IDE gene, marker D10S583, and AD. The IDE gene and D10S583 both map to contig NT_008769, which contains a region of chromosome 10 that is 9.16 Mb in size.

SEQ ID NO:61 mapped to a region of contig NT_008804_002.8 from GenBank (version 128), localizing SEQ ID NO:61 to within 9.16 Mb of the Alzheimer's disease locus on chromosome 10q. Thus, SEQ ID NO:61 is in proximity with loci shown to consistently associate with Alzheimer's disease. Therefore, in various embodiments, SEQ ID NO:61 can be used for one or more of the following: i) linkage analysis of persons and/or families to the AD disease region at 10q, ii) diagnostic assays for AD, and iii) developing therapeutics and/or other treatments for AD.

30 VII. **Analysis of Polynucleotide Expression**

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Northern analysis is a laboratory technique used to detect the presence of a transcript of a gene and involves the hybridization of a labeled nucleotide sequence to a membrane on which RNAs

from a particular cell type or tissue have been bound (Sambrook and Russell, *supra*, ch. 7; Ausubel et al., *supra*, ch. 4).

Analogous computer techniques applying BLAST were used to search for identical or related molecules in databases such as GenBank or LIFESEQ (Incyte Genomics). This analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer search can be modified to determine whether any particular match is categorized as exact or similar. The basis of the search is the product score, which is defined as:

BLAST Score x Percent Identity

5 x minimum {length(Seq. 1), length(Seq. 2)}

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The product score takes into account both the degree of similarity between two sequences and the length of the sequence match. The product score is a normalized value between 0 and 100, and is calculated as follows: the BLAST score is multiplied by the percent nucleotide identity and the product is divided by (5 times the length of the shorter of the two sequences). The BLAST score is calculated by assigning a score of +5 for every base that matches in a high-scoring segment pair (HSP), and -4 for every mismatch. Two sequences may share more than one HSP (separated by gaps). If there is more than one HSP, then the pair with the highest BLAST score is used to calculate the product score. The product score represents a balance between fractional overlap and quality in a BLAST alignment. For example, a product score of 100 is produced only for 100% identity over the entire length of the shorter of the two sequences being compared. A product score of 70 is produced either by 100% identity and 70% overlap at one end, or by 88% identity and 100% overlap at the other. A product score of 50 is produced either by 100% identity and 50% overlap at one end, or 79% identity and 100% overlap.

Alternatively, polynucleotides encoding SECP are analyzed with respect to the tissue sources from which they were derived. For example, some full length sequences are assembled, at least in part, with overlapping Incyte cDNA sequences (see Example III). Each cDNA sequence is derived from a cDNA library constructed from a human tissue. Each human tissue is classified into one of the following organ/tissue categories: cardiovascular system; connective tissue; digestive system; embryonic structures; endocrine system; exocrine glands; genitalia, female; genitalia, male; germ cells; hemic and immune system; liver; musculoskeletal system; nervous system; pancreas; respiratory system; sense organs; skin; stomatognathic system; unclassified/mixed; or urinary tract. The number of libraries in each category is counted and divided by the total number of libraries across all

categories. Similarly, each human tissue is classified into one of the following disease/condition categories: cancer, cell line, developmental, inflammation, neurological, trauma, cardiovascular, pooled, and other, and the number of libraries in each category is counted and divided by the total number of libraries across all categories. The resulting percentages reflect the tissue- and disease-specific expression of cDNA encoding SECP. cDNA sequences and cDNA library/tissue information are found in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA).

VIII. Extension of SECP Encoding Polynucleotides

Full length polynucleotides are produced by extension of an appropriate fragment of the full length molecule using oligonucleotide primers designed from this fragment. One primer was synthesized to initiate 5' extension of the known fragment, and the other primer was synthesized to initiate 3' extension of the known fragment. The initial primers were designed using OLIGO 4.06 software (National Biosciences), or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68°C to about 72°C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations was avoided.

Selected human cDNA libraries were used to extend the sequence. If more than one extension was necessary or desired, additional or nested sets of primers were designed.

High fidelity amplification was obtained by PCR using methods well known in the art. PCR was performed in 96-well plates using the PTC-200 thermal cycler (MJ Research, Inc.). The reaction mix contained DNA template, 200 nmol of each primer, reaction buffer containing Mg²⁺, (NH₄)₂SO₄, and 2-mercaptoethanol, Taq DNA polymerase (Amersham Biosciences), ELONGASE enzyme (Invitrogen), and Pfu DNA polymerase (Stratagene), with the following parameters for primer pair PCI A and PCI B: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C. In the alternative, the parameters for primer pair T7 and SK+ were as follows: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 57°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C.

The concentration of DNA in each well was determined by dispensing 100 μ l PICOGREEN quantitation reagent (0.25% (v/v) PICOGREEN; Molecular Probes, Eugene OR) dissolved in 1X TE and 0.5 μ l of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, Acton MA), allowing the DNA to bind to the reagent. The plate was scanned in a Fluoroskan II (Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantify the

concentration of DNA. A 5 μ l to 10 μ l aliquot of the reaction mixture was analyzed by electrophoresis on a 1 % agarose gel to determine which reactions were successful in extending the sequence.

The extended nucleotides were desalted and concentrated, transferred to 384-well plates, digested with CviII cholera virus endonuclease (Molecular Biology Research, Madison WI), and sonicated or sheared prior to religation into pUC 18 vector (Amersham Biosciences). For shotgun sequencing, the digested nucleotides were separated on low concentration (0.6 to 0.8%) agarose gels, fragments were excised, and agar digested with Agar ACE (Promega). Extended clones were religated using T4 ligase (New England Biolabs, Beverly MA) into pUC 18 vector (Amersham Biosciences), treated with Pfu DNA polymerase (Stratagene) to fill-in restriction site overhangs, and transfected into competent *E. coli* cells. Transformed cells were selected on antibiotic-containing media, and individual colonies were picked and cultured overnight at 37°C in 384-well plates in LB/2x carb liquid media.

The cells were lysed, and DNA was amplified by PCR using Taq DNA polymerase

(Amersham Biosciences) and Pfu DNA polymerase (Stratagene) with the following parameters: Step

1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 72°C, 2 min; Step 5: steps 2, 3, and

4 repeated 29 times; Step 6: 72°C, 5 min; Step 7: storage at 4°C. DNA was quantified by

PICOGREEN reagent (Molecular Probes) as described above. Samples with low DNA recoveries

were reamplified using the same conditions as described above. Samples were diluted with 20%

dimethysulfoxide (1:2, v/v), and sequenced using DYENAMIC energy transfer sequencing primers

and the DYENAMIC DIRECT kit (Amersham Biosciences) or the ABI PRISM BIGDYE

Terminator cycle sequencing ready reaction kit (Applied Biosystems).

In like manner, full length polynucleotides are verified using the above procedure or are used to obtain 5' regulatory sequences using the above procedure along with oligonucleotides designed for such extension, and an appropriate genomic library.

IX. Identification of Single Nucleotide Polymorphisms in SECP Encoding Polynucleotides

Common DNA sequence variants known as single nucleotide polymorphisms (SNPs) were identified in SEQ ID NO:52-102 using the LIFESEQ database (Incyte Genomics). Sequences from the same gene were clustered together and assembled as described in Example III, allowing the identification of all sequence variants in the gene. An algorithm consisting of a series of filters was used to distinguish SNPs from other sequence variants. Preliminary filters removed the majority of

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basecall errors by requiring a minimum Phred quality score of 15, and removed sequence alignment errors and errors resulting from improper trimming of vector sequences, chimeras, and splice variants. An automated procedure of advanced chromosome analysis analysed the original chromatogram files in the vicinity of the putative SNP. Clone error filters used statistically generated algorithms to identify errors introduced during laboratory processing, such as those caused by reverse transcriptase, polymerase, or somatic mutation. Clustering error filters used statistically generated algorithms to identify errors resulting from clustering of close homologs or pseudogenes, or due to contamination by non-human sequences. A final set of filters removed duplicates and SNPs found in immunoglobulins or T-cell receptors.

Certain SNPs were selected for further characterization by mass spectrometry using the high throughput MASSARRAY system (Sequenom, Inc.) to analyze allele frequencies at the SNP sites in four different human populations. The Caucasian population comprised 92 individuals (46 male, 46 female), including 83 from Utah, four French, three Venezualan, and two Amish individuals. The African population comprised 194 individuals (97 male, 97 female), all African Americans. The 15 Hispanic population comprised 324 individuals (162 male, 162 female), all Mexican Hispanic. The Asian population comprised 126 individuals (64 male, 62 female) with a reported parental breakdown of 43% Chinese, 31% Japanese, 13% Korean, 5% Vietnamese, and 8% other Asian. Allele frequencies were first analyzed in the Caucasian population; in some cases those SNPs which showed no allelic variance in this population were not further tested in the other three populations.

Labeling and Use of Individual Hybridization Probes X.

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Hybridization probes derived from SEQ ID NO:52-102 are employed to screen cDNAs, genomic DNAs, or mRNAs. Although the labeling of oligonucleotides, consisting of about 20 base pairs, is specifically described, essentially the same procedure is used with larger nucleotide fragments. Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 software (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250 μ Ci of [7-32P] adenosine triphosphate (Amersham Biosciences), and T4 polynucleotide kinase (DuPont NEN, Boston MA). The labeled oligonucleotides are substantially purified using a SEPHADEX G-25 superfine size exclusion dextran bead column (Amersham Biosciences). An aliquot containing 107 counts per minute of the labeled probe is used in a typical membrane-based hybridization analysis of human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (DuPont NEN).

The DNA from each digest is fractionated on a 0.7% agarose gel and transferred to nylon

membranes (Nytran Plus, Schleicher & Schuell, Durham NH). Hybridization is carried out for 16 hours at 40°C. To remove nonspecific signals, blots are sequentially washed at room temperature under conditions of up to, for example, 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. Hybridization patterns are visualized using autoradiography or an alternative imaging means and 5 compared.

XI. **Microarrays**

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The linkage or synthesis of array elements upon a microarray can be achieved utilizing photolithography, piezoelectric printing (ink-jet printing; see, e.g., Baldeschweiler et al., supra), mechanical microspotting technologies, and derivatives thereof. The substrate in each of the aforementioned technologies should be uniform and solid with a non-porous surface (Schena, M., ed. (1999) DNA Microarrays: A Practical Approach, Oxford University Press, London). Suggested substrates include silicon, silica, glass slides, glass chips, and silicon wafers. Alternatively, a procedure analogous to a dot or slot blot may also be used to arrange and link elements to the surface of a substrate using thermal, UV, chemical, or mechanical bonding procedures. A typical array may be 15 produced using available methods and machines well known to those of ordinary skill in the art and may contain any appropriate number of elements (Schena, M. et al. (1995) Science 270:467-470; Shalon, D. et al. (1996) Genome Res. 6:639-645; Marshall, A. and J. Hodgson (1998) Nat. Biotechnol. 16:27-31).

Full length cDNAs, Expressed Sequence Tags (ESTs), or fragments or oligomers thereof may comprise the elements of the microarray. Fragments or oligomers suitable for hybridization can be selected using software well known in the art such as LASERGENE software (DNASTAR). The array elements are hybridized with polynucleotides in a biological sample. The polynucleotides in the biological sample are conjugated to a fluorescent label or other molecular tag for ease of detection. After hybridization, nonhybridized nucleotides from the biological sample are removed, and a 25 fluorescence scanner is used to detect hybridization at each array element. Alternatively, laser desorbtion and mass spectrometry may be used for detection of hybridization. The degree of complementarity and the relative abundance of each polynucleotide which hybridizes to an element on the microarray may be assessed. In one embodiment, microarray preparation and usage is described in detail below.

Tissue or Cell Sample Preparation

Total RNA is isolated from tissue samples using the guanidinium thiocyanate method and poly(A)* RNA is purified using the oligo-(dT) cellulose method. Each poly(A)* RNA sample is

reverse transcribed using MMLV reverse-transcriptase, 0.05 pg/μl oligo-(dT) primer (21mer), 1X first strand buffer, 0.03 units/μl RNase inhibitor, 500 μM dATP, 500 μM dGTP, 500 μM dTTP, 40 μM dCTP, 40 μM dCTP-Cy3 (BDS) or dCTP-Cy5 (Amersham Biosciences). The reverse transcription reaction is performed in a 25 ml volume containing 200 ng poly(A)* RNA with GEMBRIGHT kits

(Incyte Genomics). Specific control poly(A)* RNAs are synthesized by *in vitro* transcription from non-coding yeast genomic DNA. After incubation at 37° C for 2 hr, each reaction sample (one with Cy3 and another with Cy5 labeling) is treated with 2.5 ml of 0.5M sodium hydroxide and incubated for 20 minutes at 85° C to the stop the reaction and degrade the RNA. Samples are purified using two successive CHROMA SPIN 30 gel filtration spin columns (Clontech, Palo Alto CA) and after

combining, both reaction samples are ethanol precipitated using 1 ml of glycogen (1 mg/ml), 60 ml sodium acetate, and 300 ml of 100% ethanol. The sample is then dried to completion using a SpeedVAC (Savant Instruments Inc., Holbrook NY) and resuspended in 14 μl 5X SSC/0.2% SDS. Microarray Preparation

Sequences of the present invention are used to generate array elements. Each array element is amplified from bacterial cells containing vectors with cloned cDNA inserts. PCR amplification uses primers complementary to the vector sequences flanking the cDNA insert. Array elements are amplified in thirty cycles of PCR from an initial quantity of 1-2 ng to a final quantity greater than 5 µg.

Amplified array elements are then purified using SEPHACRYL-400 (Amersham Biosciences).

Purified array elements are immobilized on polymer-coated glass slides. Glass microscope slides (Corning) are cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides are etched in 4% hydrofluoric acid (VWR Scientific Products Corporation (VWR), West Chester PA), washed extensively in distilled water, and coated with 0.05% aminopropyl silane (Sigma-Aldrich, St. Louis MO) in 95% ethanol. Coated slides are cured in a 110°C oven.

Array elements are applied to the coated glass substrate using a procedure described in U.S. Patent No. 5,807,522, incorporated herein by reference. 1 μ l of the array element DNA, at an average concentration of 100 ng/ μ l, is loaded into the open capillary printing element by a high-speed robotic apparatus. The apparatus then deposits about 5 nl of array element sample per slide.

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Microarrays are UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene).

Microarrays are washed at room temperature once in 0.2% SDS and three times in distilled water.

Non-specific binding sites are blocked by incubation of microarrays in 0.2% casein in phosphate buffered saline (PBS) (Tropix, Inc., Bedford MA) for 30 minutes at 60°C followed by washes in 0.2%

SDS and distilled water as before.

Hybridization

Hybridization reactions contain 9 μl of sample mixture consisting of 0.2 μg each of Cy3 and Cy5 labeled cDNA synthesis products in 5X SSC, 0.2% SDS hybridization buffer. The sample mixture is heated to 65°C for 5 minutes and is aliquoted onto the microarray surface and covered with an 1.8 cm² coverslip. The arrays are transferred to a waterproof chamber having a cavity just slightly larger than a microscope slide. The chamber is kept at 100% humidity internally by the addition of 140 μl of 5X SSC in a corner of the chamber. The chamber containing the arrays is incubated for about 6.5 hours at 60°C. The arrays are washed for 10 min at 45°C in a first wash buffer (1X SSC, 0.1% SDS), three times for 10 minutes each at 45°C in a second wash buffer (0.1X SSC), and dried.

Detection

Reporter-labeled hybridization complexes are detected with a microscope equipped with an Innova 70 mixed gas 10 W laser (Coherent, Inc., Santa Clara CA) capable of generating spectral lines at 488 nm for excitation of Cy3 and at 632 nm for excitation of Cy5. The excitation laser light is focused on the array using a 20X microscope objective (Nikon, Inc., Melville NY). The slide containing the array is placed on a computer-controlled X-Y stage on the microscope and raster-scanned past the objective. The 1.8 cm x 1.8 cm array used in the present example is scanned with a resolution of 20 micrometers.

In two separate scans, a mixed gas multiline laser excites the two fluorophores sequentially. Emitted light is split, based on wavelength, into two photomultiplier tube detectors (PMT R1477, Hamamatsu Photonics Systems, Bridgewater NJ) corresponding to the two fluorophores. Appropriate filters positioned between the array and the photomultiplier tubes are used to filter the signals. The emission maxima of the fluorophores used are 565 nm for Cy3 and 650 nm for Cy5. Each array is typically scanned twice, one scan per fluorophore using the appropriate filters at the laser source, although the apparatus is capable of recording the spectra from both fluorophores simultaneously.

The sensitivity of the scans is typically calibrated using the signal intensity generated by a cDNA control species added to the sample mixture at a known concentration. A specific location on the array contains a complementary DNA sequence, allowing the intensity of the signal at that location to be correlated with a weight ratio of hybridizing species of 1:100,000. When two samples from different sources (e.g., representing test and control cells), each labeled with a different fluorophore, are hybridized to a single array for the purpose of identifying genes that are differentially expressed, the calibration is done by labeling samples of the calibrating cDNA with the two fluorophores and

adding identical amounts of each to the hybridization mixture.

The output of the photomultiplier tube is digitized using a 12-bit RTI-835H analog-to-digital (A/D) conversion board (Analog Devices, Inc., Norwood MA) installed in an IBM-compatible PC computer. The digitized data are displayed as an image where the signal intensity is mapped using a linear 20-color transformation to a pseudocolor scale ranging from blue (low signal) to red (high signal). The data is also analyzed quantitatively. Where two different fluorophores are excited and measured simultaneously, the data are first corrected for optical crosstalk (due to overlapping emission spectra) between the fluorophores using each fluorophore's emission spectrum.

A grid is superimposed over the fluorescence signal image such that the signal from each spot is centered in each element of the grid. The fluorescence signal within each element is then integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used for signal analysis is the GEMTOOLS gene expression analysis program (Incyte Genomics). Array elements that exhibit at least about a two-fold change in expression, a signal-to-background ratio of at least about 2.5, and an element spot size of at least about 40%, are considered to be differentially expressed.

Expression

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For example, SEQ ID NO:62, SEQ ID NO:74, and SEQ ID NO:75 showed increased expression in Tangier disease-derived fibroblasts compared to normal fibroblasts. In addition, both types of cells were cultured in the presence of cholesterol and compared with the same cell type cultured in the absence of cholesterol. Human fibroblasts were obtained from skin explants from both normal subjects and two patients with homozygous Tangier disease. Cell lines were immortalized by transfection with human papillomavirus 16 genes E6 and E7 and a neomycin resistance selectable marker. TD-derived cells are deficient in an assay of apoA-I mediated tritiated cholesterol efflux. Therefore, SEQ ID NO:62, SEQ ID NO:74, and SEQ ID NO:75 are useful in diagnostic assays for Tangier disease.

As another example, SEQ ID NO:62 showed decreased expression in lung tissue affected by adenocarcinoma versus uninvolved lung tissue, as determined by microarray analysis. Moderately differentiated adenocarcinoma of the right lung was compared to grossly uninvolved lung tissue from a 60 year-old donor (Huntsman Cancer Institute). Therefore, SEQ ID NO:62 is useful in monitoring treatment of, and diagnostic assays for, lung adenocarcinoma and other cell proliferative disorders.

As another example, SEQ ID NO:65 showed increased expression in colon tissue affected by adenocarcinoma versus uninvolved colon tissue, as determined by microarray analysis. Gene

expression profiles were obtained by comparing normal colon tissue from a 60-year-old donor to colon adenocarcinoma tumor tissue from the same donor (Huntsman Cancer Institute, Salt Lake City, UT). Therefore, SEQ ID NO:65 is useful in monitoring treatment of, and diagnostic assays for, colon adenocarcinoma and other cell proliferative disorders.

As another example, SEQ ID NO:69 showed increased expression in breast carcinoma cells versus a nonmalignant mammary epithelial cell line. The gene expression profile of a nonmalignant mammary epithelial cell line was compared to the gene expression profiles of breast carcinoma lines at different stages of tumor progression. Cell lines compared included: a) BT-20, a breast carcinoma cell line derived in vitro from the cells emigrating out of thin slices of tumor mass isolated from a 74-year-10 old female, b) BT-474, a breast ductal carcinoma cell line that was isolated from a solid, invasive ductal carcinoma of the breast obtained from a 60-year-old woman, c) BT-483, a breast ductal carcinoma cell line that was isolated from a papillary invasive ductal tumor obtained from a 23-yearold normal, menstruating, parous female with a family history of breast cancer, d) Hs 578T, a breast ductal carcinoma cell line isolated from a 74-year-old female with breast carcinoma, e) MCF7, a 15 nonmalignant breast adenocarcinoma cell line isolated from the pleural effusion of a 69-year-old female, f) MCF-10A, a breast mammary gland (luminal ductal characteristics) cell line isolated. from a 36-year-old woman with fibrocystic breast disease, g) MDA-MB-468, a breast adenocarcinoma cell line isolated from the pleural effusion of a 51-year-old female with metastatic adenocarcinoma of the breast, and h) HMEC, a primary breast epithelial cell line isolated from a normal donor. Therefore, 20 SEQ ID NO:69 is useful in monitoring treatment of, and diagnostic assays for, breast carcinoma and other cell proliferative disorders.

As another example, SEQ ID NO:70 showed decreased expression in C3A cells treated with gemfibrozil versus untreated cells. Early confluent C3A cells were treated with various amounts of Gemfibrozil (120, 600, 800, and 1200 µg/ml) dissolved CMC, for 1, 3, and 6 hours. Parallel samples of C3A cells were treated with 1% CMC only, as a control. Therefore, SEQ ID NO:70 is useful in monitoring treatment of, and diagnostic assays for, coronary heart disease and other autoimmune/inflammatory disorders.

As another example, SEQ ID NO:80 showed differential expression in cancerous tissues versus normal tissues as determined by microarray analysis. In one experiment, the expression of SEQ ID NO:80, as determined by microarray analysis, was decreased by at least two fold in breast tumor tissues relative to normal breast tissues. The breast tumor tissues were harvested from a 43 year old female donor diagnosed with invasive lobular carcinoma. The tumor is well differentiated and

metastatic. The normal breast tissues were harvested from grossly uninvolved breast tissue of the same donor. Therefore, SEQ ID NO:80 is useful as a diagnostic marker or as a potential therapeutic target for breast cancer.

In the second experiment, the expression of SEQ ID NO:80, as determined by microarray analysis, was increased by at least two fold in sigmoid colon tissues relative to normal sigmoid colon tissues. The sigmoid colon tumor tissue which originated from a metastatic gastric sarcoma (stromal tumor) was harvested from a 48 year old female donor. The normal sigmoid colon tissue was harvested from grossly uninvolved sigmoid colon tissue of the same donor. Therefore, SEQ ID NO:80 is useful as a diagnostic marker or as a potential therapeutic target for colon cancer.

In the third experiment, SEQ ID NO:80 also showed decreased expression in tissue affected by lung adenocarcinoma versus normal lung tissue as determined by microarray analysis. A sample of right lung tissue that showed moderately differentiated adenocarcinoma was compared to grossly uninvolved lung tissue from a 60 year old donor (Huntsman Cancer Institute, Salt Lake City, UT). Therefore, SEQ ID NO:80 is useful in diagnostic assays for and monitoring treatment of lung cancer.

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In the fourth experiment, the expression of SEQ ID NO:80 was decreased by at least two fold in ovarian adenocarcinoma tissues relative to grossly uninvolved normal ovarian tissue from a 79 year old donor. Therefore, SEQ ID NO:80 is useful as a diagnostic marker or as a potential therapeutic target for ovarian cancer.

The four experiments described above showed that SEQ ID NO:80 is useful as a diagnostic marker or as a potential therapeutic target for breast, colon, lung, and ovarian cancers.

In an alternative example, the expression of SEQ ID NO:80 were increased by at least two-fold in treated human adipocytes from obese donors when compared to non-treated adipocytes from the same donors. The obese human primary subcutaneous preadipocytes were isolated from adipose tissue of a 36-year-old healthy female with a body mass index (BMI) of 27.7 (overweight but otherwise healthy). The preadipocytes were cultured and induced to differentiate into adipocytes by culturing them in the differentiation medium containing the active components, PPAR-γ agonist and human insulin. Human preadipocytes were treated with human insulin and PPAR-γ agonist for three days and subsequently were switched to medium containing insulin for 5, 9, and 12 more days before the cells were collected for analysis. Differentiated adipocytes were compared to untreated preadipocytes maintained in culture in the absence of inducing agents. An overall differentiation rate of more than 60% was observed after 15 days in culture. The experiment show that SEQ ID NO:80 is useful for the diagnosis, prognosis, or treatment of diabetes mellitus and other disorders, such as

obesity, hypertension, and atherosclerosis.

In another example, SEQ ID NO:81 showed differential expression in inflammatory responses as determined by microarray analysis. The expression of SEQ ID NO:81 was increased by at least two fold in peripheral blood mononuclear cells (PBMCs, 12% B lymphocytes, 40% T lymphocytes, 20% NK cells, 25% monocytes, and 3% various cells that include dendritic and progenitor cells) treated with 10 ng/ml recombinant interleukin 5 (IL-5) for 2 hours relative to untreated cells. IL-5 is a T cell-derived factor that promotes the proliferation, differentiation, and activation of eosinophils. IL-5 exerts its activity on target cells by binding to specific cell surface receptors. The functional high-affinity receptor for human IL-5 is composed of a low-affinity IL-5 binding a-subunit and a non-binding common b-subunit that is shared with the high-affinity receptors for GM-CSF and IL-3. Therefore, this experiment showed that SEQ ID NO:81 is useful in diagnostic assays for inflammatory responses.

In addition, the expression of SEQ ID NO:81 is up-regulated in immature dendritic cells relative to monocytes by at least three-fold. The types of cDNAs that are up-regulated in concert during the transition from monocyte to dendritic cell reflect DC's newly acquired functions, such as antigen uptake. This experiment also showed that SEQ ID NO:81 is useful in diagnostic assays for inflammatory responses.

Further, as determined by microarray analysis, SEQ ID NO:81 showed differential expression in BT483 breast carcinoma cell line versus HMEC primary mammary epithelial cells. BT483 is a breast ductal carcinoma cell line isolated from a papillary invasive ductal tumor from a 23-year-old normal, menstruating, parous female. HMEC, a primary mammary epithelial cell line was derived from normal human mammary tissue (Clonetics, San Diego, CA). The microarray experiments showed that the expression of SEQ ID NO:81 was increased by at least two fold in BT483 breast ductal carcinoma line grown either in the presence or the absence of growth factors or nutrients relative to HMEC primary mammary epithelial cells grown in the absence of growth factors and nutrients. Therefore, SEQ ID NO:81 is useful as diagnostic markers or as potential therapeutic targets for breast cancer.

SEQ ID NO:81 showed differential expression in prostate carcinoma cell lines versus normal prostate epithelial cells as determined by microarray analysis. Three prostate carcinoma cell lines, DU 145, LNCaP, and PC-3 were included in the experiments. DU 145 was isolated from a metastatic site in the brain of a 69 year old male with widespread metastatic prostate carcinoma. DU 145 has no detectable sensitivity to hormones; forms colonies in semi-solid medium; is only weekly positive for

acid phosphatase; and cells are negative for prostate specific antigen (PSA). LNCaP is a prostate carcinoma cell line isolated from a lymph node biopsy of a 50 year old male with metastatic prostate carcinoma. LNCaP expresses PSA, produces prostate acid phosphatase, and expresses androgen receptors. PC-3, a prostate adenocarcinoma cell line, was isolated from a metastatic site in the bone of a 62 year old male with grade IV prostate adenocarcinoma. The normal epithelial cell line, PrEC, is a primary prostate epithelial cell line isolated from a normal donor. In one experiment, the expression of cDNAs from the prostate carcinoma cell lines were compared to that of the normal prostate epithelial cells grown under the same conditions (in the absence of growth factors and hormones). This experiment showed that the expression of SEQ ID NO:81 was increased by at least two fold in 10 both DU145 and LNCaP prostate carcinoma lines relative to PrECs. In the other experiment, the expression of cDNAs from the prostate carcinoma cell lines grown in optimal conditions (in the presence of growth factors and hormones) were compared to that of the normal prostate epithelial cells grown under restrictive conditions (in the absence of growth factors and hormones). The experiment showed that the expression of SEQ ID NO:81 was increased by at least two fold in 15 DU145, LNCaP, and PC-3 prostate carcinoma lines relative to PrECs. Therefore, SEQ ID NO:81 is useful as a diagnostic marker or as a potential therapeutic target for prostate cancers.

In yet another example, SEQ ID NO:82 showed differential expression, as determined by microarray analysis, in liver C3A cells treated with insulin and LY294002. The human C3A cell line is a clonal derivative of HepG2/C3 (hepatoma cell line, isolated from a 15-year-old male with liver tumor), which was selected for strong contact inhibition of growth. The use of a clonal population enhances the reproducibility of the cells. C3A cells have many characteristics of primary human hepatocytes in culture: i) expression of insulin receptor and insulin-like growth factor II receptor; ii) secretion of a high ratio of serum albumin compared with a-fetoprotein iii) conversion of ammonia to urea and glutamine; iv) metabolism of aromatic amino acids; and v) proliferation in glucose-free and insulin-free medium. The C3A cell line is now well established as an in vitro model of the mature human liver (Mickelson et al. (1995) Hepatology 22:866-875; Nagendra et al. (1997) Am J Physiol 272:G408-G416). LY294002 is a PI3 kinase specific inhibitor that promotes cell cycle arrest of C3A cells and skews these cells towards a more 'liver-like' state. This factor also appears to enhance the metabolic activity of the cells, especially with respect to proteins such as P450. In this experiment, 30 C3A cells were starved of insulin for 3 days (-3 d to 0 h) and then further cultured in the presence of both insulin and the PI3-K inhibitor LY924002 for 0 hour, 24 hours, 2 hours and 3 days. The result showed that the expression of SEQ ID NO:82 was increased by at least a two-fold at in expression at

three out of the four time points (0 hour, 24 hours, and 2 hours). This experiment indicates that SEQ ID NO:82 is useful diagnostic assays for liver diseases and as a potential biological marker and therapeutic agent in the treatment of liver diseases and disorders.

In an alternative example, the expression of SEQ ID NO:85 was decreased by at least two fold in peripheral blood mononuclear cells (PBMCs, 12% B lymphocytes, 40% T lymphocytes, 20% NK cells, 25% monocytes, and 3% various cells that include dendritic and progenitor cells) treated with 10 ng/ml recombinant interleukin 4 (IL-4) for 2 hours relative to untreated cells. IL-4 is a pleiotropic cytokine produced by activated T cells, mast cells, and basophils. It was initially identified as a B cell differentiation factor (BCDF) and a B cell stimulatory factor (BSF1). IL-4 exhibits antitumor effects both in vivo and in vitro. Recently, IL-4 was identified as an important regulator for the CD4+ subset (Th1-like vs. Th2-like) development. The biological effects of IL-4 are mediated by the binding of IL-4 to specific cell surface receptors. The functional high-affinity receptor for IL-4 consists of a ligand-binding subunit (IL-4 R) and a second subunit (b chain) that can modulate the ligand binding affinity of the receptor complex. In certain cell types, the gamma chain of the IL-2 receptor complex is a functional b chain of the IL-4 receptor complex. This experiment showed that SEQ ID NO:85 is useful in diagnostic assays for inflammatory responses.

In another example, SEQ ID NO:87 showed differential expression in inflammatory responses as determined by microarray analysis. In one experiment, Raji B lymphoblast cells were stimulated in vitro with 0.1 mM soluble PMA (phorbol 12-myristate 13-acetate) and 1 mg/ml ionomycin for 0.5, 1, 2, 4, and 8 hours. Treated cells were compared to untreated Raji cells kept in culture in the absence of stimuli. Raji is a B lymphoblast cell line (Burkitt's lymphoma) that was isolated from the left jaw of an 11-year-old male. This cell line tests positive for the Epstein-Barr Nuclear Antigen (EBNA) but does not carry any detectable virus particles. Raji has been extensively used to study signaling in human B cells, identify factors produced by human B cells, and study the anti-lymphoma immune response. PMA is a broad activator of the protein kinase C-dependent pathways. Ionomycin is a calcium ionophore that permits the entry of calcium in the cell, hence increasing the cytosolic calcium concentration. The combination of PMA and ionomycin activates two of the major signaling pathways used by mammalian cells to interact with their environment. In T cells, the combination of PMA and ionomycin mimics the type of secondary signaling events elicited during optimal B cell activation. This experiment showed that expression of SEQ ID NO:87 was increased by at least two fold in three out of five time points (1, 2, and 4 hours).

In another experiment, the expression of SEQ ID NO:87 was increased by at least two fold in

peripheral blood mononuclear cells (PBMCs, 12% B lymphocytes, 40% T lymphocytes, 20% NK cells, 25% monocytes, and 3% various cells that include dendritic and progenitor cells) treated with 10 ng/ml tumor necrosis factor alpha (TNF-α) for 2 hours relative to untreated cells. TNF-α is produced by neutrophils, activated lymphocytes, macrophages, NK cells, LAK cells, astrocytes, endothelial cells, smooth muscle cells, and some transformed cells. TNF-α occurs as a secreted, soluble form and a membrane-anchored form, both of which are biologically active. TNF-α plays a critical role in mediation of the inflammatory response and in mediation of resistance to infections and tumor growth.

In the third experiment, the expression of SEQ ID NO:87 was increased by at least two fold in peripheral blood mononuclear cells (PBMCs, 12% B lymphocytes, 40% T lymphocytes, 20% NK cells, 25% monocytes, and 3% various cells that include dendritic and progenitor cells) treated with lipopolysaccharide (LPS) for 2 or 4 hours relative to untreated cells. LPS elicits a variety of inflammatory responses, and because it activates complement by the alternative (properdin) pathway, it is often part of the pathology of gram-negative bacterial infections. It is thought that LPS released into the bloodstream by lysing gram-negative bacteria is first bound by certain plasma proteins identified as LPS-binding proteins. The LPS-binding protein complex interacts with CD14 receptors on monocytes, macrophages, B cells, and other types of receptors on endothelial cells. Activation of human B cells with LPS results in mitogenesis as well as immunoglobulin synthesis. In monocytes and macrophages three types of events are triggered during their interaction with LPS: 1) Production of cytokines, including IL-1, IL-6, IL-8, TNF-a, and platelet-activating factor. These in turn stimulate production of prostaglandins and leukotrienes, powerful mediators of inflammation and septic shock that accompany endotoxin toxemia. 2) Activation of the complement cascade. 3) Activation of the coagulation cascade.

Therefore, the three experiments described above showed that SEQ ID NO:87 is useful in diagnostic assays for inflammatory responses.

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As a further example, the expression of SEQ ID NO:101 was increased by at least two fold in human umbilical vein endothelial cells (HUVECs) treated with 100 nM phorbol 12-myristate 13-acetate (PMA) and 10 ng/mltumor necrosis factor-alpha (TNF-a) for 24 hours relative to untreated HUVECs. Human umbilical vein endothelial cells are primary cells derived from the endothelium of the human umbilical vein and have been used as an experimental model for investigating the role of the endothelium in human vascular biology. PMA is an agonist of protein kinase C (PKC) which is a calcium-activated, phospho-lipid-dependent serine- and threonine-specific kinase that upon activation phosphorylates a broad range of secondary targets. TNF-a is a pleiotropic cytokine that plays a

central role in mediating the inflammatory response through activation of multiple signal transduction pathways. TNF-a is produced by activated lymphocytes, macrophages, and other white blood cells and can activate endothelial cells. Monitoring the endothelial cells' response to TNF-a at the level of mRNA expression can provide information necessary for better understanding of both TNF-a signaling pathways and endothelial cell biology. TNF-a is also known to cause translocation of PKC from the cytosol to the membrane where it phosphorylates a variety of targets. Inhibition of PKC in the TNF-a activation experimental model will help clarify the PKC-dependent events during TNF-a signaling. The experiment indicated that SEQ ID NO:101 is useful in diagnosis, prognosis, or treatment of inflammatory disorders and endothelial-related disorders, such as those defected in vascular tone regulation, coagulation, thrombosis, and atherosclerosis.

XII. Complementary Polynucleotides

Sequences complementary to the SECP-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring SECP. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same procedure is used with smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using OLIGO 4.06 software (National Biosciences) and the coding sequence of SECP. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the SECP-encoding transcript.

20 XIII. Expression of SECP

Expression and purification of SECP is achieved using bacterial or virus-based expression systems. For expression of SECP in bacteria, cDNA is subcloned into an appropriate vector containing an antibiotic resistance gene and an inducible promoter that directs high levels of cDNA transcription. Examples of such promoters include, but are not limited to, the trp-lac (tac) hybrid promoter and the T5 or T7 bacteriophage promoter in conjunction with the lac operator regulatory element. Recombinant vectors are transformed into suitable bacterial hosts, e.g., BL21(DE3). Antibiotic resistant bacteria express SECP upon induction with isopropyl beta-D-thiogalactopyranoside (IPTG). Expression of SECP in eukaryotic cells is achieved by infecting insect or mammalian cell lines with recombinant Autographica californica nuclear polyhedrosis virus (AcMNPV), commonly known as baculovirus. The nonessential polyhedrin gene of baculovirus is replaced with cDNA encoding SECP by either homologous recombination or bacterial-mediated transposition involving transfer plasmid intermediates. Viral infectivity is maintained and the strong polyhedrin promoter

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drives high levels of cDNA transcription. Recombinant baculovirus is used to infect Spodoptera frugiperda (Sf9) insect cells in most cases, or human hepatocytes, in some cases. Infection of the latter requires additional genetic modifications to baculovirus (Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945).

In most expression systems, SECP is synthesized as a fusion protein with, e.g., glutathione Stransferase (GST) or a peptide epitope tag, such as FLAG or 6-His, permitting rapid, single-step, affinity-based purification of recombinant fusion protein from crude cell lysates. GST, a 26-kilodalton enzyme from Schistosoma japonicum, enables the purification of fusion proteins on immobilized glutathione under conditions that maintain protein activity and antigenicity (Amersham Biosciences). 10 Following purification, the GST moiety can be proteolytically cleaved from SECP at specifically engineered sites. FLAG, an 8-amino acid peptide, enables immunoaffinity purification using commercially available monoclonal and polyclonal anti-FLAG antibodies (Eastman Kodak). 6-His, a stretch of six consecutive histidine residues, enables purification on metal-chelate resins (QIAGEN). Methods for protein expression and purification are discussed in Ausubel et al. (supra, ch. 10 and 16). 15 Purified SECP obtained by these methods can be used directly in the assays shown in Examples XVII, XVIII, XIX, and XX, where applicable.

XIV. Functional Assays

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SECP function is assessed by expressing the sequences encoding SECP at physiologically elevated levels in mammalian cell culture systems. cDNA is subcloned into a mammalian expression vector containing a strong promoter that drives high levels of cDNA expression. Vectors of choice include PCMV SPORT plasmid (Invitrogen, Carlsbad CA) and PCR3.1 plasmid (Invitrogen), both of which contain the cytomegalovirus promoter. 5-10 µg of recombinant vector are transiently transfected into a human cell line, for example, an endothelial or hematopoietic cell line, using either liposome formulations or electroporation. 1-2 μ g of an additional plasmid containing sequences 25 encoding a marker protein are co-transfected. Expression of a marker protein provides a means to distinguish transfected cells from nontransfected cells and is a reliable predictor of cDNA expression from the recombinant vector. Marker proteins of choice include, e.g., Green Fluorescent Protein (GFP; Clontech), CD64, or a CD64-GFP fusion protein. Flow cytometry (FCM), an automated, laser optics-based technique, is used to identify transfected cells expressing GFP or CD64-GFP and to evaluate the apoptotic state of the cells and other cellular properties. FCM detects and quantifies the uptake of fluorescent molecules that diagnose events preceding or coincident with cell death. These events include changes in nuclear DNA content as measured by staining of DNA with propidium

iodide; changes in cell size and granularity as measured by forward light scatter and 90 degree side light scatter; down-regulation of DNA synthesis as measured by decrease in bromodeoxyuridine uptake; alterations in expression of cell surface and intracellular proteins as measured by reactivity with specific antibodies; and alterations in plasma membrane composition as measured by the binding of fluorescein-conjugated Annexin V protein to the cell surface. Methods in flow cytometry are discussed in Ormerod, M.G. (1994; Flow Cytometry, Oxford, New York NY).

The influence of SECP on gene expression can be assessed using highly purified populations of cells transfected with sequences encoding SECP and either CD64 or CD64-GFP. CD64 and CD64-GFP are expressed on the surface of transfected cells and bind to conserved regions of human immunoglobulin G (IgG). Transfected cells are efficiently separated from nontransfected cells using magnetic beads coated with either human IgG or antibody against CD64 (DYNAL, Lake Success NY). mRNA can be purified from the cells using methods well known by those of skill in the art. Expression of mRNA encoding SECP and other genes of interest can be analyzed by northern analysis or microarray techniques.

15 XV. Production of SECP Specific Antibodies

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SECP substantially purified using polyacrylamide gel electrophoresis (PAGE; see, e.g., Harrington, M.G. (1990) Methods Enzymol. 182:488-495), or other purification techniques, is used to immunize animals (e.g., rabbits, mice, etc.) and to produce antibodies using standard protocols.

Alternatively, the SECP amino acid sequence is analyzed using LASERGENE software (DNASTAR) to determine regions of high immunogenicity, and a corresponding oligopeptide is synthesized and used to raise antibodies by means known to those of skill in the art. Methods for selection of appropriate epitopes, such as those near the C-terminus or in hydrophilic regions are well described in the art (Ausubel et al., *supra*, ch. 11).

Typically, oligopeptides of about 15 residues in length are synthesized using an ABI 431A peptide synthesizer (Applied Biosystems) using FMOC chemistry and coupled to KLH (Sigma-Aldrich, St. Louis MO) by reaction with N-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS) to increase immunogenicity (Ausubel et al., *supra*). Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. Resulting antisera are tested for antipeptide and anti-SECP activity by, for example, binding the peptide or SECP to a substrate, blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radio-iodinated goat anti-rabbit IgG.

XVI. Purification of Naturally Occurring SECP Using Specific Antibodies

Naturally occurring or recombinant SECP is substantially purified by immunoaffinity

chromatography using antibodies specific for SECP. An immunoaffinity column is constructed by covalently coupling anti-SECP antibody to an activated chromatographic resin, such as CNBr-activated SEPHAROSE (Amersham Biosciences). After the coupling, the resin is blocked and washed according to the manufacturer's instructions.

Media containing SECP are passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of SECP (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt antibody/SECP binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such as urea or thiocyanate ion), and SECP is collected.

XVII. Identification of Molecules Which Interact with SECP

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SECP, or biologically active fragments thereof, are labeled with ¹²⁵I Bolton-Hunter reagent (Bolton, A.E. and W.M. Hunter (1973) Biochem. J. 133:529-539). Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled SECP, washed, and any wells with labeled SECP complex are assayed. Data obtained using different concentrations of SECP are used to calculate values for the number, affinity, and association of SECP with the candidate molecules.

Alternatively, molecules interacting with SECP are analyzed using the yeast two-hybrid system as described in Fields, S. and O. Song (1989; Nature 340:245-246), or using commercially available kits based on the two-hybrid system, such as the MATCHMAKER system (Clontech).

SECP may also be used in the PATHCALLING process (CuraGen Corp., New Haven CT) which employs the yeast two-hybrid system in a high-throughput manner to determine all interactions between the proteins encoded by two large libraries of genes (Nandabalan, K. et al. (2000) U.S. Patent No. 6,057,101).

XVIII. Demonstration of SECP Activity

An assay for growth stimulating or inhibiting activity of SECP measures the amount of DNA synthesis in Swiss mouse 3T3 cells (McKay, L and I. Leigh, eds. (1993) Growth Factors: A Practical Approach, Oxford University Press, New York, NY). In this assay, varying amounts of SECP are added to quiescent 3T3 cultured cells in the presence of [3H]thymidine, a radioactive DNA precursor. SECP for this assay can be obtained by recombinant means or from biochemical preparations. Incorporation of [3H]thymidine into acid-precipitable DNA is measured over an appropriate time interval, and the amount incorporated is directly proportional to the amount of newly synthesized DNA. A linear dose-response curve over at least a hundred-fold SECP concentration range is

indicative of growth modulating activity. One unit of activity per milliliter is defined as the concentration of SECP producing a 50% response level, where 100% represents maximal incorporation of [3H]thymidine into acid-precipitable DNA.

Alternatively, an assay for SECP activity measures the stimulation or inhibition of neurotransmission in cultured cells. Cultured CHO fibroblasts are exposed to SECP. Following endocytic uptake of SECP, the cells are washed with fresh culture medium, and a whole cell voltageclamped Xenopus myocyte is manipulated into contact with one of the fibroblasts in SECP-free medium. Membrane currents are recorded from the myocyte. Increased or decreased current relative to control values are indicative of neuromodulatory effects of SECP (Morimoto, T. et al. (1995) Neuron 15:689-696).

Alternatively, an assay for SECP activity measures the amount of SECP in secretory, membrane-bound organelles. Transfected cells as described above are harvested and lysed. The lysate is fractionated using methods known to those of skill in the art, for example, sucrose gradient ultracentrifugation. Such methods allow the isolation of subcellular components such as the Golgi 15 apparatus, ER, small membrane-bound vesicles, and other secretory organelles. Immunoprecipitations from fractionated and total cell lysates are performed using SECP-specific antibodies, and immunoprecipitated samples are analyzed using SDS-PAGE and immunoblotting techniques. The concentration of SECP in secretory organelles relative to SECP in total cell lysate is proportional to the amount of SECP in transit through the secretory pathway.

Alternatively, AMP binding activity is measured by combining SECP with 32P-labeled AMP. The reaction is incubated at 37°C and terminated by addition of trichloroacetic acid. The acid extract is neutralized and subjected to gel electrophoresis to remove unbound label. The radioactivity retained in the gel is proportional to SECP activity.

XIX. Demonstration of Immunoglobulin Activity

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An assay for SECP activity measures the ability of SECP to recognize and precipitate antigens from serum. This activity can be measured by the quantitative precipitin reaction. (Golub, E.S. et al. (1987) Immunology: A Synthesis, Sinauer Associates, Sunderland, MA, pp. 113-115.) SECP is isotopically labeled using methods known in the art. Various serum concentrations are added to constant amounts of labeled SECP. SECP-antigen complexes precipitate out of solution and are 30 collected by centrifugation. The amount of precipitable SECP-antigen complex is proportional to the amount of radioisotope detected in the precipitate. The amount of precipitable SECP-antigen complex is plotted against the serum concentration. For various serum concentrations, a characteristic

precipitin curve is obtained, in which the amount of precipitable SECP-antigen complex initially increases proportionately with increasing serum concentration, peaks at the equivalence point, and then decreases proportionately with further increases in serum concentration. Thus, the amount of precipitable SECP-antigen complex is a measure of SECP activity which is characterized by sensitivity to both limiting and excess quantities of antigen.

Alternatively, an assay for SECP activity measures the expression of SECP on the cell surface. cDNA encoding SECP is transfected into a non-leukocytic cell line. Cell surface proteins are labeled with biotin (de la Fuente, M.A. et al. (1997) Blood 90:2398-2405). Immunoprecipitations are performed using SECP-specific antibodies, and immunoprecipitated samples are analyzed using SDS-PAGE and immunoblotting techniques. The ratio of labeled immunoprecipitant to unlabeled immunoprecipitant is proportional to the amount of SECP expressed on the cell surface.

Alternatively, an assay for SECP activity measures the amount of cell aggregation induced by overexpression of SECP. In this assay, cultured cells such as NIH3T3 are transfected with cDNA encoding SECP contained within a suitable mammalian expression vector under control of a strong promoter. Cotransfection with cDNA encoding a fluorescent marker protein, such as Green Fluorescent Protein (CLONTECH), is useful for identifying stable transfectants. The amount of cell agglutination, or clumping, associated with transfected cells is compared with that associated with untransfected cells. The amount of cell agglutination is a direct measure of SECP activity.

XX. SECP Secretion Assay

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A high throughput assay may be used to identify polypeptides that are secreted in eukaryotic cells. In an example of such an assay, polypeptide expression libraries are constructed by fusing 5'-biased cDNAs to the 5'-end of a leaderless β -lactamase gene. β -lactamase is a convenient genetic reporter as it provides a high signal-to-noise ratio against low endogenous background activity and retains activity upon fusion to other proteins. A dual promoter system allows the expression of β -lactamase fusion polypeptides in bacteria or eukaryotic cells, using the lac or CMV promoter, respectively.

Libraries are first transformed into bacteria, e.g., E. coli, to identify library members that encode fusion polypeptides capable of being secreted in a prokaryotic system. Mammalian signal sequences direct the translocation of β -lactamase fusion polypeptides into the periplasm of bacteria where they confer antibiotic resistance to carbenicillin. Carbenicillin-selected bacteria are isolated on solid media, individual clones are grown in liquid media, and the resulting cultures are used to isolate library member plasmid DNA.

Mammalian cells, e.g., 293 cells, are seeded into 96-well tissue culture plates at a density of about 40,000 cells/well in 100 µl phenol red-free DME supplemented with 10% fetal bovine serum (FBS) (Life Technologies, Rockville, MD). The following day, purified plasmid DNAs isolated from carbenicillin-resistant bacteria are diluted with 15 µl OPTI-MEM I medium (Life Technologies) to a volume of 25 µl for each well of cells to be transfected. In separate plates, 1 µl LF2000 Reagent (Life Technologies) is diluted into 25 μl/well OPTI-MEM I. The 25 μl diluted LF2000 Reagent is then combined with the 25 µl diluted DNA, mixed briefly, and incubated for 20 minutes at room temperature. The resulting DNA-LF2000 reagent complexes are then added directly to each well of 293 cells. Cells are also transfected with appropriate control plasmids expressing either wild-type βlactamase, leaderless β-lactamase, or, for example, CD4-fused leaderless β-lactamase. 24 hrs following transfection, about 90 μl of cell culture media are assayed at 37°C with 100 μM Nitrocefin (Calbiochem, San Diego, CA) and 0.5 mM oleic acid (Sigma Corp. St. Louis, MO) in 10 mM phosphate buffer (pH 7.0). Nitrocefin is a substrate for β-lactamase that undergoes a noticeable color change from yellow to red upon hydrolysis. β-lactamase activity is monitored over 20 min in a microtiter plate reader at 486 nm. Increased color absorption at 486 nm corresponds to secretion of a B-lactamase fusion polypeptide in the transfected cell media, resulting from the presence of a eukaryotic signal sequence in the fusion polypeptide. Polynucleotide sequence analysis of the corresponding library member plasmid DNA is then used to identify the signal sequence-encoding cDNA. (Described in U.S. Patent application 09/803,317, filed March 9, 2001.)

For example, SEQ ID NO:28 was shown to be a secreted protein using this assay.

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Various modifications and variations of the described compositions, methods, and systems of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. It will be appreciated that the invention provides novel and useful proteins, and their encoding polynucleotides, which can be used in the drug discovery process, as well as methods for using these compositions for the detection, diagnosis, and treatment of diseases and conditions.

Although the invention has been described in connection with certain embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Nor should the description of such embodiments be considered exhaustive or limit the invention to the precise forms disclosed. Furthermore, elements from one embodiment can be readily recombined with elements from one or more other embodiments. Such combinations can form a number of embodiments within the scope of the invention. It is intended that the scope of the invention be

defined by the following claims and their equivalents.

Table

7					
Incyte rroject ID	rolypephae	Incyte	Polynucleotide	Incyte	
	SEC 10 NO:	Polypeptide ID	SEQ ID NO:	Polynucleotide	
				D	Incyte Full Length Clones
7506904		7506904CD1	52	7506904CB1	3216178CA2, 5291455CA2, 90072791CA2, 90109312CA2,
					90109351CA2, 90109375CA2, 90109403CA2, 90109435CA2,
					90109459CA2, 90109467CA2, 90109511CA2, 90109550CA2,
	···				90109559CA2, 90109603CA2, 90109627CA2, 90109635CA2,
			•	•	90109643CA2, 90109651CA2, 90109659CA2, 90109673CA2,
					90109675CA2, 90109682CA2, 90109691CA2
1206909	7	7506909CD1	53	7506909CB1	90109350CA2, 90109404CA2, 90109412CA2, 90109426CA2,
=					90109503CA2, 90109567CA2, 90109573CA2, 90109618CA2,
					90109658CA2, 90109683CA2, 90109690CA2
7507096	3	7507096CD1	54	7507096CB1	90115488CA2, 90115532CA2
7507098	4	7507098CD1	55	7507098CB1	
7507099	5	7507099CD1	56	7507099CB1	
7501399	9	7501399CD1	57	7501399CB1	
7504768	7	7504768CD1	58	7504768CB1	4111686CA2
7500757	8	7500757CD1	59	7500757CB1	90158490CA2
1730616	6	1730616CD1	8	1730616CB1	
190404	10	190404CD1	19	190404CB1	
7500679	11	7500679CD1	62	7500679CB1	
7500687	12	7500687CD1	63	7500687CB1	
7500688	13	7500688CD1	3	7500688CB1	90207830CA2
7500697	14	7500697CD1	65	7500697CB1	4330768CA2, 5072466CA2
7500709	15	7500709CD1	99	7500709CB1	
7500711	16	7500711CD1	29	7500711CB1	90213704CA2
7500723	17	7500723CD1	89	7500723CB1	7978329CA2, 95074077CA2
7500764	18	7500764CD1	69	7500764CB1	7577131CA2, 90197034CA2, 90197102CA2
7500772	19	7500772CD1	70	7500772CB1	
7501350	20	7501350CD1	7.1	7501350CB1	90197157CA2, 90203017CA2
7506396	21	7506396CD1	72	7506396CB1	
7505917	22	7505917CD1	73	7505917CB1	

Table

Incyte Project ID	Polypeptide	Incyte	Polynucleotide	Incyte	
,	SEQ ID NO:	Polypeptide ID	SEQ ID NO:	Polynucleotide	
					Incyte Full Length Clones
7500701	23	7500701CD1	74	7500701CB1	
7500702	24	7500702CD1	75	7500702CB1	90210690CA2
6044343	25	6044343CD1	76	6044343CB1	
7503990	26	7503990CD1	77	7503990CB1	
7504655	27	7504655CD1	78	7504655CB1	
7504690	28	7504690CD1	79	7504690CB1	90053710CA2, 90053818CA2
7504720	29	7504720CD1	80	7504720CB1	8079636CA2
7504722	30	7504722CD1	81	7504722CB1	7256019CA2
7504733	31	7504733CD1	82	7504733CB1	1456958CA2, 95134365CA2
7507100	32	7507100CD1	83	7507100CB1	
7503330	33	7503330CD1	84	7503330CB1	
7504519	34	7504519CD1	85	7504519CB1	90023474CA2, 90024742CA2, 90024758CA2
7504705	35	7504705CD1	98	7504705CB1	5486686CA2
7504738	36	7504738CD1	87	7504738CB1	4349337CA2, 559380CA2
7510280	37	7510280CD1	88	7510280CB1	3182544CA2, 5861015CA2, 7354785CA2, 7946775CA2
7503700	38	7503700CD1	68	7503700CB1	
7504685	39	7504685CD1	8	7504685CB1	2072758CA2, 2208146CA2, 2326887CA2, 2635337CA2,
					3598548CA2, 4152229CA2, 4156112CA2, 4805920CA2,
					95069520CA2, 95069528CA2, 95069552CA2, 95069592CA2,
					95069652CA2, 95069668CA2, 95069676CA2, 95069752CA2,
					95069776CA2, 95069820CA2, 95069828CA2, 95069852CA2,
					95069892CA2
7506844	\$	7506844CD1	91	7506844CB1	90123506CA2
7510259	41	7510259CD1	25	7510259CB1	56010433CA2, 56010441CA2, 56010449CA2, 56010541CA2,
					56010549CA2, 56010557CA2, 90135770CA2, 90135786CA2
7510444	42	7510444CD1	93	7510444CB1	90001102CA2, 90001110CA2, 90001210CA2
7510494	43	7510494CD1	94	7510494CB1	90017546CA2, 90017614CA2
6486485	4	6486485CD1	95	6486485CB1	95166102CA2, 95166118CA2
7503772	45	7503772CD1	96	7503772CB1	

Table 1

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Incyte Project ID	Polypeptide	Incyte	Polynucleotide Incyte	Incyte	
	SEQ ID NO:	Polypeptide ID SEQ ID NO:	SEQ ID NO:	Polynucleotide	
				<u>A</u>	Incyte Full Length Clones
7503773	46	7503773CD1	6	7503773CB1	
7504698	47	7504698CD1	86		5974456CA2, 6024316CA2, 95118339CA2
7510361	48	7510361CD1	8	7510361CB1	
7507013	49	7507013CD1	100	7507013CB1	90122156CA2. 90122188CA2
7510507	50	7510507CD1	101		
90106370	51	90106370CD1	102	_	90106314CA2, 90106346CA2, 90106370CA2, 90106378CA2.
					90106422CA2, 90106470CA2, 90106494CA2, 90106495CA2

Polypeptide SEQ Incyte	Incyte	GenBank ID NO: Probability	Probability	Annotation
D NO:	Polypeptide ID	or PROTEOME	Score	
		ID NO:		
6	1730616CD1	g23320913	1.0E-100	lung cancer metastasis-related protein [Homo sapiens]
23	7500701CD1	g10444289	3.7E-58	[Homo sapiens] MYG1 (probable metal-dependent hydrolase) homolog
24	7500702CD1	g10444289	3.7E-58	[Homo sapiens] MYG1 homolog
27	7504655CD1	568682 SPUF	2.5E-27	[Homo sapiens] Protein with low similarity to a region of PMBP, which is a putative steroid
				membrane receptor that is preferentially expressed in the placenta
34	7504519CD1	g24285996	5.0E-22	G-protein coupled receptor GPR 114 [Homo sapiens]
35	7504705CD1	336408 MGP	4.2E-11	[Homo sapiens][Structural protein][Extracellular matrix (cuticle and basement membrane);
		-		Extracellular (excluding cell wall)] Matrix Gla protein, a vitamin K-dependent calcium-
				binding component of extracellular matrix that inhibits the calcification of arteries and
				cartilage; mutation of the corresponding gene causes Keutel syndrome
				(Chen, L. et al. (1990) Oncogene 5:1391-1395; Munroe, P. B. et al. (1999) Nat. Genet.
				21:142-144.)
37	7510280CD1	690518 CKL.F1	6.8E-11	[Homo sapiens] Chemokine-like factor 1, a secreted chemoattractant for leucocytes,
				neutrophils, monocytes and lymphocytes, stimulates inflammatory response and muscle
				stem cell proliferation; expression is inhibited by IL 10
				(Han, W. et al. (2001) Biochem. J. 357:127-135.)
45	7503772CD1	g20269129	0.0	MEGF6 [Homo sapiens]
		443807 Y64G10 7.3E-204	7.3E-204	(Caenorhabditis elegans) Putative ortholog of D. melanogaster N (Notch)
		A.7		(Zhou, Z. et al. (2001) Cell 104:43-56.)
		716683 MEGF11 7.8E-168	7.8E-168	[Homo sapiens] Protein containing five laminin epidermal growth factor (EGF)-like and 15
				epidermal growth factor (EGF)-like domains, has low similarity to mouse Notch2
		659407 ced-1	2.9E-151	[Caenorhabditis elegans][Receptor (signalling)][Plasma membrane] Notch family member;
	_			putative cell surface phagocytic receptor that recognizes cell corpses in apoptosis
				(Liu, Q. A. et al. (1998) Cell 93:961-972; Wu, Y. C. et al. (2000) Genes And Development
				14:536-548.)
46	7503773CDI	g20269129	0.0	[Rattus norvegicus] MEGF6
				(Nakayama, M. et al. (1998) Genomics 51:27-34.)

Table?

Polypeptide SEQ Incyte	Incyte	GenBank ID NO: Probability	Probability	Annotation
D NO:	Polypeptide ID	or PROTEOME ID NO:	Score	
		662841Egf13	0.0	[Rattus norvegicus] Multiple epidermal growth factor (EGF)-like domains 6, contains 30
				epidermal growth factor-like motifs, putative secreted protein, predicted to bind calcium
				(Nakayama, M. et al. supra)
		443807 Y64G10 0.0	0.0	[Caenorhabditis elegans] Putative ortholog of D. melanogaster N (Notch)
		A.7		(Zhou, Z. et al. (2001) supra.)
		716683 MEGF11 3.3E-192	3.3E-192	[Homo sapiens] Protein containing five laminin epidermal growth factor (EGF)-like and 15
				epidermal growth factor (EGF)-like domains, has low similarity to mouse Notch2
47	7504698CD1	g7416941 2.0E-29	2.0E-29	[Homo sapiens] MS-14
		476049 LOC513	1.7E-30	[Homo sapiens] M5-14 protein, a putative transmembrane protein that is ubiquitously
	-	8		expressed
				(Escarceller, M. et al. (2000) DNA Seq. 11:335-338)
48	7510361CD1		8.0E-92	[Homo sapiens] parotid secretory protein
		582625 Psp	1.7E-23	[Mus musculus][Extracellular (excluding cell wall)] Parotid secretory protein, a bacteria-
	<u></u>			binding protein that may function as an innate antimicrobial agent, aberrant expression is
				observed in nonobese diabetic mice
				(Robinson, C. P. et al. (1997) Am. J. Physiol. 272:G863-871; Gonzalez, M. J. et al. (2000)
				Exp. Mol. Pathol. 69:91-101.)
		751684 Psp	2.4E-22	[Rattus norvegicus][Extracellular (excluding cell wall)] Parotid secretory protein
				(submandibular gland protein A), a major secreted product of the submandibular gland
	·			acinar-cell progenitors and parotid glands, may play a role in bacterial binding, expression
				may be downregulated in diabetes
		,		(Mirels, L. et al. (1998) Biochem. J. 330:437-444; Szczepanski, A. et al. (1998) Eur. J.
				Morphol. Suppl:240-246.)
49	7507013CD1	g2338292	1.5E-12	[Homo sapiens] proline-rich Gla protein 2
				(Kulman, J. D. et al. (1997) Proc. Natl. Acad. Sci. U.S.A. 94:9058-9062.)
		337254PRRG2	1.2E-13	[Homo sapiens][Plasma membrane] Proline-rich Gla (G-carboxyglutamic acid) polypeptide
				2, member of the vitamin K-dependent family of proteins
				(Kulman, J. D. et al. (1997) Proc Natl Acad Sci U S A 94:9058-9062.)

Table

Polypeptide SEQ Incyte ID NO:	Incyte Polypeptide ID	Incyte GenBank ID NO: Probability Polypeptide ID or PROTEOME Score	Probability Score	Annotation
3		ID NO:		
20	(/21020/CDI	812826025	1.6E-184	[Mus musculus] (AK016991) WD domain, G-beta repeat containing protein-data
				source: Pfam, source key: PF00400, evidence: ISS~putative
				(Caminci, P. et al. (1999) Meth. Enzymol. 303:19-44; Caminci, P. et al. (2000) Genome
				Res. 10, 1617-1630.)
		742592 DKFZP4 7.6E-99	7.6E-99	[Homo sapiens] Protein containing WD domains (WD-40 repeat), which may mediate
		34C245		protein-protein interactions
		753721 WDR5	9.4E-37	[Homo sapiens] WD repeat domain 5, contains seven WD domains (WD-40 repeats), which
				likely mediate protein-protein interactions

			5, 75, 1	
SEC SEC		ច	d Signature Sequences, Domains and Motits	Analytical Methods
a ë	Polypeptide ID	Residues		and Databases
_	7506904CD1	120	Signal_cleavage: M1-G37	SPSCAN
			Signal Peptide: M20-G37, M20-T40, M20-C43, M1-G37, M20-S44	HMMER
			Leucine zipper pattern: L5-L26	MOTIFS
			Potential Phosphorylation Sites: S62 T96	MOTIFS
2	7506909CD1	79	Signal_cleavage: M1-G37	SPSCAN
			Signal Peptide: M20-G37, M20-T40, M20-Q43, M1-G37, M20-S42	HMMER
			Leucine zipper pattern: L5-L26	MOTIFS
			Potential Phosphorylation Sites: S64	MOTIFS
3	7507096CD1	270	Signal_cleavage: M1-A21	SPSCAN
			Signal Peptide: M1-S15, M1-A19, M1-A21, M1-G23, M1-S22, M1-W16	HMMER
			Potential Phosphorylation Sites: S22 S31 S41 S222 S240 T155	MOTIFS
4	7507098CD1	225	Signal_cleavage: M1-A60	SPSCAN
			Signal Peptide: M40-S54, M40-A58, M40-A60, M40-G62, M40-S61, M40-W55	HMMER
			MADS-box domain signature and profile: Y87-G169	PROFILESCAN
			Potential Phosphorylation Sites: S61 S70 S80 S171 S181 S220 T190 Y206	MOTIFS
2	7507099CD1	283	Signal_cleavage: M1-A60	SPSCAN
			Signal Peptide: M40-S54, M40-A58, M40-A60, M40-G62, M40-S61, M40-W55	HIMMER
			Potential Phosphorylation Sites: S61 S70 S80 S181 S235 S253 T190	MOTIFS
9	7501399CD1	294	Signal Peptide: M1-V16, M1-K18, M1-T21, M1-T23, M1-R25, M1-K22	HMMER
			Potential Glycosylation Sites: N159 N190 N194	MOTIFS
			Potential Phosphorylation Sites: S68 S161 S174 S176 S195 S235 T23 T35 T196 Y162	MOTIFS
_	7504768CD1	\$	Signal_cleavage: M1-A20	SPSCAN
			Signal Peptide: M1-S18, M1-A20, M1-C24	HMMER
∞	7500757CD1	187	Signal_cleavage: M1-P27	SPSCAN
			Signal Peptide: M1-A24	HIMMER
ŀ			Immunoglobulin domain: G55-V127	HIMMER_PFAM
			Prenyl group binding site (CAAX box):	MOTIFS
			Potential Phosphorylation Sites: S63 S83 S107 S179 T46 T51 T54 T88 Y123	MOTIFS
			Potential Glycosylation Sites: N105	MOTIFS

SEQ	Incyte	Amino Acid	d Signature Sequences, Domains and Motifs	Analytical Methods
ДÖ	Polypeptide ID	Residues		and Databases
6	1730616CD1	261	Signal_cleavage: M1-A29	SPSCAN
	*		Potential Phosphorylation Sites: S143 S146 S211 S259 T209	MOTIFS
			Potential Glycosylation Sites: N20 N141	MOTIFS
2	190404CD1	206	Signal_cleavage: M1-R20	SPSCAN
			Potential Phosphorylation Sites: S4 S18 S50 S136 S169 S207 S363 S364 T199 T309 T353	MOTIFS
			Potential Glycosylation Sites: N327	MOTIFS
11	7500679CD1	69	Signal_cleavage: M1-P44	SPSCAN
			Signal Peptide: M17-P32, M17-L35, M17-P41, M17-G48, M17-P44, M17-S38	HMMER
			Cell attachment sequence: RS0-D52	MOTIFS
			Potential Phosphorylation Sites: T16	MOTIFS
2	7500687CD1	121	Signal_cleavage: M1-S26	SPSCAN
			Signal Peptide: M1-H28, M1-S23, M1-S26	HMMER
			Potential Phosphorylation Sites: S81 S115	MOTIFS
	ı		Potential Glycosylation Sites: N50 N79	MOTIFS
5	7500688CD1	290	Signal_cleavage: M1-G27	SPSCAN
			Signal Peptide: M1-R30, M1-G24, M1-G25, M1-G27	HMMER
			Potential Phosphorylation Sites: S76 S82 S250 S284 T28 T44 T132	MOTIFS
			Potential Glycosylation Sites: N74 N87 N130 N143 N160 N173 N219 N248	MOTIFS
7		27	Signal Peptide: M1-S19, M1-C20, M1-Q22	HMMER
15	7500709CD1	500	Signal_cleavage: M1-S67	SPSCAN
			Cytosolic domain: M1-R48;	TMHMMER
			Transmembrane domain: A49-P71	
			Non-cytosolic domain: C72-A500	
			tial Phosphorylation Sites: S29 S78 S124 S288 S470 T147 T316 T317 T359 T363 T391	MOTIFS
			T393	
	- 1		Potential Glycosylation Sites: N116 N120 N144 N198 N206 N314 N389 N421	MOTIFS
16	7500711CD1	543	Signal_cleavage: M1-S67	SPSCAN

Table

SEO	Incyte	Amino Acid	Amino Acid Signature Sequences. Domains and Motifs	Analytical Methods
<u>A</u>	Polypeptide	Residues		and Databases
ö	О			
			Cytosolic domain: M1-R48	TMHMMER
			Transmembrane domain: A49-P71	
			Non-cytosolic domain: C72-A543	
			Potential Phosphorylation Sites: S29 S78 S124 S279 S435 S437 S513 T138 T307 T308 T350	MOTIFS
			T354 T382 T384	
			Potential Glycosylation Sites: N116 N120 N135 N189 N197 N305 N380 N464	MOTIFS
11	7500723CD1	72	Signal_cleavage: M1-G28	SPSCAN
			Signal_cleavage: W12-G28, S11-A32, A9-A32, M1-A32, M1-P29	HMMER
			Potential Phosphorylation Sites: S11 T5	MOTIFS
18	7500764CD1	22	Signal Peptide: M1-K22	HMMER
61	7500772CD1	26	Signal Peptide: M1-A16, M1-S19, M1-S20, M1-T22, M1-L24, M1-P18	HMMER
			Potential Phosphorylation Sites: S19 T22	MOTIFS
8	7501350CD1	27	Signal Peptide: MI-P18, MI-A20, MI-G22, MI-A16	HIMMER
			Potential Phosphorylation Sites: S19	MOTIFS
77	7506396CD1	253	Signal_cleavage: M1-G54	SPSCAN
			PROTEIN F20D22.3 C47D12.2 PD043239: V21-D160, K205-F248	BLAST_PRODOM
			Potential Phosphorylation Sites: S58 S71 S138 S151 S157 S172 T7 T9 T18 T50	MOTIFS
			Potential Glycosylation Sites: N56 N66 N95 N114	MOTIFS
22	7505917CD1	511	Signal_cleavage: M1-A35	SPSCAN
			PROTEIN CONSERVED PUTATIVE NICOTINATE PHOSPHORIBOSYLTRANSFERASE BLAST_PRODOM	BLAST_PRODOM
			TRANSFERASE GLYCOSYLTRANSFERASE YUEK CY130.15C 392AA PD008895: F92-	
			E223, E268-E480	
			CONSERVED HYPOTHETICAL PROTEIN PD183751: P377-L505	BLAST_PRODOM
			PROTEIN CONSERVED PUTATIVE NICOTINATE PHOSPHORIBOSYLTRANSFERASE BLAST_PRODOM	BLAST_PRODOM
			TRANSFERASE GLYCOSYLTRANSFERASE YUEK CY130.15C PD011757: L16-L80	
			Potential Phosphorylation Sites: S220 S462 S495 T105 T437	MOTIFS
ន	7500701CD1	127	Signal_cleavage: M1-T21	SPSCAN
			Signal Peptide: M1-T21	HMMER

Table .

0	1	Link 2.1.4		
ב ב ב	מביל חווכאופ	אנווווס אכום	Annue Acid Jagnature Sequences, Domains and Motifs	Analytical Methods
Д Ö	Polypeptide ID	Residues		and Databases
			PROTEIN HYDROLASE K08H10.8 BEM2NCB1 INTERGENIC REGION PREDICTED METAL DEPENDENT PD105898: K40-R110	BLAST_PRODOM
			Potential Glycosylation Sites: N51	MOTIFS
73	7500702CD1	137	Signal_cleavage: M1-T21	SPSCAN
			Signal Peptide: M1:T21	HMMER
			PROTEIN HYDROLASE KOSHIO.8 BEM2NCBI INTERGENIC REGION PREDICTED	BLAST_PRODOM
			Potential Cluberdation State, NC1	Commo
į	ı.	200	ICN: NOT	MOTIFS
3	6044343CD1	207		SPSCAN
			13 S181 T80 T84 T92 T98 T102 T106 T114 T146 T150	MOTIFS
	ı		Potential Glycosylation Sites: N174	MOTIFS
8	7503990CD1	1008	signal_cleavage: M1-A34	SPSCAN
			Potential Phosphorylation Sites: S95 S168 S245 S276 S337 S375 S407 S411 S434 S457 S501	MOTIFS
			S531 S548 S564 S580 S625 S670 S680 S684 S761 S792 S800 S804 S848 S850 S882 S891	
			S924 S971 T49 T68 T162 T166 T347 T362 T419 T588 T731 T777 T778 T912 T967 T975	
			T1006	
	- 1		Potential Glycosylation Sites: N47 N142 N172 N207 N225 N226 N230 N586	MOTIFS
23	7504655CD1	70	Signal_cleavage: M1-G25	SPSCAN
				HMMER
			Potential Phosphorylation Sites: S60 T28 T34 T48	MOTIFS
8	7504690CD1	142	Signal_cleavage: M1-L19	SPSCAN
			Potential Phosphorylation Sites: S83 S110 T88 T92 T105 T136	MOTIFS
			Signal Peptide: M1-A17, M1-L19, M1-Q21, M1-S24	HMMER
82	7504720CD1	43	Signal_cleavage: M1-G20	SPSCAN
			Signal Peptide: M1-G20, M1-T22, M1-K24, M1-R28	HMMER
			Sites: S40 T22 T26 T33	MOTIFS
8	7504722CD1	8	Signal_cleavage: M1-R33	SPSCAN

SEO	Incyte	Amino Acid	Signature Sequences, Domains and Motifs	Analytical Methods
		Residues		and Databases
ÿ Ž	a			
			Potential Phosphorylation Sites: S42	MOTIFS
31	7504733CDI	23	Signal Peptide: M1-S15	HMMER
			Signal Peptide: M1-V18, M1-P20, M1-L16, M1-P20	HMMER
32	7507100CD1	349	Signal_cleavage: M1-A60	SPSCAN
			Signal Peptide: M40-S54, M40-A58, M40-A60, M40-G62, M40-S61, M40-W55	HMMER
			Potential Phosphorylation Sites: S61 S70 S80 S181 S240 S289 T190 T265 T344 Y206	MOTIFS
			Potential Glycosylation Sites: N238 N288	MOTIFS
33	7503330CD1	\$9	Signal_cleavage: M1-A27	SPSCAN
			Signal Peptide: M9-A27, M9-S28, M9-A31, M9-L33, M1-S28, M1-A31	HMMER
			Potential Phosphorylation Sites: S5	MOTIFS
34	7504519CD1	2	Signal_cleavage: M1-A19	SPSCAN
			Signal Peptide: MI-A19, MI-T21, MI-T23, MI-E25, MI-E22	HMMER
			Potential Phosphorylation Sites: S29 S45 S46 T23	MOTIFS
			Potential Glycosylation Sites: N18	MOTIFS
33	7504705CD1	94	Signal_cleavage: MI-C19	SPSCAN
			Signal Peptide: MI-V16, MI-C19, MI-E21, MI-S22, MI-E24, MI-L18	HMMER
			Potential Phosphorylation Sites: S87	MOTIFS
			Potential Glycosylation Sites: N78	MOTIFS
36	7504738CD1	52	Signal_cleavage: M1-S23	SPSCAN
			Signal Peptide: M1-A16, M1-C18, M1-P20, M1-S23, M1-A24, M1-P25	HMMER
			Cytosolic domain: M1-T6	TMHMMER
			Transmembrane domain: V7-A24	
			Non-cytosolic domain: P25-N52	
37	7510280CD1	33	Potential Phosphorylation Sites: S16	MOTIFS
88	7503700CD1	242	Signal_cleavage: M1-A20	SPSCAN
			Signal Peptide: M1-A19, M1-S21, M1-A29, M1-A20, M1-G24	HMMER
			Potential Phosphorylation Sites: S21 S174 T46 T176 T203	MOTIFS
			Potential Glycosylation Sites: N220	MOTIFS
39	7504685CD1	47	Signal_cleavage: M1-A18	SPSCAN

Table :

SEQ		cid	Signature Sequences, Domains and Motifs	Analytical Methods
ЭŠ	Polypeptide ID	Residues		and Databases
			Signal Peptide: M1-A18, M1-R20	HMMER
			Cytosolic domain: Q24-Q47	TMHMMER
			Transmembrane domain: T4-L23	
			Non-cytosolic domain: M1-S3	
\$	7506844CD1	53	Signal_cleavage: MI-E24	SPSCAN
			Signal Peptide: M1-G21, M1-E24	HMMER
4	7510259CD1	95	Signal_cleavage: M1-G46	SPSCAN
			Cytosolic domain: M1-R29	TIMHIMMER
			Transmembrane domain: L30-V52	
			Non-cytosolic domain: V53-R95	
			Guanylate cyclases signature: L35-R93	PROFILESCAN
			Potential Phosphorylation Sites: S9 S14 S27 S64 S80	MOTIFS
42	7510444CD1	57	Signal_cleavage: M1-L19	SPSCAN
			Signal Peptide: M1-L19, M1-S20, M1-A22, M1-K24, M1-I25, M1-T27	HMMER
			Cytosolic domain: M1-V6	TMHMMER
			Transmembrane domain: L7-S29	
			Non-cytosolic domain: T30-D57	
			Potential Phosphorylation Sites: T30 T49	MOTIFS
&	7510494CD1	29	Signal_cleavage: M1-A21	SPSCAN
			Signal Peptide: M1-S17, M1-C18, M1-A21, M1-H23, M1-A24, M1-L30	HMMER
			.8 S51	MOTIFS
			ites: N48	MOTIFS
4	6486485CD1	311	Signal Peptide: M1-A26	HMMER
			Signal Peptide: M1-S31, M9-S31	HMMER
			Immunoglobulin domain: T97-L194	HMMER_SMART
			Ig superfamily from SCOP: T93-S197	HMMER_INCY
			Cytosolic domain: R232-G311	TMHMMER
			Transmembrane domain: L209-T231	
			Non-cytosolic domain: M1-P208	

SEQ	SEQ Incyte	Amino Acid	Amino Acid Signature Sequences, Domains and Motifs	Analytical Methods
ДŻ	Polypeptide ID	Residues		and Databases
			Potential Phosphorylation Sites: S36 S67 S147 S197 S250 S297 T166 T179 T186 T231	MOTIFS
			Potential Glycosylation Sites: N99 N 168 N303	MOTIFS
\$	7503772CD1	1097	signal_cleavage: M1-G26	SPSCAN
			Signal Peptide: M1-A22, M1-V24, M1-Y29, M1-G26	HMMER
			EGF-like domain: C60-C95, C101-C137, C143-C178, C184-C219, C230-C265, C271-C305,	HIMMER_PFAM
			C311-C346, C415-C446, C459-C489, C502-C532, C536-C577, C590-C622, C635-C664,	
			C677-C708, C721-C751, C764-C795, C808-C838, C851-C881, C894-C924, C937-C967,	
			C980-C1010, C1023-C1053	
		-	Epidermal growth factor-like domain: E59-L96, S100-V138, P142-E179, E183-Y220, S229-	HMMER SMART
			1266, D270-B306, B310-S347, D414-N447, N458-B490, H501-H533, T535-Q578, E580-	ļ
			G623, N634-E665, G676-Q709, S720-Q752, D763-E796, G807-E839, D850-A882, N893-	
			L925, E927-Q968, A979-E1011, G1022-N1054	
			Calcium-binding EGF-like domain: D56-L96, A97-V138, C143-E179, D180-Y220, V227.	HIMMER SMART
			1266, D267-G304, D307-S347, C459-E490, Q618-E665, C681-Q709, C721-Q752, C894-	
			L925, E927-Q968, C980-E1011	
			Laminin-type epidermal growth factor-like domain: C463-C502, C506-C545, C549-C590,	HMMER SMART
			C594-C635, C639-C677, C681-C721, C725-C764, C768-C808, C812-C851, C855-C894,	
			C898-C937, G942-C980, C984-C1023, C1027-C1069	
			Calcium-binding EGF-like domain proteins pattern proteins BL01187; C265-C276, C71-F86	BLIMPS_BLOCKS
			Type III EGF-like signature PR00011: C471-C489, C690-C708	BLIMPS PRINTS
			Thrombomodulin signature PR00907: C237-P253, L258-C281, G286-C311	BLIMPS PRINTS
			MEGF6 GLYCOPROTEIN EGF-LIKE DOMAIN	BLAST PRODOM
			PD169326; L349-D414	·
			PD165309: N507-C536	
			SURFACE ANTIGEN PROTEIN PRECURSOR SIGNAL REPEAT MEMBRANE GPI	BLAST PRODOM
			ANCHOR 156G 168G PD001714: T470-C924	 I
			PROTEIN TRANSCRIPTIONAL REPEAT TRANSCRIPTION REGULATION DNA.	BLAST PRODOM
			BINDING NUCLEAR SHUTTLE CRAFT PUTATIVE PD014613: C406-C881	1

SEQ	Incyte	Amino Acid	d Signature Sequences, Domains and Motifs	Analytical Methods
ВÄ	D Polypeptide NO: D	Residues		and Databases
			EGF DM00003 P98163 1373-1460: S229-V308, C277-E350, C143-E223 DM00003 P35556 2219-2292: S274-C346	BLAST_DOMO
			EGF-LIKE DOMAIN DM00864 I55476 159-241: N234-C311, C277-D355, R146-E223	BLAST_DOMO
			SUSHI REPEAT DM04887 P16581 1-609: C459-A882	BLAST_DOMO
			Aspartic acid and asparagine hydroxylation site: C71-C82, C195-C206, C281-C292, C322-C333	MOTIFS
			EGF-like domain signature 1: C435-C446, C478-C489, C521-C532, C566-C577, C653-C664, MOTIFS C697-C708, C740-C751, C784-C795, C827-C838, C870-C881, C913-C924, C956-C967, C999-C1010, C1042-C1053	MOTIFS
			EGF-like domain signature 2: C80-C95, C122-C137, C163-C178, C204-C219, C250-C265, C290-C303, C331-C346, C435-C446, C566-C577, C697-C708, C740-C751, C784-C795, C827-C838, C913-C924, C956-C967, C999-C1010, C1042-C1053	MOTIFS
			Calcium-binding EGF-like domain pattern signature: D56-C80, D180-C204, D267-C290, D307-C331	MOTIFS
			Potential Phosphorylation Sites: S49 S201 S313 S347 S567 S1008 T264 T399 T401 T418 T487 T660 T836 T953	MOTIFS
			Potential Glycosylation Sites: N147 N447 N458 N634 N769 N856 N867 N893 N1054	MOTIFS
8	7503773CD1	1350	signal_cleavage: M1-G26	SPSCAN
			Signal Peptide: M1-A22, M1-V24, M1-Y29, M1-G26	HMMER
			05,	HMMER_PFAM
			C677-C708, C721-C751, C764-C795, C808-C838, C851-C881, C894-C924, C937-C967, C980-C1010, C1014-C1054, C1067-C1097, C1110-C1140, C1153-C1183, C1196-C1226, C133, C136, C1	
			C1239-C1269, C1282-C1312	

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מנק	Incyle	Amino Acid	Annuo Acid Signature Sequences, Domains and Motits	Analytical Methods
e ë	ID Polypeptide NO: ID	Residues		and Databases
			Epidermal growth factor-like domain: E59-L96, S100-V138, P142-E179, E183-Y220, S229-I266, D270-E306, E310-S347, D414-N447, N458-E490, H501-H533, T535-Q578, E580-	HMMER_SMART
			G623, N634-E665, G676-Q709, S720-Q752, D763-E796, G807-E839, D850-A882, N893-	
			L925, E927-Q968, A979-E1011, G1022-Q1055, G1066-N1098, N1109-E1141, G1152-	
			E1184, A1195-E1227, P1229-E1270, G1281-N1313	
			Calcium-binding EGF-like domain: D56-L96, A97-V138, C143-E179, D180-Y220, V227-	HMMER_SMART
			1266, D267-G304, D307-S347, C459-E490, Q618-E665, C681-Q709, C721-Q752, C894-	
			L925, E927-Q968, C980-E1011, C1153-E1184, E1199-E1227	
				HMMER_SMART
_			C594-C635, C639-C677, C681-C721, C725-C764, C768-C808, C812-C851, C855-C894,	
			C898-C937, G942-C980, C984-C1023, C1027-C1067, C1071-C1110, C1114-C1153, C1157-	
			C1196, C1200-C1239, C1243-C1282, C1286-N1313	
			Calcium-binding EGF-like domain proteins pattern proteins BL01187; C71-F86	BLIMPS BLOCKS
			Type III EGP-like signature PR00011: C471-C489, C690-C708	BLIMPS PRINTS
			L258-C281, G286-C311	BLIMPS PRINTS
			MEGF6 GLYCOPROTEIN EGF-LIKE DOMAIN	BLAST PRODOM
			PD169326: L349-D414	
			PD165309: N507-C536	
			PROTEIN PRECURSOR SIGNAL REPEAT MEMBRANE GPI	BLAST_PRODOM
			PD001714: H774-C1215	
	-		PROTEIN TRANSCRIPTIONAL REPEAT TRANSCRIPTION REGULATION DNA.	BLAST PRODOM
			BINDING NUCLEAR SHUTTLE CRAFT PUTATIVE	
			PD014613: C406-C881, G802-C1312	
			BGF	BLAST DOMO
			DM00003 P98163 1373-1460: S229-V308, C277-E350, C143-E223	
			DM00003 P35556 2219-2292: S274-C346	
			DM00003 A57278 2213-2286: S274-C346	

Table 3

SEO	Incyte	Amino Acid	Signature Sequences, Domains and Motifs	Analytical Methods
A	Polypeptide	Residues		and Databases
<u> </u>			EGF-LIKE DOMAIN DM00864lfs476l159-241: N234-C311, C277-D355, R146-E223	BLAST_DOMO
			C206, C281-C292, C322-	MOTIFS
			EGF-like domain signature 1: C435-C446, C478-C489, C521-C532, C566-C577, C653-C664, MOTIFS C697-C708, C740-C751, C784-C795, C827-C838, C870-C881, C913-C924, C956-C967, C999-C1010, C1043-C1054, C1086-C1097, C1129-C1140, C1172-C1183, C1215-C126,	MOTIFS
			C1258-C1269, C1301-C1312	
			C80-C95, C122-C137, C163-C178, C204-C219, C250-C265, C-C446, C566-C517, C607-C708, C740-C751, C784-C795	MOTIFS
			C827-C838, C913-C924, C956-C967, C999-C1010, C1043-C1054, C1086-C1097, C1172-C1183, C1215-C1226, C1258-C1269	
			ttern signature: D56-C80, D180-C204, D267-C290,	MOTIFS
			Potential Phosphorylation Sites: S49 S201 S313 S347 S567 S1008 S1018 S1158 S1206 S1255 MOTIFS S1277 S1337 T264 T399 T401 T418 T487 T660 T836 T953 T1179 T1214 T1341	MOTIFS
			Potential Glycosylation Sites: N147 N447 N458 N634 N769 N856 N867 N893 N1098 N1109 MOTIFS N1169 N1204 N1205	MOTIFS
14	7504698CD1	71	signal_cleavage: M1-A24	SPSCAN
			Signal Peptide: M1-A25, M1-A30, M1-A24	HMMER
			Vitamin K-dependent carboxylation domain: V3-F71	PROFILESCAN
			ATP/GTP-binding site motif A (P-loop): G63-S70	MOTIFS
			Potential Phosphorylation Sites: S53	MOTIFS
48	7510361CD1	220	signal_cleavage: MI-S18	SPSCAN
			Signal Peptide: M1-T15, M1-E19, M1-S20	HMMER
L_			PROTEIN SALIVARY PRECURSOR GLAND SIGNAL BSP30 PAROTID SECRETORY PSP SUBMANDIBULAR PD011295; M1-1192	BLAST_PRODOM
			I SI SOBMEM DESCRIPTION I POSTESSION IN THE PROPERTY OF THE PR	

SEO	SEQ Incyte	Amino Acid	Signature Sequences, Domains and Motifs	Analytical Methods
ДÖ	Polypeptide ID	Residues		and Databases
			PAROTID SECRETORY PROTEIN DM04779[P07743]12-234: G12-L186 DM04779[B42337]12-235: G12-L186	BLAST_DOMO
			Leucine zipper pattern: L44-L65	MOTIFS
			Potential Phosphorylation Sites: S20 S184 T50 T95	MOTIFS
			Potential Glycosylation Sites: N124 N132	MOTIFS
\$	7507013CD1	39	Signal_cleavage: M1-D19	SPSCAN
			Signal Peptide: M1-D19, M1-P22, M1-E25, M1-S23, M1-T20	HMMER
			PEP-utilizing enzymes signatures: M1-V38	PROFILESCAN
			PROLINERICH GLA PROTEIN 2 PD059428: MI-G33	BLAST_PRODOM
			Potential Phosphorylation Sites: S21 T16 T26	MOTIFS
လ	7510507CD1	451	Signal_cleavage: MI-AS3	SPSCAN
			WD domain, G-beta repeat: F136-D172, L220-D256, K94-S130, R52-188, L262-R298, L10-N46, C178-D214	HMMER_PFAM
		,	WD repeats: D7-N46, P49-188, K91-S130, R133-D172, N175-D214, V217-D256, E259-R298 HMMER SMART	HAMER SMART
			one copy of WD repeat: L10-N46, R52-188, G93-S130, R135-D172, T174-D214, V217-D256, HMMER_SMART G260-R298	HMMER_SMART
			Trp-Asp (WD) repeat proteins proteins BL00678: S161-W171	BLIMPS_BLOCKS
			Trp-Asp (WD-40) repeats signature: C149-G198, D249-F278, V192-F236, T64-A112, T22-S79, S107-P154	PROFILESCAN
			Beta G-protein (transducin) signature PR00319: I159-T173, P196-W213	BLIMPS_PRINTS
			G-protein beta WD-40 repeat signature PR00320: 1159-T173	BLIMPS_PRINTS
			PROTEIN REPEAT WD TRPASP REPEATS CONTAINING CHROMOSOME NUCLEAR FACTOR I PDOMOKI: \$203-1.247 \$96-W120	BLAST_PRODOM
			Trp-Asp (WD) repeats signature: L33-F47, IIS9-T173, I201-V215	MOTIFS
			Potential Phosphorylation Sites: S3 S125 S161 S349 S393 S420 T22 T83 T119 T167 T174 T209 T251 T299 T310 T352 T378 T379 T383 T427 Y363	MOTIFS
			Potential Glycosylation Sites: N181	MOTIFS

Table :

S D SE	SEQ Incyte Amino A D Polypeptide Residues NO: ID	Amino Acid Residues	Amino Acid Signature Sequences, Domains and Motifs Residues	Analytical Methods and Databases
51	90106370CD1 224		Signal_cleavage: MI-AS3	SPSCAN
	•		Signal Peptide: M35-A53	HMMER
			Cytosolic domain: E54-F224 Transmembrane domain: C31-A53	TMHMMER
			Non-cytosolic domain: M1-P30	
			Potential Phosphorylation Sites: S9 S66 S168 T87 T104 T141 Y96	MOTIFS

Polymicleotide	Caniance Proments
Incyte ID/	
Sequence Length	
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Polynucleotide SEQ ID NO:/	Sequence Fragments
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1776	



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	1550-2277, 1561-2097, 1586-2127, 1615-1998, 1644-2062, 1657-2171, 1762-2129, 1764-2087, 1816-2090, 1819-2058, 1819-2275, 1831-
	2482, 1889-2141, 1917-2168, 1936-2199, 1936-2201, 1941-2200, 1952-2330, 1958-2589, 1964-2230, 1991-2499, 2008-2231, 2038-2264.
	2063-2465, 2063-2468, 2090-2326, 2111-2400, 2115-2679, 2132-2674, 2149-2432, 2157-2390, 2158-2340, 2167-2410, 2245-2912, 2256-
	2708, 2270-2494, 2270-2723, 2299-2636, 2317-3080, 2337-2717,
	2340-2562, 2340-2580, 2340-2860, 2340-2951, 2340-3032, 2352-3001, 2360-3063, 2365-3050, 2375-2882, 2379-3060, 2380-2624, 2397.
-	2653, 2428-2708, 2429-2680, 2449-3047, 2451-3047, 2451-3064, 2463-3080, 2473-3024, 2476-2714, 2493-3080, 2505-3067, 2548-3080,
	2563-3080, 2573-2718, 2573-2817, 2588-2949, 2613-2978, 2619-3067, 2620-3065, 2621-3065, 2632-3080, 2640-3069, 2643-3065, 2649-
_	2909, 2652-3070, 2653-3065, 2659-3064, 2666-3065, 2686-3069, 2721-2956, 2721-3076, 2729-3069, 2734-2847, 2753-3063, 2762-3068.
	2763-3067, 2773-3063, 2780-3052, 2782-3069, 2797-3039, 2799-3057, 2804-3082, 2807-3082, 2813-3072, 2820-2970, 2820-3069, 2850-
	3073, 2861-3051, 2867-3066, 2910-3080
102/	1-580, 1-601, 1-648, 1-743, 1-747, 1-764, 1-771, 1-774, 1-779, 1-781, 1-822, 1-825, 1-844, 1-846, 1-852, 1-854, 1-857, 1-869, 2-579, 2-
90106370CB1/	700, 2-778, 5-808, 5-869, 9-869, 31-869, 36-869, 37-869, 43-516, 43-604, 43-680, 43-685, 43-705, 43-711, 43-735, 43-803, 43-834, 43-
1548	852, 43-854, 71-869, 73-869, 88-869, 159-869, 166-869, 231-869, 234-987, 265-869, 351-577, 627-1367, 634-1269, 662-1532, 712-1424
	751-862, 766-1239, 781-1548

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Table 5

	Incyte Project ID:	Representative Library
ID NO:	7506904CB1	CORPNOTO
52 53	7506909CB1	CORPNOTO2
54		KIDNNOT05
	7507096CB1	KIDNNOC01
55	7507098CB1	KIDNNOC01
56	7507099CB1	KIDNNOC01
57	7501399CB1	BRACDIK08
58	7504768CB1	PROSDITO1
59	7500757CB1	BRAINOT09
60	1730616CB1	FIBPFEN06
61	190404CB1	SYNORAB01
62	7500679CB1	LUNLTMT01
63	7500687CB1	BRAENOT02
64	7500688CB1	BRAENOT02
65	7500697CB1	KIDNNOT32
66	7500709CB1	SINTNOR01
67	7500711CB1	SINTNOR01
68	7500723CB1	EOSITXT01
69	7500764CB1	PENITUT01
70	7500772CB1	SINTNOR01
71	7501350CB1	MONOTXT02
72	7506396CB1	LUNGAST01
73	7505917CB1	SINTFER02
74	7500701CB1	BRAPDITOI
75	7500702CB1	BRSTNOT09
76	6044343CB1	BRAUTDR02
77	7503990CB1	BRSTNOT05
78	7504655CB1	PANCNOT01
79	7504690CB1	NERDTDN03
80	7504720CB1	NGANNOT01
81	7504722CB1	293TF3T01
82	7504733CB1	COLNFET02
83	7507100CB1	KIDNNOC01
84	7503330CB1	KIDNFEE02
85	7504519CB1	LNODNOT05
86	7504705CB1	MYOMNOT01
87	7504738CB1	MPHGLPT02
88	7510280CB1	BRABNOE02
89	7503700CB1	LUNGNOT02
90	7504685CB1	ADRENOTO7
91	7506844CB1	LUNGFET05
	7510259CB1	
92		null lib
93	7510444CB1	LIVRTMR01
94	7510494CB1	MPHGNOT03
95	6486485CB1	MIXDUNB01
96	7503772CB1	PROSNOT28
97	7503773CB1	ENDCNOT03
98	7504698CB1	HEARNON08
99	7510361CB1	LSUBNOT03
100	7507013CB1	BRSTNOT02

165

WO 03/068943 PCT/US03/04712

Table 5

Polynucleotide SEQ	Incyte Project ID:	Representative Library
ID NO:	<u> </u>	
101	7510507CB1	SCOMDIC01
102	90106370CB1	SPLNDIC01

Table (

Library	Vector	Library Description
293TF3T01	pINCY	Library was constructed using RNA isolated from a serum-starved transformed embryonal cell line (293-EBNA) derived from kidney epithelial tissue. The cells were transformed with adenovirus 5 DNA.
ADRENOT07	pINCY	Library was constructed using RNA isolated from adrenal tissue removed from a 61-year-old female during a bilateral adrenalectomy. Patient history included an unspecified disorder of the adrenal glands.
BRABNOE02	PBK-CMV	This 5' biased random primed library was constructed using RNA isolated from vermis tissue removed from a 35-year-old Caucasian male who died from cardiac failure. Pathology indicated moderate leptomeningeal fibrosis and multiple microinfarctions of the cerebral neocortex. Patient history included dilated cardiomyopathy, congestive heart failure, cardiomegaly, and an enlarged spleen and liver. Patient medications included simethicone, Lasix, Digoxin, Colace, Zantac, captopril, and Vasotec.
BRACDIK08	PSPORT1	This amplified and normalized library was constructed using RNA isolated from diseased corpus callosum tissue removed from the brain of a 57-year-old Caucasian male who died from a cerebrovascular accident. Serologies were negative. Patient history included Huntington's disease, emphysema, and tobacco abuse (3-4 packs per day for 40 years).
BRAENOT02	pINCY	Library was constructed using RNA isolated from posterior parietal cortex tissue removed from the brain of a 35-year-old Caucasian male who died from cardiac failure.
BRAINOT09	pINCY	Library was constructed using RNA isolated from brain tissue removed from a Caucasian male fetus, who died at 23 weeks' gestation.
BRAPDIT01	pINCY	Library was constructed using RNA isolated from diseased pons tissue removed from the brain of a 57-year-old Caucasian male, who died from a cerebrovascular accident. Serology was negative. Patient history included Huntington's disease, emphysema, and tobacco abuse.
BRAUTDR02	PCDNA2.1	This random primed library was constructed using RNA isolated from pooled amygdala and entorhinal cortex tissue removed from a 55-year-old Caucasian female who died from cholangiocarcinoma. Pathology indicated mild meningeal fibrosis predominately over the convexities, scattered axonal spheroids in the white matter of the cingulate cortex and the thalamus, and a few scattered neurofibrillary tangles in the entorhinal cortex and the periaqueductal gray region. Pathology for the associated tumor tissue indicated well-differentiated cholangiocarcinoma of the liver with residual or relapsed tumor. Patient history included cholangiocarcinoma, post-operative Budd-Chiari syndrome, biliary ascites, hydrothorax, dehydration, malnutrition, oliguria and acute renal failure. Previous surgeries included cholecystectomy and reserving 68.95, of the liver
BRSTNOT02	PSPORTI	Library was constructed using RNA isolated from diseased breast tissue removed from a 55-year-old Caucasian female during a unilateral extended simple mastectomy. Pathology indicated proliferative fibrocysytic changes characterized by apocrine metaplasia, sclerosing adenosis, cyst formation, and ductal hyperplasia without atypia. Pathology for the associated tumor tissue indicated an invasive grade 4 mammary adenocarcinoma. Patient history included atrial tachycardia and a benign neoplasm. Family history included cardiovascular and cerebrovascular disease.

Table (

Library	Vector	Library Description
BRSTNOT05	PSPORT1	Library was constructed using RNA isolated from breast tissue removed from a 58-year-old Caucasian female during a unilateral extended simple mastectomy. Pathology for the associated tumor tissue indicated multicentric invasive grade 4 lobular carcinoma. Patient history included skin cancer, rheumatic heart disease, osteoarthritis, and tuberculosis. Family history included cerebrovascular and cardiovascular disease, breast and prostate cancer, and type I diabetes.
BRSTNOT09	pincy	Library was constructed using RNA isolated from breast tissue removed from a 45-year-old Caucasian female during unilateral extended simple mastectomy. Pathology for the associated tumor tissue indicated invasive nuclear grade 2-3 adenocarcinoma, with 3 of 23 lymph nodes positive for metastatic disease. Immunostains for estrogen/progesterone receptors were positive, and uninvolved tissue showed proliferative changes. The patient concurrently underwent a total abdominal hysterectomy. Patient history included valvuloplasty of mitral valve without replacement, rheumatic mitral insufficiency, and rheumatic heart disease. Family history included acute myocardial infarction, athenosclerotic coronary artery disease, and the properties.
COLNFET02	pINCY	Library was constructed using RNA isolated from the colon tissue of a Caucasian female fetus, who died at 20 weeks' gestation.
CORPNOT02	pINCY	Library was constructed using RNA isolated from diseased corpus callosum tissue removed from the brain of a 74-year-old Caucasian male who died from Alzheimer's disease.
ENDCNOT03	pINCY	Library was constructed using RNA isolated from dermal microvascular endothelial cells removed from a neonatal Caucasian male.
EOSITXT01	pINCY	Library was constructed using RNA isolated from eosinophils stimulated with IL-5.
FIBPFEN06	pINCY	The normalized prostate stromal fibroblast tissue libraries were constructed from 1.56 million independent clones from a prostate fibroblast library. Starting RNA was made from fibroblasts of prostate stroma removed from a male fetus, who died after 26 weeks' gestation. The libraries were normalized in two rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228 and Bonaldo et al., Genome Research (1996) 6:791, except that a significantly longer (48-hours/round)reannealing hybridization was used. The library was then linearized and recircularized to select for insert containing clones as follows: plasmid DNA was prepped from approximately 1 million clones from the normalized prostate stromal fibroblast tissue libraries following soft agar transformation.
HEARNON08	PBLUESCRIPT	PBLUESCRIPT This normalized, pooled mixed untreated cardiomyocytes and heart tissue library was constructed from 1 million independent clones from a mixed heart and cardiomyocyte library. Starting RNA was made from polyA RNA isolated from pooled untreated cardiomyocytes removed from a 16-week-old Caucasian fetus (donor A) and heart tissue removed from a 21-year-old Caucasian female (donor B) who died from cardiopulmonary arrest. Donor B's history included delivery four months prior to death and seizures. Patient medications included unspecified birth control pills. The library was normalized in 2 rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research 6 (1996):791, except that a significantly longer (48 hours/round) reannealing hybridization was used.

Library	Vector	Library Description
EE02	2.1	This 5' biased random primed library was constructed using RNA isolated from kidney tissue removed from a Caucasian male fetus who was stillborn with a hypoplastic left heart at 23 weeks' gestation. Serology was negative.
KIDNNOC01	pINCY	This large size-fractionated library was constructed using RNA isolated from pooled left and right kidney tissue removed from a Caucasian male fetus, who died from Patau's syndrome (trisomy 13) at 20-weeks' gestation.
KIDNNOT05	PSPORTI	Library was constructed using RNA isolated from the kidney tissue of a 2-day-old Hispanic female, who died from cerebral anoxia. Family history included congenital heart disease.
KIDNNOT32	pINCY	Library was constructed using RNA isolated from kidney tissue removed from a 49-year-old Caucasian male who died from an intracranial hemorrhage and cerebrovascular accident. Patient history included tobacco abuse.
LIVRTMR01	PCDNA2.1	This random primed library was constructed using RNA isolated from liver tissue removed from a 62-year-old Caucasian female during partial hepatectomy and exploratory laparotomy. Pathology for the matched tumor tissue indicated metastatic intermediate grade neuroendocrine carcinoma, consistent with islet cell tumor, forming nodules ranging in size, in the lateral and medial left liver lobe. The pancreas showed fibrosis, chronic inflammation and fat necrosis consistent with pseudocyst. The gallbladder showed mild chronic cholecystitis. Patient history included malignant neoplasm of the pancreas tail, pulmonary embolism, hyperlipidemia, thrombophlebitis, joint pain in multiple joints, type II diabetes, benign hypertension, cerebrovascular disease, and normal delivery. Previous surgeries included distal pancreatectomy, total splenectomy, and partial hepatectomy. Family history included pancreas cancer with secondary liver cancer. Penian hymertension and hymericial
LNODNOT05	pINCY	Library was constructed using RNA isolated from lymph node tissue obtained from a 14-year-old Caucasian female, who died from cardiac arrest secondary to burns. Serology was negative.
LSUBNOT03	pINCY	Library was constructed using RNA isolated from submandibular gland tissue obtained from a 68-year-old Caucasian male during a sialoadenectomy. Family history included acute myocardial infarction, atherosclerotic coronary artery disease, and type II diabetes.
LUNGASTO	PSPORT1	Library was constructed using RNA isolated from the lung tissue of a 17-year-old Caucasian male, who died from head trauma. Patient history included asthma.
LUNGFET05	PSPORT1	Library was constructed using RNA isolated from lung tissue removed from a Caucasian female fetus, who died at 20 weeks' gestation from anencephalus.
LUNGNOT02	PBLUESCRIPT	PBLUESCRIPT Library was constructed using RNA isolated from the lung tissue of a 47-year-old Caucasian male, who died of a subarachnoid hemorrhage.

ı		Library Description
LUNLTMT01	pINCY	The library was constructed using RNA isolated from right middle lobe lung tissue removed from a 63-year-old Caucasian female during a segmental lung resection. Pathology for the associated tumor tissue indicated grade3 adenocarcinoma in the right lower lobe and right middle lobe that infiltrated the parietal pleural surface. Metastatic grade 3 adenocarcinoma was found in the diaphragm. The lymph nodes contained metastatic grade 3 adenocarcinoma and involved the superior mediastinal and inferior mediastinal lymph nodes. Patient history included hyperlipidemia. Family history included benign hypertension, cerebrovascular disease, breast cancer, and hyperlipidemia.
MIXDUNB01	pINCY	Library was constructed using RNA isolated from myometrium removed from a 41-year-old Caucasian female (A) during vaginal hysterectomy with a dilatation and curettage and untreated smooth muscle cells removed from the renal vein of a 57-year-old Caucasian male. Pathology for donor A indicated the myometrium and cervix were unremarkable. The endometrium was secretory and contained fragments of endometrial polyps. Benign endo- and ectocervical mucosa were identified in the endocervix. Pathology for the associated tumor tissue indicated uterine leiomyoma. Medical history included an unspecified menstrual disorder, ventral hernia, normal delivery, a benign ovarian neoplasm, and tobacco abuse in donor A. Previous surgeries included a bilateral destruction of fallopian tubes, removal of a solitary ovary, and an exploratory laparotomy in donor A. Medications included ferrous sulfate in donor A.
MONOTXT02 pINCY	pINCY	The library was constructed using RNA isolated from treated monocytes from peripheral blood removed from a 42-year-old female. The cells were treated with interleukin-10 (IL-10) and lipopolysaccharide (LPS). IL-10 was added at time 0 at 10 ng/ml, LPS was added at 1 hour at 5 ng/ml. The monocytes were isolated from buffy coat by adherence to plastic. Incubation time was 24 hours.
	PSPORTI	Library was constructed using RNA isolated from adherent mononuclear cells, which came from a pool of male and female donors. The cells were stimulated with LPS.
MPHGNOT03	PBLUESCRIPT	PBLUESCRIPT Library was constructed using RNA isolated from plastic adherent mononuclear cells isolated from buffy coat units obtained from unrelated male and female donors.
MYOMNOT01 PSPORT1	PSPORT1	Library was constructed using RNA isolated from uterine myometrial tissue removed from a 43-year-old Caucasian female during a vaginal hysterectomy and removal of the fallopian tubes and ovaries. Family history included lung cancer, stroke, type II diabetes, hepatic lesion, chronic liver disease, hyperlipidemia, congenital heart anomaly, and mitral valve prolanse.

Table (

Library	Vector	Library Description
DNO3	pINCY	This normalized dorsal root ganglion tissue library was constructed from 1.05 million independent clones from a dorsal root ganglion tissue library. Starting RNA was made from dorsal root ganglion tissue removed from the cervical spine of a 32-year-old Caucasian male who died from acute pulmonary edema, acute bronchopneumonia, bilateral pleural effusions, pericardial effusion, and malignant lymphoma (natural killer cell type). The patient presented with pyrexia of unknown origin, malaise, fatigue, and gastrointestinal bleeding. Patient history included probable cytomegalovirus infection, liver congestion, and steatosis, splenomegaly, hemorrhagic cystitis, thyroid hemorrhage, respiratory failure, pneumonia of the left lung, natural killer cell lymphoma of the pharynx, Bell's palsy, and tobacco and alcohol abuse. Previous surgeries included colonoscopy, closed colon biopsy, adenotonsillectomy, and nasopharyngeal endoscopy and biopsy. Patient medications included Diflucan (fluconazole), Deltasone (prednisone), hydrocodone, Lortab, Alprazolam, Reazodone, ProMace-Cytabom, Etoposide, Cisplatin, Cytarabine, and dexamethasone. The patient received radiation therapy and multiple blood transfusions. The library was normalized in 2 rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research 6 (1996):791, except that a significantly longer (48 hours/round) reannealing hybridization was used.
	PSPORTI	PSPORT1 Library was constructed using RNA isolated from tumorous neuroganglion tissue removed from a 9-year-old Caucasian male during a soft tissue excision of the chest wall. Pathology indicated a ganglioneuroma. Family history included asthma.
PANCNOT01	PBLUESCRIPT	Library was constructed using RNA isolated from the pancreatic tissue of a 29-year-old Caucasian male who died from head trauma.
PENITUTO	pINCY	Library was constructed using RNA isolated from tumor tissue removed from the penis of a 64-year-old Caucasian male during penile amputation. Pathology indicated a fungating invasive grade 4 squamous cell carcinoma involving the inner wall of the foreskin and extending onto the glans penis. Patient history included benign neoplasm of the large bowel, atherosclerotic coronary artery disease, angina pectoris, gout, and obesity. Family history included malignant pharyngeal neoplasm, chronic lymphocytic leukemia, and chronic liver disease.
PROSDITO	pINCY	The library was constructed using RNA isolated from diseased prostate tissue removed from a 58-year-old Caucasian male during radical prostatectomy, regional lymph node excision, and prostate needle biopsy. Pathology indicated adenofibromatous hyperplasia. Pathology for the associated tumor tissue indicated adenocarcinoma Gleason grade 3+3, which formed a predominant mass involving the right posterior superior prostate. Another microscopic focus of tumor was identified in the left posterior inferior. The tumor invaded the capsule but didnot extend beyond it. The patient presented with elevated prostate specific antigen (PSA), nocturia, hematuria, and induration. Patient history included benign hypertension. Family history included benign hypertension and prostate cancer.

7 :1.		
Liorary	Vector	Library Description
PROSNOT28	pINCY	Library was constructed using RNA isolated from diseased prostate tissue removed from a 55-year-old Caucasian male during
		a radical prostatectomy and regional lymph node excision. Pathology indicated adenofibromatous hyperplasia. Pathology for
		the associated tumor tissue indicated adenocarcinoma, Gleason grade 5+4. The patient presented with elevated prostate
		specific antigen (PSA). Family history included lung and breast cancer.
SCOMDIC01	PSPORT1	This large size-fractionated library was constructed using RNA isolated from diseased spinal cord tissue removed from the
		base of the medulla of a 57-year-old Caucasian male, who died from a cerebrovascular accident. Serologies were negative.
		Patient history included Huntington's disease, emphysema, and tobacco abuse (3-4 packs per day, for 40 years)
SINTFER02	pINCY	This random primed library was constructed using RNA isolated from small intestine tissue removed from a Caucasian male
		fetus who died from fetal demise.
SINTNOROI	PCDNA2.1	This random primed library was constructed using RNA isolated from small intestine tissue removed from a 31-year-old
		Caucasian female during Roux-en-Y gastric bypass. Patient history included clinical obesity.
SPLNDIC01	pINCY	This large size-fractionated library was constructed using pooled cDNA from two different donors. cDNA was generated using
		mRNA isolated from spleen tissue removed from an 8-year-old Black male (donor A) who died from anoxia and from diseased
		spleen tissue removed from a 14-year-old Asian male (donor B) during a total splenectomy. Pathology for donor B indicated
		changes consistent with idiopathic thrombocytopenic purpura. Serologies were negative for donor A. Donor B presented with
		bruising. Patient medications included DDAVP, Versed, labetalol (donor A), and Vincristine (donor B).
SYNORABOI	PBLUESCRIPT	SYNORAB01 PBLUESCRIPT Library was constructed using RNA isolated from the synovial membrane tissue of a 68-year-old Caucasian female with
		rheumatoid arthritis.

Program	Description	Reference	Parameter Threshold
ABIFACTURA	hat removes vector sequences and masks bases in nucleic acid sequences.		
ABI/PARACEL FDF	A Fast Data Finder useful in comparing and annotating amino acid or nucleic acid sequences.	Applied Biosystems, Foster City, CA; Paracel Inc., Pasadena, CA.	Mismatch <50%
ABI AutoAssembler	A program that assembles nucleic acid sequences.	Applied Biosystems, Foster City, CA.	
BLAST	A Basic Local Alignment Search Tool useful in	Altschul, S.F. et al. (1990) J. Mol. Biol.	ESTs: Probability value = 1.0E-
	sequence similarity search for amino acid and nucleic 215:403-410; Altschul, S.F. et al. (1997)	215:403-410; Altschul, S.F. et al. (1997)	8 or less; Full Length sequences:
	acid sequences. BLAST includes five functions: blastp, blastn, blastx, tblastn, and tblastx.	Nucleic Acids Res. 25:3389-3402.	Probability value = 1.0E-10 or less
FASTA	A Pearson and Lipman algorithm that searches for	Pearson, W.R. and D.J. Lipman (1988) Proc. ESTs: fasta E value = 1.06E-6;	ESTs: fasta E value = 1.06E-6;
	similarity between a query sequence and a group of	Natl. Acad Sci. USA 85:2444-2448; Pearson, Assembled ESTs: fasta Identity	Assembled ESTs: fasta Identity
	sequences of the same type. FASTA comprises as	W.R. (1990) Methods Enzymol. 183:63-98;	= 95% or greater and Match
	least five functions: fasta, tfasta, fastx, tfastx, and	and Smith, T.F. and M.S. Waterman (1981)	length = 200 bases or greater;
	ssearch.	Adv. Appl. Math. 2:482-489.	fastx E value = 1.0E-8 or less;
			Full Length sequences: fastx
			score = 100 or greater
BLIMPS	A BLocks IMProved Searcher that matches a	Henikoff, S. and J.G. Henikoff (1991)	Probability value = 1.0E-3 or
	sequence against those in BLOCKS, PRINTS,	enikoff,	
		J.G. and S. Henikoff (1996) Methods	
	quence homology, and structural	Enzymol. 266:88-105; and Attwood, T.K. et	
	fingerprint regions.	al. (1997) J. Chem. Inf. Comput. Sci. 37:417-	
HMMER	rching a query sequence against	Krogh, A. et al. (1994) J. Mol. Biol.	PFAM, INCY, SMART or
	hidden Markov model (HMM)-based databases of	235:1501-1531; Sonnhammer, E.L.L. et al.	TIGRFAM hits: Probability
	protein family consensus sequences, such as PFAM,	(1988) Nucleic Acids Res. 26:320-322;	value = 1.0E-3 or less; Signal
	INCY, SMART and TIGRFAM.	Durbin, R. et al. (1998) Our World View, in	peptide hits: Score = 0 or greater
		a Nutshell, Cambridge Univ. Press, pp. 1-	
ProfileScan	An algorithm that searches for structural and	Gribskov, M. et al. (1988) CABIOS 4:61-66; Normalized quality score ≥ GCG	Normalized quality score ≥ GCG
	sequence motifs in protein sequences that match	Gribskov, M. et al. (1989) Methods	specified "HIGH" value for that
	sequence patterns defined in Prosite.	Enzymol. 183:146-159; Bairoch, A. et al.	particular Prosite motif.
		(1997) Nucleic Acids Res. 25:217-221.	Generally, score = 1.4-2.1.

Program	Description	Reference	Parameter Threshold
Phred	A base-calling algorithm that examines automated	Ewing, B. et al. (1998) Genome Res. 8:175-	
	sequencer traces with high sensitivity and probability. 185; Ewing, B. and P. Green (1998) Genome Res. 8:186,194	185; Ewing, B. and P. Green (1998) Genome Res. 8:186-194	
Phrap	A Phils Revised Assembly Program including	Smith, T.F. and M.S. Waterman (1981) Adv. Score = 120 or greater: Match	Score = 120 or greater: Match
	SWAT and CrossMatch, programs based on efficient Appl. Math. 2:482-489; Smith, T.F. and	Appl. Math. 2:482-489; Smith, T.F. and	length = 56 or greater
	implementation of the Smith-Waterman algorithm,	M.S. Waterman (1981) J. Mol. Biol. 147:195-	
	useful in searching sequence homology and	197; and Green, P., University of	
	assembling DNA sequences.	Washington, Seattle, WA.	
Consed	A graphical tool for viewing and editing Phrap	Gordon, D. et al. (1998) Genome Res. 8:195-	
		202.	
SPScan	A weight matrix analysis program that scans protein	Nielson, H. et al. (1997) Protein Engineering Score = 3.5 or greater	Score = 3.5 or greater
	sequences for the presence of secretory signal	10:1-6; Claverie, J.M. and S. Audic (1997)	
	peptides.	CABIOS 12:431-439.	
TMAP	A program that uses weight matrices to delineate	Persson, B. and P. Argos (1994) J. Mol. Biol.	
	transmembrane segments on protein sequences and	237:182-192; Persson, B. and P. Argos	
	determine orientation.	(1996) Protein Sci. 5:363-371.	
TMHMMER	A program that uses a hidden Markov model (HMM) Sonnhammer, E.L. et al. (1998) Proc. Sixth	Sonnhammer, E.L. et al. (1998) Proc. Sixth	
	to delineate transmembrane segments on protein	Intl. Conf. On Intelligent Systems for Mol.	
	sequences and determine orientation.	Biol., Glasgow et al., eds., The Am. Assoc.	
		for Artificial Intelligence (AAAI) Press,	
		Menlo Park, CA, and MIT Press, Cambridge,	
		MA, pp. 175-182.	
Motifs	A program that searches amino acid sequences for	Bairoch, A. et al. (1997) Nucleic Acids Res.	
	patterns that matched those defined in Prosite.	25:217-221; Wisconsin Package Program	
		Manual, version 9, page M51-59, Genetics	
		Computer Group, Madison, WI.	_

Table 8

SEQ PID	ESTID	OI ANS	EST	CB1	EST	Allele		Allele Amino Acid	Caucasian	African	Asian	Hispanic
			SNP	SK BYS	Allele	-	7		Allele 1	Allele 1	Allele 1	Allele 1
									frequency	frequency	frequency	frequency
7506904	1700625H1	SNP00039444	77	1249	▼	A	Ð	noncoding	p/u	n/a	n/a	n/a
7506904	1700625H1	SNP00072668	152	1324	A	Ą	Đ	noncoding	p/u	n/a	r/a	n/a
7506904	6331607H1	SNP00039444	138	1239	٧	A	Ð	noncoding	p/u	n/a	n/a	n/a
7506904	6331607H1	SNP00072668	213	1314	ß		ຽ	Γ		n/a	n/a	n/a
7506909	1700625H1	SNP00039444	77	947	V		Ð		n/d	n/a	n/a	n/a
7506909	1700625H1	SNP00072668	152	1022	٧	A	Ð	noncoding	p/u	n/a	n/a	n/a
7506909	6331607H1	SNP00039444	138	937	Y	A	Ŋ	noncoding	p/u	ī/a	n/a	n/a
7506909	6331607H1	SNP00072668	213	1012				Π	p/u	n/a	n/a	n/a
7507096	1560345H1	SNP00047855	58	1755	Ţ	Т		l	0.59		4	0.69
7507096	3213196H1	SNP00129679	16	1008				noncoding	n/a	n/a		n/a
7507098	1560345H1	SNP00047855	58	1900	1	T						69.0
7507098	3213196H1	SNP00129679	16	1153	T		T	l "	n/a			n/a
7507099	1560345H1		58	1677				noncoding	0.59	0.61	_	0.69
7507099	3213196H1	SNP00129679	16	930	T	C		١,				n/a
7501399	1273836H1	SNP00100328	155	1137		G	A	noncoding	n/a	n/a	n/a	n/a
7501399	3744343H1	SNP00100327	9	18				noncoding	n/a			n/a
7501399	3744344H1	SNP00100327	7	16	၁	C	Ţ		n/a			n/a
7501399	4784901H2	SNP00068331	206	1687					n/a			n/a
7501399	5353388H1	SNP00068331	118	1688				noncoding	n/a	n/a		n/a
7501399	6404778H1	SNP00068330	29	1310	C	T	C	Г	0.12			n/a
7501399	6869706H1	SNP00128804	§	1197		G		noncoding	n/a	n/a		n/a
7501399	6869706H1	SNP00068330	488	1313	L	T		noncoding	0.12			n/a
7504768	1343743H1	SNP00052446	160	391		G		Г	n/a			n/a
7504768	1343743H1	SNP00067987	200	431		၁	Ţ	Γ	n/a			n/a
7504768	1343743H1	SNP00068446	107	338		Ö			0.99	p/u		p/u
7504768	1343743H1	SNP00075667	114	345		၁	T					n/a
7504768	1343743H1	SNP00132838	56	287	၁	ر		noncoding	n/a	n/a		n/a
7504768	1343757H1		100	644				Т				1/a
7504768	1343782H1	SNP00063753	28	24	ر د	် ပ	4	P4				7/8

icy frequency n/d
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187
SNP00056531 18
412
134444H1 SN

Table {

PD	ESTID	SNPID	EST	CB1	EST	Allele	Allele	Allele Amino Acid	Caucasian	African	Asian	Hispanic
	_		SNP	SNP	Allele	-	8		Allele 1	Allele 1	Allele 1	Allele 1
									frequency	frequency	frequency	frequency
7504768	1549314H1	SNP00075667	179	355	၁	င	Ţ	noncoding	n∕a	Za	n/a	n/a
7504768	1549314H1	SNP00132838	121	297	ပ	C	Ţ	noncoding	n/a	n/a	n/a	n/a
7504768	1549314H1	SNP00140832	46	222	C	င	A	noncoding	n/a	n/a	n/a	n/a
7504768	1549323H1	SNP00011894	182	718	၁	၁	Ţ	noncoding	n/a	n/a	n/a	n/a
7504768	1549323H1	SNP00129147	163	669	၁	C	T	noncoding	n/a	n/a	n/a	n/a
7504768	1549587H1	SNP00063754	25	194	ß	G	၁	noncoding	n/a	n/a	n/a	n/a
7504768	1549587H1	SNP00076561	24	126		Ð		A38	n/a	n/a	n/a	n/a
7504768	1549587H1	SNP00140832	119	221	ر	င	A	noncoding	n/a	n/a	n/a	n/a
7504768	1549671H1	SNP00056531	196	630			¥	noncoding	n/d	n/a	n/a	n/a
7504768	1682001H1	SNP00042307	30	314		ပ	A	noncoding	0.20	0.07	0.08	0.41
7504768	1682982H1	SNP00037414	29	530			Ţ	noncoding	n/a	n/a	n/a	n/a
7504768	1683569H1	SNP00076561	115	911			C	T34	n/a	n/a	n/a	n/a
7504768	1683572H1	SNP00047368	26	63	ر ت		Ţ	417	n/a	n/a	n/a	n/a
7504768	1683572H1	SNP00046952	63	72				A20	n/a	n/a	n/a	n/a
7504768	1685714H1	SNP00044426	103	244	A	G	A	noncoding	p/u	P/a	p/u	p/u
7504768	1686274H1	SNP00046952	75	71				A19	n/a	r/a	n/a	n/a
7504768	1687168H1	SNP00054365	25	561			င	noncoding	n/a	n/a	n/a	n/a
7504768	1688106H1	SNP00052446	77	410			A	noncoding	n/a	n/a	n/a	n/a
7504768	1688106H1	SNP00067987	117	449	C	င	Ţ	noncoding	n/a	n/a	n/a	n/a
7504768	1688106H1	SNP00068446	8	357			Ą	noncoding	0.99	n/d	p/u	p/u
7504768	1688106H1	SNP00075667	31	364	ပ	C	T	noncoding	n/a	n/a	n/a	n/a
7504768	1688329H1	SNP00011894	8	719			T	noncoding	n/a	n/a	n/a	n/a
7504768	1690353H1	SNP00155250	88	373	၁	Č	A	noncoding	n/a	n/a	n/a	n/a
7504768	1690881H1	SNP00044426	226	225	A	ß	A	noncoding	p/u	p/u	p/u	p/u
7504768	1709318H1	SNP00056531	188	632			A	noncoding	ργ	n/a	n/a	n/a
7504768	1711676H1	SNP00046952	73	73	Ð	A	G	G20	n/a	n/a	n/a	n/a
7504768	1724316H1	SNP00044426	54	977			A	Γ	p/u	p/u	p/u	וי/מ
7504768	1725201H1	SNP00054416	70	70	Ţ	T		617	n/a	n/a	n/a	r/a
7504768	1808532H1	SNP00042307	179	185) ၁	C	A	noncoding	0.20	0.07	0.08	0.41

Table 8

			_				Ψ.	_	_	_	_		, .	,		,		_				_		_	_	_	-				
Hispanic	Allele 1	frequency	n/a	n/a	n/đ	n/a	n/a	n/a	n/a	p/u	n/a	n/a	n/d	n/a	0.03	n/a	0.03	n/a	p/u	n/a	n/a	n/d	n/a	n/a	n/a	n/a	n/a	p/u	īva	n/a	n/a
Asian	Allele 1	frequency	n/a	n/a	1/q	n/a	n/a	n/a	n/a	p/u	n/a	n/a	p/u	n/a	0.04	n/a	0.04	n/a	p/u	n/a	n/a	p/u	n/a	n/a	n/a	n/a	n/a	p/u	n/a	n/a	
African	Allele 1	frequency	n/a	n/a	p/u	n/a	n/a	n/a	n/a	p/a	п/а	n/a	p/u	n/a	0.04	n/a	90.0	n/a	n/d	n/a	n/a	p/u	n/a	n/a	n/a	n/a	n/a	p/u	n/a	n/a	n/a
Caucasian	Allele 1	frequency	n/a	n/a	p/u	n/a	n/a	n/a	n/a	0.99	n/a	n/a	n/d	n/a	90.0		90'0	n/a	n/d	1/a	n/a	0.99	n/a	Zå	n/a		n/a	0.99	n/a	n/a	n/a
Allele Amino Acid			V21	V12	noncoding	noncoding	noncoding	noncoding	noncoding	noncoding	noncoding	R16	noncoding	noncoding	V3	W14	noncoding	SII	noncoding	П	noncoding	noncoding		L22	W14	noncoding		noncoding			Г
Allele	7			၁	A				T	A		1	A	T			၁	A		A	A			Ð		A	T		T		Ö
Allele	_			Ţ		ß	Ü	Ŋ	ر ن			ິບ				ß		ß		C	G	ט			T	Ð	C	ß	၁		V
EST	Allele		G			G	ں ت		၁			ပ ပ				ß				၁			ر د) D		G	ပ ၁		ر ق
GB.	SNP		75	49		390	430	386	426		0			0		56				25	389		343		54	400	440	347		296	06
EST	SNP		. 9/	53	92	115	155	98	126	33			272	172	39	72	17	20	152	35	89		22		29	179	219	126	9	75	16
SNP ID		- 1	SNP00046952			SNP00052446	SNP00067987	SNP00052446	SNP00067987	_	- 1			SNP00129147			SNP00046892		SNP00044426		- 1	\neg	SNP00075667		SNP00054416	SNP00052446		SNP00068446		\neg	SNP00046952
ESTID			1809090H1	_	_	1809992H1	1809992H1	1810488H1		1810488H1													1914285H1		1915830H1						1917215H1
PID			7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768
SEO	A	히	\neg		П	58	88	88		丁	\neg	コ	28		٦	コ	\neg			П	ヿ	T		ヿ			\neg	T		T	28

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Hispanic	Allele 1	frequency	n/a	n/a	17/a	n/a	n/a	p/u	n/a																						
Asian	Allele 1	frequency	n/a	n/a	n/a	ī/a	n/a	n/a	p/u	n/a																					
African	Allele 1	frequency	n/a	n/a	p/u	n/a	n/a	n/a	π/a	n/a	n/a	n/a	n/a	n/a	n/a																
Caucasian	Allele 1	frequency	n/a	p/u	n/a	n/a	0.99	n/a																							
Amino Acid			H17	nohcoding	noncoding	noncoding	noncoding	S7	noncoding	noncoding	87	noncoding	A3	V3	1.3	noncoding	115	noncoding	noncoding	noncoding	noncoding	noncoding	noncoding	noncoding	A34	noncoding	noncoding	noncoding	Tı	noncoding	noncoding
Allele	7		ပ		Ţ	T	T	A		T	A	A		A			T		A			T			, ၁	_			Α.	A	
Allele	-		T	ß	ວ	C	C	C		Ċ	၁	C	C	C	၁		C	င	G	င			၁	G		၁	င	C	C		C
EST	Allele		Ţ	Ŋ	C	C	၁	C	G	C	င	ပ	c	င	C	C	T	င	G	ပ	G	င	C	G		C	ပ	ပ	ပ		ပ
GB	SNP		જ	586	720	701	643	34	415	455	36	653	22	23	21	14	59	<u>2</u>	387	427	334	341	284	182	114	509	429	285	16	382	422
EST	SNP		89	36	91	72	36	35	29	69	37	188	23	33	24	17	35	33	161	201	108	115	27	181	113	208	163	19	18	22	જ
SNP ID			SNP00054416	SNP00054365	SNP00011894	SNP00129147	SNP00068474	SNP00063753	SNP00052446	SNP00067987	SNP00063753	SNP00056531	SNP00063753	SNP00063753	SNP00063753	SNP00063753	SNP00047368	SNP00068474	SNP00052446 161	SNP00067987	SNP00068446	SNP00075667	SNP00132838	SNP00063754	SNP00076561	SNP00140832	SNP00067987	SNP00132838	SNP00063753	SNP00052446	SNP00067987
ESTID			1917215H1	2068818H1	2187806H1	2187806H1	2233404H1	2271117H1	2276979H1	2276979H1	2388554H1	2941522H1	3272323H1	3272538H1	3272748H1	3272754H1	3272891H1	3273142H1	3273256H1	3273256H1	3273256H1	3273256H1	3273256H1	3276649H1	3276649H1	3276649H1	3276751H1	3276751H1	3276756H1	3277293H1	3277293Н1
PID			7504768	7504768	7504768	7504768	7504768		7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768
SEQ	А	Ö		╗	\neg		. 28	28			28	П	コ			\neg		88	T			Ì	Ì				\exists	Ì	1	T	28

ESTD	 	SNP ID	EST	ig B	EST	Allele	Allele	Allele Amino Acid	Caucasian	African	Asian	Hispanic
			SNP	SNP	Allele		~		Allele 1	Allele 1	Allele 1	Allele 1
- 1	+								frequency	frequency	frequency	frequency
3316682HI	છ	SNP00009321	44	455	၁	Ţ	၁	noncoding	0.92	n/a	n/a	n/a
3316682H		SNP00037413	15	426	g	Ð	A	noncoding	n/a	n/a	11/a	n/a
		SNP00068446	123	337	G	G		noncoding	0.99	p/u	p/u	n/d
:17:1	3317420H1 SI	SNP00075667	130	344	ت ن	ပ	F	Г	n/a	n/a	n/a	n/a
361	3317420H1 SI	SNP00132838	72	286				Г	n/a	n/a	n/a	n/a
321	3317786H1 ST	SNP00063753	23	20	၁	Ü		Г	n/a	n/a	n/a	n/a
Œ	_	SNP00009321	45	452		F.		noncoding	0.92	n/a	n/a	n/a
3318147H		SNP00037413	16	423	G	ß			n/a	n/a	n/a	n/a
平		SNP00063833	140	133		G	S	G40	p/u	P/a	n/d	p/u
3351152H		SNP00054416	41					L10	n/a	n/a	n/a	n/a
3356872H		SNP00063754	54	184		ß	Г	noncoding	n/a	n/a		n/a
3356872H		SNP00140832		211				Г	n/a	n/a		n/a
3358153HI				19				Г	n/a	n/a		n/a
3358196Н				57	T	C .	T	FIS	n/a	r/a		n/a
3359959H			0	,		, G		noncoding	0.99	p/u		μ/d
3360072H							T	noncoding	ī√a	n/a	n/a	ī⁄a
3360072H		SNP00129147	29	9		ပ			n/a	n/a	n/a	n/a
3361360H			82	70	Ü		Ŋ	G19	n/a	n/a	n/a	n/a
3362266Н				76				G21	n/a	n/a	n/a	n/a
3362266Н	_						C		n/a	n/a		n/a
3362413H	_	SNP00047368	75	65		ິ		H17	n/a	n/a	n/a	n/a
3362834H			4	529	၁	C		noncoding	n/a	n/a	n/a	n/a
3362834H		- 1	8	593		Ð	၁	noncoding	n/a	n/a	n/a	n/a
3365106H	_	SNP00140832	4	210		' ၁	¥			n/a	n/a	n/a
3497647H		SNP00068474	85	642					n/a	n/a		n/a
3498973H			5	59	T	T (၁		n/a	n/a		n/a
3499611H	2							noncoding	n/a	n/a		n/a
3962030H		SNP00146294	137	277	ပ ပ	S	Ð					n/a
3962418H	<u>~</u>	NP00146294	11	279	၁	၁	Ð	П	n/a			n/a

		_	_	_	_	_	,	_	_	_	_	_	_	_	_	-	_	_	_		_	_	_	_	_	_	_	_	_		
Hispanic	Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	0.41	n/a	n/a	n/a	n/a	n/a	n/d	p/u	n/a	n/a	n/a	n/a	n/a	17/8	n/a	n/a	p/u	n/a	n/a	n/a	n/a	n/a	n/a	n/a
Asian	Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	0.08	n/a	n/a	n/a	n/a	n/a	p/u	p/u	n/a	r/a	p/u	n/a	n/a	n/a	n/a	n/a								
African	Allele 1	frequency	17/3	n/a	n/a	n/a	n/a	0.07	n/a	n/a	n/a	n/a	n/a	n/d	n/d	n/a	p/u	n/a	n/a	n/a	n/a	n/a	n/a								
Caucasian	Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	0.20	n/a	n/a	n/a	n/a	n/a	0.99	p/u	n/a	0.99	n/a	0.92	n/a	n/a	n/a	n/a	n/a							
Allele Amino Acid			noncoding	S34	noncoding	noncoding	noncoding	noncoding	115	noncoding	noncoding	R33	noncoding		noncoding	MI3	A13	noncoding	noncoding												
Allele	7		C	ر	T	4	Ţ	A		Ţ	Ţ	U	Ð	V		T		T	1		S		T	A	T	ر ت	H	_ ပ	V	υ υ	T
Allele	-		G	G	ပ	Ð	C	ပ	Т	၁	င	G	၁	D		င	၁	၁	ပ	ပ	G		C	G	ß	Ţ	Ð	T		Ð	C
EST	Allele		D D	Ð	၁	Ð	ာ	၁	L	T	၁	g		Ð		C	C	T	၁			G	င	G	G	T	Ö	Ţ	G	G	၁
CB1	SNP		183	115	236	388	428	312	58	716	697	112	278	335	235	527	371	726	707	612	646	361	401	309	909	454	602	52	51	283	342
EST	SNP		184	116	58	74	114	39	61	82	63	50	131	62	104	53	173	210	191	95	129	8	130	37	18	46	5	99	55	45	5
SNP ID			SNP00063754	SNP00076561	SNP00132838	SNP00052446	SNP00067987	SNP00042307	SNP00054416	SNP00011894	SNP00129147	SNP00076561	SNP00146294	SNP00068446	SNP00044426	SNP00037414	SNP00155250	SNP00011894	SNP00129147	SNP00037414	SNP00054365	SNP00052446	SNP00067987	SNP00068446	SNP00043769	SNP00009321	SNP00043769			SNP00054365	SNP00075667
ESTID			3962841HI	3962841H1	3963802H1	3964102H1	3964102H1	3964673H1	3964682H1	3965811H1	3965811H1	3966081H1	3966329H1	3966939H1	3967020H1	3967122H1	3967837H1	3968413H1	3968413H1	3968755H1	3968755H1	3978889H1	3978889H1	3978889H1	_	4109074H1	4109695H1	_		4111048H1	4111426H1
Eg.			7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768
SEQ	A	Ö	28	28	28	28	28	28	58	88	58			58	28	28	88	88	28	88	28	28	28	28	88	28	28	28	88	٦	28

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Hispanic	Allele 1	frequency	n/a	n/a	n/a	n/a	ī/a	n/a	0.41	n/a	n/a	17/a	n/a	n/a	n/a	n/a	p/d	n/a	1/م	n/a	n/a	7/a	0.03	n/a	n/a	0.03	n/a	n/a	n/a	n/a	1/a
Asian	Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	n/a	0.08	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/d	n/a	υ/d	n/a	n/a	n/a	0.04	n/a	n/a	0.04	n/a	r/a	n/a	n/a	n/a
African	Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	n/a	0.07	n/a	r/a	n/a	n/a	n/a	n/a	n/a	n/d	n/a	p/u	n/a	17/a	n/a	0.04	n/a	n/a	0.04	n/a	n/a	n/a		
Caucasian	Allele 1	frequency	n/a	0.92	n/a	n/a	n/a	n/a	0.20	n/a	n/a	n/a			n/a	n/a	n/d	n/a		n/a			90.0	n/a		0.06	n/a				n/a
Amino Acid			C14	noncoding	noncoding	R14	F22	noncoding	noncoding	S13	115	noncoding	noncoding	1	S27	noncoding	noncoding	noncoding	A40	noncoding	-		A3	W14	noncoding		G14	noncoding	Π	819	noncoding
Allele	7		T	U		F	U	4				L			Ð		4			F		T	C		T		4		O	ပ	Ü
Allele	-		ر د	Į.		ပ			U							ပ			r D			် ပ				H		Ö	A		U
EST	Allele		ت ن	Ţ		ر د			ပ ပ		Ţ	၁		G	ß	C			ß		ß	ပ	ບ	G	ں ت	ပ	Ü	Ü	י פ	T	၁
CBI	SNP		26	453	370	54		372	310	51	58	346	535	869	94	640		137	134	533	267	2	22	55	543		54	5	93		257
EST	SS		9	6	200	99	83	93	34	59	61	157	140	174	78	28	81	183	6	88	122	230	22	28	101	29	79	22	6	74 (3
SNP ID		7	SNP00047368	SNP00009321	SNP00155250	SNP00047368	SNP00054416	SNP00155250	SNP00042307	SNP00054416	SNP00047368			_	SNP00046952		SNP00044426	SNP00076561		SNP00037414	SNP00054365	\neg	SNP00046892	-	SNP00037414	SNP00046892	SNP00124735	SNP00043769			SNP00146294
ESTID			4111558H1	4111717H1	4112155H1	_	4113023H1	4276507H1	4278118H1	4404036H1											4595478H1										4646632H1
E G			7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768
SEO	<u>A</u>	ÿ	28	28	8	28	88	58	88	28	8	28	28	28	28	28	28	8	28	8	88	28	28	28	28	58	28	% %	28	28	58

Table 8

Uicacia	Allele 1	frequency																													
Ľ	-		√ 2	n/a	n/a	/S	n/a	2	2/a	2	1/a	1/a	\rac{1}{a}	2	1/a	n/a	n/a	4	2/2	2 2	월 물 물	5 8 8 5	2/2 8/2/ B/2/ B/2/ B/2/ B/2/ B/2/ B/2/ B	2/8 2/2 2/8 2/2 2/8 2/9 2/9 2/9 2/9 2/9 2/9 2/9 2/9 2/9 2/9	2/2	7 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2	7/8 2/4 2/4 8/2 8/2 8/2 8/2 8/2 8/2 8/2 8/2 8/2 8/2	7 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2	7/8 2/9 2/9 2/9 2/9 2/9 2/9 2/9 2/9 2/9 2/9	1/8 1/8 1/9 1/9 1/9 1/9 1/9 1/9 1/9 1/9 1/9 1/9	7/8
Acion	Allele 1	frequency	n∕a	n/a	ī⁄a	n/a	n/a	n/a	n/a	n/a	n/a	p/u		n/a	n/a n/a	n/a n/a n/d	n/a n/a n/d n/a	n/a n/a n/d n/a n/a	178 178 179 178 178 179	n/a n/d n/a n/a n/a	1/8 1/8 1/8 1/8 1/8 1/8 1/8 1/8	n/a n/a n/a n/a n/a n/a n/a n/a n/a	10/8 10/4 10/8 10/8 10/8 10/8 10/8 10/8	10/8 10/4 10/8 10/8 10/8 10/8 10/8 10/8 10/8 10/8	10/8 10/4 10/8 10/8 10/8 10/8 10/8 10/8 10/8						
African	Allele 1	frequency	n/a	n/a	n/a	n/a	17/8	n/a	17/8	n/a	D/U		n/a	n/a n/a	n/a n/a n/d	n/a n/a n/d n/a	n/a n/a n/a n/a	1/8 17/8 17/8 17/8 17/8	n/a n/a n/d n/d n/d	n/a n/a n/a n/a n/a n/a	n/a n/a n/d n/a n/a n/a n/a	n/a n/a n/a n/a n/a n/a n/a	n/a n/a n/a n/a n/a n/a n/a n/a	n/a n/a n/a n/a n/a n/a n/a n/a							
Caucasian		frequency	n/a	n/a	n/a	n/a	n/a	p/u	n/a	p/a	7,1	28	n/a	n/a n/d	n/a n/d n/a	n/a n/d n/a n/a	n/a n/d n/a n/a n/a	17/8 17/8 17/8 17/8 17/8	11/8 11/8 11/8 11/8 11/8 11/8	17/8 17/8 17/8 17/8 17/8 17/8 17/8	17/8 17/8 17/8 17/8 17/8 17/8 17/8	11/8 11/8 11/8 11/8 11/8 11/8 11/8 11/8	11/8 11/8 11/8 11/8 11/8 11/8 11/8 11/8								
Amino Acid		.,,,,	noncoding	noncoding	G35	noncoding	noncoding	noncoding	noncoding	r.	noncoding	noncoding	noncoding	noncoding	noncoding	L10	R2	A40	820	1	F22	F22 noncoding	F22 noncoding noncoding	F22 noncoding noncoding	F22 noncoding noncoding noncoding	F22 noncoding noncoding noncoding noncoding	F22 noncoding noncoding noncoding noncoding C31 C24	F22 noncoding noncoding noncoding noncoding C31 C24 noncoding	F22 noncoding noncoding noncoding noncoding O31 C24 noncoding	F22 noncoding noncoding noncoding noncoding Q31 C24 noncoding noncoding	F22 noncoding noncoding noncoding Q31 C24 noncoding noncoding noncoding
Allele	7)	A	ပ		4	4	4	4	¥	L	၁	H	ပ	Ŧ			ပ				ນ ∢									
Ailele		•	ပ	g	g	ပ	Ö	ပ	ပ	၁	C	G		G		T	၁		A		ī	Ţ	F 6 0	G G T	F 0 0 0 0						
EST	Allele		ပ	G	Ð	ပ	უ	ပ ပ	ပ	၁	ပ	G	ပ	ð			၁														
CBI	SNP		13	981	118	213	425		368	15	542	929	553	587	703	42	18	132	101		78	,	VO ++	V2 ++ 10	V # V =	VO 77 10 - 1 -	× × ×	V2 77 15 - 15 15	N # 10 - 1 N 0		
EST	SNP		19	209	141	236	28		233	18	95	129			221		25	128	109												
SNPID			SNP00063753	SNP00063754	SNP00076561	SNP00140832	SNP00037413	SNP00056531	SNP00155250	SNP00063753	SNP00037414	SNP00054365	SNP00037414	SNP00054365	SNP00129147	SNP00054416	SNP00063753	SNP00063833	SNP00046952	7.11.2000CL	SNP00054416	SNP00054416 SNP00044426	SNP00054416 SNP0004426 SNP00155250	SNP00054416 SNP0004426 SNP00155250 SNP00155250	SNP00054416 SNP0004426 SNP00155250 SNP00155250 SNP00155250	SNP00054416 SNP0004426 SNP00155250 SNP00155250 SNP0004426 SNP0004426	SNP00054416 SNP0004426 SNP00155250 SNP00155250 SNP0004426 SNP00046952 SNP00054416	SNP00054416 SNP0004426 SNP00155250 SNP00155250 SNP0004426 SNP00046952 SNP00054416	SNP00054416 SNP00044426 SNP00155250 SNP00155250 SNP0004426 SNP00046952 SNP00054416 SNP0005416	SNP00054416 SNP00044426 SNP00155250 SNP0004426 SNP00046952 SNP00054416 SNP0005414 SNP00054365 SNP00054365	SNP00054416 SNP0004426 SNP00155250 SNP00155250 SNP0004426 SNP0004426 SNP0005416 SNP0005416 SNP00054365 SNP00054365 SNP00054365
ESTID			4646933H1	4648313H1	4648313H1	4648313H1	4648743H1	4818867H1	4818918H1	4819335H1	4822515H1	4822515H1	4822570H1	4822570H1	4887930H1	4888934H1	4889107H1	4890251H1	\$003015H1		5003015H1	5003015H1 5003615H1	5003015H1 5003615H1 5102495H1	5003615H1 5003615H1 5102495H1 5102562H1	5003015H1 5003615H1 5102495H1 5102562H1 5102562H1	5003015H1 5003615H1 5102495H1 5102562H1 5102562H1 5102902H1	5003015H1 5003615H1 5102495H1 5102562H1 5102502H1 5102902H1	5003015H1 5003615H1 5102495H1 5102562H1 5102502H1 5102902H1 5103232H1	5003015H1 5003615H1 5102495H1 5102562H1 5102562H1 5102902H1 5103232H1	5003015H1 5003615H1 5102495H1 5102562H1 5102562H1 5102902H1 5103232H1 5103232H1 5103232H1	5003015H1 5003615H1 5102495H1 5102562H1 5102562H1 5102902H1 5102902H1 5103232H1 5103232H1 510476H1
E.			7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	7504768	0/87046	/304/08	7504768	7504768 7504768	7504768 7504768 7504768	7504768 7504768 7504768 7504768	7504768 7504768 7504768 7504768 7504768	7504768 7504768 7504768 7504768 7504768	7504768 7504768 7504768 7504768 7504768 7504768	7504768 7504768 7504768 7504768 7504768 7504768 7504768	7504768 7504768 7504768 7504768 7504768 7504768 7504768	7504768 7504768 7504768 7504768 7504768 7504768 7504768 7504768
SEO	A	ÖZ	28	П	\neg	58		58	\neg	88	88			丁				\neg	\neg	_	T	Π									

Table 8

PD	ESTID	SNPID	EST	CB1	EST	Allele	Allele	Allele Amino Acid	Caucasian	African	Asian	Hispanic
			SNP	SNP			7		Allele 1	Allele 1	Allele 1	Allele 1
									frequency	frequency	frequency	frequency
	5107578H1	SNP00037414	49	546	၁	C	Ţ	noncoding	n/a	n/a	n/a	n/a
	5107578H1	SNP00054365	83	280	ß	Ð	ပ	noncoding	n/a	n/a	r/a	n/a
-	S107620H1	SNP00044426	228	242	A	G	4	noncoding	p/u	p/u	n/d	n/d
-	5108414H1	SNP00046952	76	74	g	Α .	ß	A20	n/a	n/a	n/a	n/a
5	5764353H1	SNP00146294	343	372			Ö	noncoding	n/a	n/a	n/a	n/a
8	5765817H1	SNP00044426	298	867	ß	ט	4	noncoding	p/u	n/d	n/d	p/u
-	784132H1	SNP00011894	102	747		ر ن	F	noncoding	n/a	n/a	n/a	n/a
00	825709H1	SNP00052446	8	380	Ø	O			r/a	n/a	n/a	n/a
00	825709H1	SNP00067987	48	420			-	noncoding	n/a	n/a	n/a	n/a
00	826297H1	SNP00054416	61	72		Ţ		Г	n/a	n/a	n/a	17/8
00	829954H1	SNP00011894	160	269	ن ن		F	noncoding	n/a	n/a	17/8	n/a
00	829954H1	SNP00129147	141	878	ິວ		H	noncoding	n/a	n/a	n/a	n/a
00	832039H1	SNP00011894	189	705				noncoding	n/a	n/a	r/a	n/a
00	832039H1	SNP00129147	170	989	ບ				n/a	n/a	n/a	n/a
∞∣	842177H1	SNP00063753	∞	4				noncoding	n/a	n/a	n/a	n/a
∞	843350H1	SNP00037414	29	531			Ţ	noncoding	n/a	n/a	n/a	n/a
∞∣	843350H1	SNP00054365	93	565					n/a	n/a	n/a	n/a
	1269986H1	SNP00028173	222	1745	ں ت	O		noncoding	n/a	n/a	n/a	n/a
-	1475518H1	SNP00106848	126	1287				noncoding	n/d	p/u	p/u	n/d
7	2156540H1	SNP00106847	158	1145	၁	ပ	ß		p/u	p/u	p/u	p/u
3	3915209H1	SNP00106846	111	872				noncoding	0.23	0.42	0.39	0.23
S	5773656H1	SNP00005518	250	1750				noncoding	n/a	n/a	n/a	n/a
∞	8624396H1	SNP00154333	46	1864	T	T		П	n/a	n/a	n/a	n/a
3	3188917H1	SNP00132428	171	255	C			A65	n/d	n/a	n/a	n/a
3	3316961HI	SNP00104542	168	1543	A	A	O	K500	n/d	p/u	p/u	p/u
S	5406235H1		51	1324	g		G	G427	0.87	0.87	0.85	0.88
-1	1235717H1		78	1041	ပ	C	A	noncoding	0.99	0.69		0.82
_1	1235717H1	SNP00011317		1098	ß	၂		noncoding	r/a	n/a		n/a
-1	1285865H1	SNP00130638 183		389	Α .	A	G	noncoding	n/a	n/a	n/a	n/a

Table 8

_			_	_	_		_	_	_	_	_	_	_	_	_	_	_					_	_	_	_	_			_		
Hispanic	Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	n/a	0.65	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	0.65	n/a	n/a	n/a	n/a	n/a	n/a	0.77	n/a	n/a	n/a	n/a	p/d	9/4
Asian	Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	n/a	0.36	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	0.36	n/a	n/a	n/a	n/a	n/a	n/a	0.79	n/a	n/a	n/a	n/a	0.98	
African	Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	n/a	0.24	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	0.24	n/a	n/a	n/a	n/a	n/a	n/a	0.84	n/a	n/a	n/a	n/a		
Caucasian	Allele 1	frequency	n/a	0.12	p/u	n/a	n/a	n/a	0.34	n/a	p/u	0.75	0.12	p/u	n/a	n/a	п⁄а	0.34	n/a	p/u	0.75	p/u	p/u	p/u	0.88	n/a	n/a	n/a	n/a		n/a
Allele Amino Acid			L68	noncoding	noncoding	noncoding	noncoding	noncoding	P54	760	noncoding	noncoding	noncoding	noncoding	noncoding	noncoding	noncoding	P223	0266	noncoding	noncoding	noncoding	noncoding	noncoding	R495	noncoding	T393	noncoding			noncoding
Allele	7		A	A	V	ບ		ຽ		A		g	A		ပ		ŋ	O	V				L L		H		U	G	F		₩.
Allele	-		G	G	G	T	T	Ą		ט		A			Ţ		A			S		C				S	A	A	ບ		Ö
EST	Allele		Ö	A	Ŋ	T	T	G	A	A		G	A	Ð	T	T	G	A	A		G	C	၁	C	၁	C	A	A	ر د	C	Ð
CB1	SNP		261	771	830	583	1187	965	221	349	818	1119	1278	1337	1090	1694	1472	728		1325	1626	405	899	2083	1943	2995	1637	384		1594	2781
EST	SNP		55	જ	124	118	10	249	127	74	159	29	65	124	118	10	249	127	74	159	29	163	96		7	166	136	79	113	455	431
CII ANS			SNP00132088	ľ		SNP00132471			SNP00001723	SNP00111346			SNP00011955			SNP00044590	SNP00059865	SNP00001723	SNP00111346	SNP00044588			SNP00060602	SNP00016820		_	SNP00098520	SNP00130277			SNP00016821
ESTID			1285865H1	1352080H1	1352080H1	1400156H1	2186481H1	2269083H1	2593286H1	2697792H1	409698H1	4148849H1	1352080H1	1352080H1	1400156H1	2186481H1	2269083H1	2593286H1	2697792H1	409698H1			3535269H1	1402206H1		1989003H1					6338096H1
PID			7500679	7500687	7500687	7500687	7500687	7500687	7500687	7500687	7500687	7500687	7500688	7500688	7500688	7500688	7500688	7500688	7500688	7500688	7500688	7500697	7500697	7500709	7500709	7500709	7500709	7500709	7500709	7500709	7500709
SEQ	A :	Ö	8	63	છ	છ	83	63	છ	63	8	8	3	8	প্ত	8	8	8	8	8	8	જ	8	8	99	98	99	9	8	Т	8

Table 8

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Hispanic	Allele 1	frequency	n/a	p/u	n/a	n/a	0.77	n/a	n/a	P/a	n/a	n/a	n/a	n/a	n/a	a/a	n/a	n/a	n/a	1 /a	Z/a	n/a	n/a	n/a	B/2	P/4	n/a	- F2	n/a	n/a	
Asian	Allele 1	frequency	n/a	0.98	n/a	n/a	0.79	n/a	n/a	0.98	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	p/a	n/a	n/a	n/a	n/a	2/6
African	Allele 1	frequency	n/a	0.99	n/a	n/a	0.84	n/a	n/a			n/a	n/a	n/a		n/a	n/a	1/8	n/a	r/a	r/a	n/a				P/Z					
Caucasian	Allele 1	frequency	n/a	0.99	n/a	p/u	0.88			0.99	n/a	n/a	n/a	n/a			n/a				n/a		n/a	n/a						n/a	n/a
Allele Amino Acid			D8S	A431	L111	noncoding	R538	noncoding	T384	H421	noncoding	noncoding	Г	D85	L111	L7	noncoding		Γ	Π									П	S135	7917
Allele	7			T		A	T					F			ь		4	۳							V		Ö		Ö		
Allele	-		G		C	C		၁		င		၁	G	g	ပ		ပ	ပ				G		ບ		ပ		H	T	၁	٣
EST	Allele		G		၁			C	Ą	၁		C			C							ט				U	V V		T	C	Ð
CB1	SNP		713	1752	162	2212		3124	0191	1723	384	2740	2910	713	162	147	1401	830	104	419	1489	1516	1484	1343 (1209	1133	363	584	541	562 (659
EST	SNP		368	175	318	6			136	792	79	113	431	398	80	84	203	168	53	86		174	142	71	240	111	310	222	179	120	135
SNP ID		_	SNP00146224	-7	\neg		SNP00016818 7		SNP00098520	SNP00068738	_	SNP00140107	SNP00016821	SNP00146224	SNP00152059	SNP00006328	SNP00006329	SNP00055365			SNP00010848 6		SNP00148038	SNP00010847 7	SNP00149612	SNP00115242	SNP00124832	SNP00007856		SNP00036749 1	SNP00074946
ESTID			6481368H1	7059872H1	7469363H1	1402206H1	1595983H1	1989003H1	2304976H1	2699021H1	3074049H1	3289688H1	6338096H1	6481368H1	7469363H1	1005080H1	1220531H1	1283008H1	1375166H1	1933092H1	1473474H1	1553396H1	1553396H1	1995636H1	2618624H1	3435845H1	6881314H1	1451293H1	1451293H1	1393846H1	1519711H1
윤			7500709	7500709	7500709	7500711	7500711	7500711	7500711	7500711	7500711	7500711	7500711	7500711	7500711	7500723	7500723	7500723	7500723	7500723	7500764	7500764	7500764	7500764	7500764	7500764	7500764	7501350	7501350	7506396	7506396
SEQ	<u>e</u>	ö	8	99	%	29	29	29	29	╗	63		\neg	29	\neg		\neg	89	1		\neg	7	٦	П			\$	71	\neg	1	72

Table 8

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Hispanic	Allele 1	frequency	n/a	24	n/a	n/a	n/a	r/a	n/a	n∕a	n/a	n/a	n/a	n/a	0.64	7⁄a															
Asian	Allele 1	frequency	n/a	p/u	n/a	0.74	n/a																								
African	Allele 1	frequency	n/a	p/u	n/a	ī/a	0.50	n/a																							
Caucasian	Allele 1	frequency	n/a	p/u	n/a	Z/a	n/a	n/a	n/a	29.0	n/a																				
Allele Amino Acid			P199	M50	G375	A401	G401	A367	P298	L305	A304	P298	L297	S304	R375	R401	A372	P367	A375	1.297	V303	R365	A276	P399	L272	F279	stop394	P405	P429	D100	S432
	7		G	T	G	T	Ŧ		Ţ			Ţ						T		T				T		C					T
Allele	-		ပ	၁	၁	၁	၁	၁	၁	Т	T	၁	ပ	Ţ	၁	င	၁	၁	၁	ပ	Т	ပ	Ţ	Ŋ	၁	T	ن د	၁	၁	S	C
EST	Ailele		ပ	Ţ	၁	၁	၁	C	C	Т	၁	၁	C	Т	ပ	ပ	၁	ပ	ပ	C	T	Ŋ	Т	ပ	ပ	T	T	ပ	၁	ပ	H
CBI	SNP		753	307	1406	1483	1484	1381	1175	1194	1193	1174	1172	1191	<u>설</u>	1482	1396	1380	1405	1170	1189	1374	1109	1477	1095	1116	1461	1494	1367	581	1575
EST	SNP		112	143	12	68	79	87	2	83	246	195	4	63	9	3	231	214	21	138	157	326	57	433	123	142	24	124	202	399	421
SNPID			SNP00074947	SNP00026722	SNP00019538	SNP00097325	SNP00097325	SNP00001985	SNP00006863	SNP00031912	SNP00031912	SNP00006863	SNP00006863	SNP00031912	SNP00019538	SNP00097325	SNP00019538	SNP00001985	SNP00019538	SNP00006863	SNP00031912	SNP00001985	SNP00031912	SNP00097325	SNP00006863	SNP00031912	SNP00001985	SNP00019538	SNP00097325	- 1	SNP00097325
ESTID			1649353H1	2614262H1	1224269H1	1224269H1	1364234H1	1467725H1	1734934H1	1734934H1	2875519H1	3408257H1	3803327H1	3803327H1	3876468H1	3876468H1	3995861H1	3997411H1	3997411H1	5992953H1	5992953H1	6307209H1	6307209H1	6307209H1	6307313H1	6307313H1	6719860H1	6719860H1	6719860H1		7333804HI
E E			7506396	7506396	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917	7505917
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Table 8

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Hispanic	Allele 1	frequency	n/a	n/a	n/a	p/u	1/8	n/a	p/u	n/a	n/a	n/a	n/a	n/a	n/a	1/a	0.78	n/a	0.77	n/a	0.88	ρg	n/a	n/a	0.87	n/a	2/3	2/3	n/a	n/a	9/4
Asian	Allele 1	frequency	n/a	n/a	n/a	0.96	n/a	n/a	96.0	n/a	n/a	n/a	n/a	n/a	n/a	n/a	p/u	n/a	p/u	n/a	p/u	p/u	n/a	n/a	0.99	n/a	n/a	n/a	n/a		
African	Allele 1	frequency	n/a	n/a	n/a	n/d	n/a	n/a	p/u	n/a	n/a	n/a	n/a	n/a	n/a		0.91		0.93	n/a	0.97	PZ.	n/a	n/a	0.92	n/a	n/a	n/a			
Caucasian	Allele 1	frequency	n/a	n/a	n/d	p/u	n/a	n/d	Zq Lq	n/d	p/u			n/a	2		0.73		2		0.85		n/a					n/a			
Allele Amino Acid			A395	noncoding	V94	R80	noncoding	767	R80	noncoding		Г	Г			noncoding			Γ	Y817	F820	S256	noncoding		F988		noncoding	Г			Г
Allele	7		L	T	٨		<u>-</u>	4			ر ت							ت ن			T			C					C		
Allele	-		C	C	G	G	U	Ö	O	Ţ	Ţ	[++	Ţ	Ð	Ţ		Ð			ပ				T	Ţ		H	U	H		U
EST	Allele		ပ		G	Ö	U		G		T	Ţ	G		C			Ţ	ບ	T			U		T	ī	H		T	ر د) ပ
CB1	S S		1465	593		293	746	336	293	1772	1937	2768	2757	3235	3285	3947	4317		2834	. 9662	3002			3284		3282	3945	4060	3942	4057 (4064 (
EST	SNS		432	191	135	191	161	135	161	188	146	16	4	399		171	87	15	83	245	254				235	50	172	786	142	257 4	100
SNPID		_		_		SNP00022822	SNP00020669				SNP00104924				_		SNP00004158					_1	\neg		_	7				$\overline{}$	SNP00115522
ESTID			7398053H1	1422265H1	2483938H1	3133415H1	1422265H1	2483938H1	3133415H1	1294191H1	3449686H1	4342984H1	4839169H1	6855931HI	1341905H1	1915202H1	2021533H1	2280153H1	2600125H1	2600125H1	2600125H1	2736801H1	3121639H1	3228112H1	3228112H1	3270946H1	3661149H1	3661149H1	3679593Н1	3679593Н1	3681589H1
E			7505917	7500701	7500701	7500701	7500702	7500702	7500702	6044343	6044343	6044343	6044343	6044343	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990
SEQ	<u>a</u> ;	ğ	2	74	- 1		22	2	75	92	92	92	9	9	1	7	7		П	11	11		11	1	1	11	1	11	1		11

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Hispanic	Allele 1	frequency	0.82	n/a	0.78	0.78	n/a	0.82	n/a	1 / a	0.78	n/a	n/a	n/a	0.78	n/a	p/u	0.86	0,60	09.0	n/a	0.77	n/a	0.88	n/a	n/a	n/a	n/a	n/a	n/a	n/a
Asian	Allele 1	frequency	66.0	n/a	n/d	₽/u	n/a	0.99	n/a	n/a	p/u	n/a	n/a	n/a	p/u	n/a	p/u	p/u	0.61	0.61	n/a	p/u	n/a	p/u	n/a	n/a	n/a	n/a	n/a	n/a	n/a
African	Allele 1	frequency	0.92	n/a	0.91	0.91	n/a	0.92	n/a	n/a	0.91	n/a	n/a	n/a	0.91	n/a	μ/d	0.95	n/a	n/a	n/a	0.93	n/a	0.97	n/a	n/a	n/a	n/a		n/a	n/a
Caucasian	Allele 1	frequency	0.77	n/a	0.73	0.73		0.77	n/a	0.46	0.73	n/a			0.73	n/a	n/d	62:0	0.87	0.87	n/a		0.78	0.85	n/a	n/a	p/u	n/a		n/a	n/a
Allele Amino Acid			L987	T221	noncoding	noncoding	Г	1988	noncoding	Γ	noncoding	noncoding		noncoding			R464	S684	S337	R300	L138	F763	D817	F820	попсодіпд	noncoding	noncoding	V145	noncoding	noncoding	noncoding
Allele	7			T	A		U		် ပ			S						υ U		G I					C		C		C		T
Allele	_		L	ပ	G	G	F	Ŧ	Ŧ	T		Ţ	Ð	Ţ	G	T	Ţ	£.		င			ن ن	C	T	ပ	T	G	T		U
EST	Allele		T	Ç	Y	Y	၁	Ţ	Ţ	၁	G	T	G	T		T				ပ			ບ			C		T	T		ن ن
CBI	SNP		3507	1209	4316	4306	4204	3510	4213	3283	4304	4202	959	4216	4252	4193	1939	7		1445	961	2836	2998	3007	4196	4043	3956	981	1		872
EST	SNP		91	128	71	134	32	58	197	113	134	32	2 2	62	107	各	11	173	367	317	208	269	431	440	84	237	504	706		271	129
SNP ID			SNP00096747	SNP00151675	SNP00004158	SNP00004158	SNP00024758	SNP00096747	SNP00024758	SNP00004157	SNP00004158	SNP00024758	SNP00062181	SNP00024758	SNP00004158	SNP00024758	SNP00115520	SNP00092898	SNP00092496	SNP00092496	SNP00062181	SNP00024755	SNP00024756	SNP00024757	SNP00024758	SNP00115522	SNP00115521	SNP00062181	SNP00024758	SNP00000037	SNP00044450
ESTID			3917071H1	4163704H1	4183129H1	4348376H1	4348376H1	4455065H1	4553662H1	4571868H1	5026526H1	5026526H1	5046938H1	5223889H1	5726796H1	645839H1	6549708H1	6573209H1	6804476J1	7152502H1	7161660H1	7262641H1	7262641H1	7262641H1	7326295H1	7326295H1	7603261H1	862601111	968309H1	6395382H1	1261613H1
PID			7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7503990	7504655	7504690
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SEQ	E.	ESTID	SNPID	EST	CB1	EST	Allele	Allele	Allele Amino Acid	Caucasian	African	Asian	Hispanic
日 :				SNP	SNP	Allele	-	7		Allele 1	Allele 1	Allele 1	Allele 1
Ö										frequency	frequency	frequency	frequency
79	7504690	1405907H1	SNP00044451	83	1414	T	၁	Ţ	noncoding	n/a	n/a	1/a	n/a
8	7504690	1680256H1	SNP00044450	176	698	ပ ပ		F	П	n/a	n/a	n/a	n/a
79	7504690	2040386H1	SNP00044451	83	1413	ں ن	U			n/a	n/a	n/a	n/a
5	7504690	SS75755H1	SNP00044450	119	871	ပ ပ		F	Γ	n/a	n/a	n/a	17/a
62	7504690	6397357H1	SNP00044450	24	865	ပ ပ	ပ	T	Г	n/a	n/a	n/a	n/a
8	7504720	025832H1	SNP00006719	154	992				Γ	n/a	n/a	n/a	8/4
80	7504720	1000318H1	SNP00070116	15	33	T	Ţ	် ပ		n/a	n/a	n/a	1/8
8	7504720	1349406H1	SNP00144339	210	8	∀	4		Π	n/a	11/8		2/2
8	7504720	138512H1	SNP00070116	27	32	T		U	Г	n/a	n/a		p/a
T	7504720	1464560H1	SNP00006719	69	986	<u></u>	9	A		n/a	n/a		n/a
&	7504720	167768H1	SNP00070116	28	21				Г	n/a	17/a		17/a
8	7504720	1932685H1	SNP00006719	242	993	G	<u>'</u>	A		n/a	n/a	r/a	17/a
&	7504720	1985942H1	SNP00144339	212	905					n/a	11/8		n/a
Т	7504720	1997242H1	SNP00006719	<u>18</u>	965	D D	່ ອ	A		n/a	n/a		n/a
8	7504720	2013940H1	SNP00144339	194	897	۷ ۷	A	_ O	noncoding	n/a	n/a	n/a	n/a
T	7504720	2108806H1	SNP00006719	166	982	O D			noncoding	n/a	n/a	n/a	n/a
2	7504720	2968021H1	SNP00070116	17	31	T	T	C	Г	n/a	n/a	n/a	n/a
T	7504720	3236115H1	SNP00006719	220	066	9			Г	n/a	n/a	n/a	n/a
T	7504720	3284457H1	SNP00070116	17		် ပ	T (C		n/a	n/a	n/a	n/a
T	7504720	3370771H1	SNP00006719	151	991					n/a	n/a	n/a	n/a
T	7504720	3472889H1	SNP00070116	23	27	C	T (C	noncoding	n/a	n/a		n/a
T	7504720	3607738H1	SNP00006719	125		9	9	A	Г	n/a	n/a		n/a
T	7504720	3705874H1	SNP00006719	270	686) Y	9				n/a		n/a
T	7504720	3735181H1	SNP00144339	212	968	/ D) V	r G	noncoding	n/a	n/a		n/a
T	7504720	3811868HI	SNP00006719	193	1000	Ð		A	noncoding	n/a	n/a		n/a
T	7504720	3860814H1	SNP00144339	231	894	/ Y	A (G					n/a
Т	7504720	4334301H1	SNP00006719	227		Ð	9	A	noncoding	n/a	n/a	n/a	n/a
T	7504720	6722906H1	SNP00006719	334				A	noncoding	n/a	n/a	n/a	n/a
8	7504720	688061H1	SNP00006719	137	985	r O	/ D	A		n/a			n/a

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Hispanic	Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	10/a	n/a	n/a	n/a	n/a	n/a	n/a	1/a	n/a	1/a	n/a	n/a	n/a	n/a	n/a	n/a	8/4	n/a	n/a
Asian	Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	
African	Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	1/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	P/a	ī/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a		
Caucasian	Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a										
Amino Acid			noncoding	noncoding	noncoding	noncoding	noncoding	noncoding	noncoding	noncoding				noncoding			L19	noncoding		coding	Г	noncoding	Γ	A17	noncoding	Т	oding		noncoding	Г	noncoding
9	7		T		T		1	T				T	T.	1	4				T-		Ŧ		1			T ,		Ţ		T	
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EST	Allele		Ü	Ü	C	C	ပ		U	U		ر ت			Ð			H		T		H	S		Ļ	Ü			H	U	
Ē	SNP		813	1122	558	811	812	. 608	805	810	1119	807	764	557	1120	289	239	290	223		238	288	509	234	284	237	287	228	277	236	П
EST	SNS		8	92	135	11	138	162	176	182	84	195	149	366	49	140	94	141	8	137	26	139	71	19	108		135		135	87	134
SNP ID		\neg	SNP00124454	SNP00092908	SNP00018343	SNP00124454	SNP00124454	SNP00124454	SNP00124454	SNP00124454	SNP00092908	SNP00124454	SNP00124454	SNP00018343		SNP00140932	SNP00034039	SNP00140932	SNP00034039	SNP00140932	SNP00034039	SNP00140932	SNP00034039	SNP00034039	SNP00140932	SNP00034039	SNP00140932	SNP00034039	SNP00140932	SNP00034039	SNP00140932
ESTID			030692H1	1286313H1	154958H1	2399811H1	3109003H1	3158872H1	4108913H1	4211203H1	4531755HI	5699517H1	6737674H1	6929007H1	879871H1	025850H1	026018H1	026018H1	138450H1	138450H1		139638H1	139751H1	166203H1	166203H1	166663H1	166663H1	166896H1		167123H1	167123H1
OLA M			7504722	7504722	7504722	7504722	7504722	7504722	7504722	7504722	7504722	7504722	7504722	7504722	7504722	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733
SEO	<u> </u>	ÿ Z	<u>~</u>	8	∞	∞	12	81	∞	∞	∞	81	81	18	81	82	82	82	82	82	22	82	82	82	82	82	82	82	82	82	82

Table 8

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Hispanic	Allele 1	frequency	n/a	n/a	n/a	n/a	r/a	n/a	n/a	n/a	n/a	n/a	n/a	0.69	n/a	n/a	n/a	n/a	n/a												
Asian	Allele 1	frequency	n/a	n/a	n/a	1/8	n/a	n/a	n/a	17a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	r/a	n/a	n/a	n/a	0.54	n/a	n/a	n/a	n/a	n/a
African	Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	0.61	n/a	n/a	n/a	n/a													
Caucasian	Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	0.59	n/a	n/a	n/d	n/a	n/a												
Amino Acid			noncoding	D17	noncoding	P19	noncoding	S14	noncoding	noncoding	noncoding	RIS	noncoding	L11	noncoding	T13	noncoding	noncoding	noncoding	noncoding	P16	noncoding	noncoding	noncoding	noncoding		K8	noncoding	noncoding		I^-
ä	7		ß	T	C			H				T			၁		g			Ð		C			C			פ	G		C
Allele	-		Ţ	C	T	C			Ţ		Ţ			C			Ţ			Ţ			A		Т	T	A	A	A		T
EST	Allele		T	င	Ţ	C	T		Ţ	င	T		T					L.		Ţ				T		T	٧		A		ပ
CB1	SNP		387	235	285	240	291	225	272	257	306	227	275	217			366		384	385	231		369	283	297	1891	70	74		2843	2841
EST	SKP		23	76	123	92	139	19	801		123	74	1		117					225			152		156	58	59	61		257	221
CII ANS			SNP00034041	SNP00034039	SNP00140932	_	SNP00034041		SNP00034041		SNP00034039	SNP00140932	SNP00034040	SNP00140932				SNP00026362	~		SNP00131351										
ESTID			2048217H1	2513378H1	2513378HI	2516759Н1	2516759H1	271049H1	271049H1	272900H1	272900H1	293166H1	293166H1	293308H1	293308H1	294628H1	294952H1	2957315H1	4086311H1	4416404H1	4797147H1	4797147H1	480396H1	5951186H1	7024134H1	1560345H1	3783254H1	2432516H1		2655227H1	3223832H1
E E			7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7504733	7507100	7503330	7504519	7504519	7504519	7504519
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Table 8

SEQ	PID	ESTID	CI days	EST	CB1	EST	Allele	Allele	Allele Amino Acid	Caucasian	African	Asian	Hienanic
				SNP	SNP		-	7		Allele 1	Allele 1	Allele 1	Allele 1
										frequency	frequency	frequency	frequency
	7504519	3223832H1	SNP00131352	271	1687	၁	် ပ	A	noncoding		n/a	n/a	n/a
	7504519	7169295H1	SNP00131352	437	2893) ၁	` ن			n/a	n/a	n/a	n/a
	7504738	010054H1	SNP00061272	134	393			၁	noncoding	n/a	n/a	n/a	n/a
	7504738	010089H1	SNP00058947	176	73	၁	. ၁	Ţ			n/a	n/a	n/a
	7504738	010482H1	SNP00146399	254	159		₅		1		n/a	n/a	n/a
	7504738	010838H1	SNP00002068	14		'	4	U	noncoding	n/a	n/a	n/a	n/a
	7504738	010934H1	SNP00019749	38	55		O	H		n/a	n/a	n/a	n/a
	7504738	010934H1	SNP00149440	83			0			n/a	n/a	n/a	7/a
	7504738	1289761H1	SNP00149440	73	66	Ð	, ເ		בוו	n/a	n/a	n/a	n/a
	7504738	154781H1	SNP00002068	14	30		Y	၁	noncoding	n/a	ī/a	n/a	n/a
	7504738	155182H1	SNP00061272	131	379	/ V					n/a	n/a	n/a
	7504738	2228722H1	SNP00149442	50	152					n/a	n/a	n/a	n/a
	7504738	2439274H1		63					noncoding	n/a	n/a	n/a	n/a
	7504738	3162461HI	SNP00058947	192	70	ິວ		T			n/a	n/a	n/a
	7504738	3229391H1	SNP00061272	30					noncoding	n/a	n/a	n/a	n/a
	7504738	3294683H1	SNP00058947	191		၁	C			n/a	n/a	n/a	n/a
	7504738	3685171H1	SNP00146399	272				A	A32	n/a	n/a	n/a	n/a
	7504738	3844355H1	SNP00052991	123					noncoding	p/u	n/a		n/a
	7504738	3845160H1	SNP00146399	274					G31	n/a	n/a		n/a
	7504738	4345360H1	SNP00052991	86	230		ر د	T		p/u	n/a	n/a	n/a
	7504738	4345360H1	SNP00146399	13	157	Ð	9		E31		n/a		n/a
	7504738	4345451H1	SNP00058947	163			C				n/a		n/a
	7504738	4347872H1	SNP00058947	151	69))			11	n/a	r/a		n/a
	7504738	4994751H1	SNP00002068	27	28	/ V	V		noncoding		r/a	n/a	r/a
	7504738	560674H1	SNP00052991	118	231) ၁	υ U	H	noncoding	n/d		n/a	n/a
	7504738	5978392H1	SNP00058947	178			C			n/a			n/a
	7504738	5978861H1	SNP00058947	172	74	د ر		F	Γ				2/a
	7504738	5982159H1	SNP00058947	185	29) ၁	ر ر						n/a
	7504738	5986062H1	SNP00146399 259	259	154	0	S S	4	E30	n/a			n/a

Table 8

			_	_	_	1	Τ_	_	1	7-	_	_	_	1	1	1		Т			_	_	_	_	Τ-	_	1	_	_	_	_
Hispanic	Allele 1	frequency	17/q	n/a	n/a	n/a	17/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	17/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	1/a
Asian	Allele 1	frequency	90.0	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n∕a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a
African	Allele 1	frequency	0.01	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a
Caucasian	Allele 1	frequency	p/u	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a		n/a	ī/a	0.99	n/a	66'0		n/a	0.99	66.0		0.05	n/a	n/a	n/a			n/a
Allele Amino Acid			noncoding	noncoding	noncoding	K239	noncoding	K239	noncoding	noncoding	noncoding	noncoding	noncoding	П	noncoding		noncoding	noncoding	noncoding		noncoding	Γ		noncoding	noncoding		П	noncoding		П	
Allele	7		C	T			L		Т						4			¥						V				A			ر ن
Allele	-		T	ာ	င		ပ		၁	ט	g					Ð			Ö		၁	ڻ ن	හ	۲	C	U	G	ບ	C		ß
EST	Allele		Ţ		၁	A	C	A	C	Ð	A					Ð				G			ڻ ق		C	ß	G	ر د	၁	G	Ð
GB.	SNP		. 09	381	393	1137	1388	1138	1386	286	548		437		438		136	434		134	369	135	127	436 (4506	4647	4772	4252 (4645 (
EST	SZ DZ		691	81	143	61	194	143	45	335	192	220	213	48	35	125	186	54	178	98	9	52	173	193	31	172	200	75 4	1	77	202
CII ANS			SNP00002069	SNP00105730	_	SNP00151238		SNP00151238		SNP00000806	SNP00000806			SNP00140641	SNP00140641									SNP00140641				SNP00036239			SNP00036243
ESTID			6091956H1	3735009H1	8103858H1	1336111H1	3417040H1	4596758H1	5943105H1	2483756H1	6119436H1	6119636H1	1456284H1	1541920H1	1622811H1	2110750H1	2138747H1	2322502H1	4527351H1	4639711H1	4946835H1	5163294H1	5165765H1	5533895H1	1212254H1	1212254H1	1215514H1	2611731H1	2611731H1	3087123H1	3087123H1
PID			7504738	7510280	7510280	7503700	7503700	7503700	7503700	7504685	7504685	7504685	7506844	7506844	7506844	7506844	7506844	7506844	7506844	7506844	7506844	7506844	7506844	7506844	7503772	7503772	7503772	7503772	7503772	7503772	7503772
SEO	A 9	ÿ	87	88	- 1		68	<u>&</u>	8	8	8	8	16	16	2	5	2	ೱ	2	ᇙ	16	2	2	2	8	8	8	8	જ્	8	8

Table 8

_			7	_	_	1	_	_	-	_	_	Ψ-	_	т	_	_	_	_	_	_	_	_	_	_	_	_	_	_	_		_
Hispanic	Allele 1	frequency	ī/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	7⁄a	n/a	n/a								
Asian	Allele 1	frequency	n/a	n/a	n/a	D/a	n/a	1/a	7/a	ī⁄a	g/a	ī/a	n/a	r∕a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	r/a	1/a	n/a	n/a	n/a	n/a	n/a	n/a
African	Allele 1	frequency	n/a	n/a	n/a	n/a	n/a	1/a	n/a	n/a	n/a	r/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a		n/a		n/a	n/a	n/a	n/a
Caucasian	Allele 1	frequency	n/a	n/a			n/a		n/a	n/a	n/a	2	n/a	n/a	0.05	n/a	n/a	n/a	n/a	0.05	ī/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	
Amino Acid			noncoding			П	Г	Г		Г	Г		noncoding	noncoding		P762	H774	noncoding	P538	noncoding			noncoding			noncoding	Г	noncoding	noncoding	noncoding	
Allele /	7			T	U					T		T		C					C				A				A				
Allele	_		ر د	S	O	U	ر ر	0		ပ ၁	0	C	G D		C	ر د		H	T	C	9 9			C	9	G G		ر 2	S G	ပ ပ	T
EST	Allele		S	၁	Ö	O	T	Ü) ၁) ၁		T (g		ပ ပ		၁			၁	Ð			C C		G		ر د			T
CB1	SNP		4254	4291	4763	4767	4290	4643	4255	4292	4769	3570	4633	4758		2389	2427		1718	4459	4600	4725	4205	4241	4598	4723	4207	4244		\neg	4243
EST	SNP		214	250	81	173	167	36	177	213	235	22	45			329			139	31			72	_	11	202		250			167
CII ANS			SNP00036239	SNP00036240	SNP00036243	SNP00036243	SNP00036240	SNP00036242	SNP00036239	SNP00036240	SNP00036243	SNP00036241		_	$\neg \tau$		\neg	SNP00138319	SNP00138319	$\neg \neg$	\neg			_	$\neg \neg$		- 1		- 1	\neg	SNP00036240
ESTID			3286120H2	3286120H2	3372558H1	3373510H1	3565279H1	4188096H1	4466544H1	4466544H1	4895923H1												2611731H1					~			3565279H1
PID			7503772	7503772	7503772	7503772	7503772	7503772	7503772	7503772	7503772	7503772	7503772	7503772	7503772	7503772	7503772	7503772	7503772	7503773	7503773	7503773	7503773	7503773	7503773	7503773	7503773	7503773	7503773	7503773	7503773
SEQ	<u>A</u>	Ö	8	8	96	96	96	96	8	8	96	8	8	96	96	8	8	96	8	2	8	22	2	2	6	22	97	5	8	7	97

SEQ	E C	ESTID	SNPID	EST	CB1	EST	Allele	Allele	Amino Acid	Caucasian	African	Acian	Hienanic
A				SNP	SNP	Allele		2		Allele 1	Allele 1	Allele 1	Allele 1
ÿ										frequency	frequency	frequency	frequency
8	7503773	4188096H1	SNP00036242	36	4596	Ŋ	ß	A	noncoding	n/a	n/a	n/a	n/a
8	7503773	4466544H1	SNP00036239	177	4208	၁	U	A	nóncoding	n/a	n/a	n/a	n/a
2	7503773	4466544H1	SNP00036240	213	4245	၁	ပ	F	noncoding	n/a	n/a	n/a	n/a
2	7503773	4895923H1	SNP00036243	235	4722	Ð	Ü	U	noncoding	n/a	n/a	n/a	n/a
2	7503773	6493623H1	SNP00036241	84	3652	1	ပ	H	C1183	0.05	n/a	n/a	n/a
2	7503773	6531015H1	SNP00036242	45	4586	Ð	0	4	noncoding	n/a	n/a	n/a	8/2
2	7503773	6531015H1	SNP00036243	170	4711	ပ	O	U	noncoding	n/a	n/a	n/a	n/a
2	7503773	6544492H1	SNP00036241	182	4457	ပ		ı	noncoding	0.05	n/a	n/a	n/a
22	7503773	8037780H1	SNP00138319	139	1718	ပ	۲		P538		n/a		8/2
8	7504698	030732H1	SNP00149756	215	453				noncoding		n/a		n/a
8	7504698	031619H1	SNP00149756	216	451	ß	Ö	A	noncoding		n/a	n/a	1/a
8	7504698	072413H1	SNP00149756	118	449	ß	Ð	A	noncoding		n/a		1/a
8	7504698	2188139H1	SNP00149756	248	424		ŋ		noncoding		n/a		n/a
8	7504698	2723635H1		35	527		G		noncoding	0	n/a	n/a	10/8
88	7504698	3090344H1	SNP00149756	102	452	Ð	ß	¥	noncoding		n/a		n/a
8	7504698	3572162H1	SNP00003052	50	526		ŋ		noncoding				n/a
88	7504698	3806693H1	SNP00149756 93	93	448		ט		noncoding	n/a			1/a
8	7504698	4071296H1		27	525	ß	Ö	4	noncoding				p/a
8	7504698	5276116H1	SNP00003052	24	520				noncoding	1.00			n/a
8	7504698	5768664H1	NP00141546	431	936		ວ		noncoding	n/a	n/a		n/a
8	7504698	5949629H1	- 1	250	450			A	noncoding	n/a	n/a		n/a
8	7504698	6211253H1	NP00149756	73	426	ŋ			noncoding	n/a			n/a
8	7504698	6938844H1	- 1	289	421				noncoding	1.00	n/a		n/a
8	7510361	1325602H1	SNP00105072	246	999		T	C	W207	n/a	n/a		n/a
- 1	7510361	1326305H1	SNP00137858	절	460		C		P138	n/a			n/a
- 1	7510361	1824780H1	SNP00105070	252	252	A	A		S69	n/d	p/u		p/u
8	7510361	1824858H1	SNP00105071	167	525				T160				9/2
8	7510361	1824858H1	SNP00105072	242	600	L	Ţ		L185				8/2
I	7510361	1825455H1	SNP00105072	195	\$65		H	U	L206				n/a

Table 8

					_		_								
Hispanic	Allele 1	frequency	μ/d	p/u	n/a	p/u	1/3	n/a	n/a	2/8	n/a	n/a	n/a	2/0	9/4
Asian	Allele 1	frequency	p/u	p/u	n/a	n/a	n/a	n/a	n/a	n/a			n/a		
African	Allele 1	frequency	ıv/d	υ/d	n/a	n/d	n/a	n/a	n/a	n/a	n/a	n/a	n/a	n/a	
Caucasian	Allele 1	frequency	p/u	n/d	n/a	n/d	n/a	0.99	n/a	n/a	n/a	n/a	n/a	n/a	
Allele Allele Amino Acid Caucasian			E220	N181	S207	noncoding	noncoding		G198	P199	C198	ding	1	t	
Allele	7		Ð	O	ပ	ပ	O	Ţ	F				O		
Allele	-		¥	4	Ţ	Ţ	V	U	ပ	ر د	U	V	A	ပ	
EST	Allele		٧	A	ပ	ပ	4	ပ	S	ပ	Ţ	Y	A	U	۲
CB1	SNP		<i>1</i> 0 <i>1</i>	685	299	723	2896	1955	878	879	876	2894	2855		1187
EST	SNP		13	295	370	323	001	137	136	242	262	247	171	288	86
SNPID			SNP00111130	SNP00105071 295	SNP00105072	SNP00058963 323	SNP00151312	SNP00031810 137	SNP00039104	SNP00039104	SNP00039104	SNF00151312	SNP00151312	SNP00039104 288	SNP00138949 98
ESTID			7653234H1	7939670H1	7939670H1	6041339H1	1748724H1	2137158H1	2864359H1	3327158H1	4376488H1	S691218H1	6409380H1	6482421H1	6623018J1
E.			7510361	7510361	7510361	7507013	7510507	7510507	7510507	7510507	7510507	7510507	7510507	7510507	7510507
SEO	A	Ö	8	83	8	8	101	101	[2]	. 101	101	101	. 101	. 101	101

What is claimed is:

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- 1. An isolated polypeptide selected from the group consisting of:
- a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-51,
- b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-5, SEQ ID NO:7, SEQ ID NO:10-11, SEQ ID NO:14, SEQ ID NO:17-20, SEQ ID NO:23-25, SEQ ID NO:27-38, SEQ ID NO:40-46, and SEQ ID NO:48-51
- a polypeptide comprising a naturally occurring amino acid sequence at least 92% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:6, SEQ ID NO:39, and SEQ ID NO:47,
- a polypeptide comprising a naturally occurring amino acid sequence at least 93%
 identical to the amino acid sequence of SEQ ID NO:21,
- e) a polypeptide comprising a naturally occurring amino acid sequence at least 94% identical to the amino acid sequence of SEQ ID NO:9,
 - f) a polypeptide comprising a naturally occurring amino acid sequence at least 98% identical to the amino acid sequence of SEQ ID NO:13,
 - g) a polypeptide consisting essentially of a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:8, SEQ ID NO:12, SEQ ID NO:15, SEQ ID NO:22, SEQ ID NO:26,
 - h) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, and
 - i) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-51.
 - 2. An isolated polypeptide of claim 1 comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-51.
- 30 3. An isolated polynucleotide encoding a polypeptide of claim 1.
 - 4. An isolated polynucleotide encoding a polypeptide of claim 2.

5. An isolated polynucleotide of claim 4 comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:52-102.

- 6. A recombinant polynucleotide comprising a promoter sequence operably linked to a5 polynucleotide of claim 3.
 - 7. A cell transformed with a recombinant polynucleotide of claim 6.
 - 8. A transgenic organism comprising a recombinant polynucleotide of claim 6.
 - 9. A method of producing a polypeptide of claim 1, the method comprising:
 - a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide, and said recombinant polynucleotide comprises a promoter sequence operably linked to a polynucleotide encoding the polypeptide of claim 1, and
 - b) recovering the polypeptide so expressed.

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- 10. A method of claim 9, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-51.
 - 11. An isolated antibody which specifically binds to a polypeptide of claim 1.
 - 12. An isolated polynucleotide selected from the group consisting of:
 - a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:52-102,
 - b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:52-53, SEQ ID NO:59-61, SEQ ID NO:65-67, SEQ ID NO:69, SEQ ID NO:71, SEQ ID NO:73-75, SEQ ID NO:77-79, SEQ ID NO:85-86, SEQ ID NO:89-92, SEQ ID NO:95-97, and SEQ ID NO:99-102,
 - a polynucleotide comprising a naturally occurring polynucleotide sequence at least
 91% identical to a polynucleotide sequence selected from the group consisting of SEQ

- ID NO:62, and SEQ ID NO:88,
- a polynucleotide comprising a naturally occurring polynucleotide sequence at least
 93% identical to a polynucleotide sequence selected from the group consisting of SEQ
 ID NO:54-56, SEQ ID NO:63, SEQ ID NO:83, and SEQ ID NO:93,
- 5 e) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 94% identical to the polynucleotide sequence of SEQ ID NO:64,
 - f) a polynucleotide comprising a naturally occurring polynucleotide sequence at least
 96% identical to a polynucleotide sequence selected from the group consisting of SEQ
 ID NO:68, SEQ ID NO:81, and SEQ ID NO:87,
- a polynucleotide comprising a naturally occurring polynucleotide sequence at least 98% identical to the polynucleotide sequence of SEQ ID NO:94,
 - a polynucleotide comprising a naturally occurring polynucleotide sequence at least
 99% identical to a polynucleotide sequence selected from the group consisting of SEQ
 ID NO:57, SEQ ID NO:82, and SEQ ID NO:98,
 - i) a polynucleotide consisting essentially of a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:58, SEQ ID NO:70, SEQ ID NO:72, SEQ ID NO:76, SEQ ID NO:80, and SEQ ID NO:84,
 - j) a polynucleotide complementary to a polynucleotide of a),
 - k) a polynucleotide complementary to a polynucleotide of b),
 - l) a polynucleotide complementary to a polynucleotide of c),
 - m) a polynucleotide complementary to a polynucleotide of d),
 - n) a polynucleotide complementary to a polynucleotide of e),
 - o) a polynucleotide complementary to a polynucleotide of f),
 - p) a polynucleotide complementary to a polynucleotide of g),
 - q) a polynucleotide complementary to a polynucleotide of h),
 - r) a polynucleotide complementary to a polynucleotide of i), and
 - s) an RNA equivalent of a)-r).

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30 13. An isolated polynucleotide comprising at least 60 contiguous nucleotides of a polynucleotide of claim 12.

14. A method of detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 12, the method comprising:

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- hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and
- b) detecting the presence or absence of said hybridization complex, and, optionally, if present, the amount thereof.

15. A method of claim 14, wherein the probe comprises at least 60 contiguous nucleotides.

- 16. A method of detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 12, the method comprising:
 - a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and
 - b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.
- 20 17. A composition comprising a polypeptide of claim 1 and a pharmaceutically acceptable excipient.
 - 18. A composition of claim 17, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-51.
 - 19. A method for treating a disease or condition associated with decreased expression of functional SECP, comprising administering to a patient in need of such treatment the composition of claim 17.
- 30 20. A method of screening a compound for effectiveness as an agonist of a polypeptide of claim 1, the method comprising:
 - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and

b) detecting agonist activity in the sample.

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21. A composition comprising an agonist compound identified by a method of claim 20 and a pharmaceutically acceptable excipient.

22. A method for treating a disease or condition associated with decreased expression of functional SECP, comprising administering to a patient in need of such treatment a composition of claim 21.

- 23. A method of screening a compound for effectiveness as an antagonist of a polypeptide of claim 1, the method comprising:
 - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
 - b) detecting antagonist activity in the sample.
- 24. A composition comprising an antagonist compound identified by a method of claim 23 and a pharmaceutically acceptable excipient.
 - 25. A method for treating a disease or condition associated with overexpression of functional SECP, comprising administering to a patient in need of such treatment a composition of claim 24.
 - 26. A method of screening for a compound that specifically binds to the polypeptide of claim 1, the method comprising:
 - a) combining the polypeptide of claim 1 with at least one test compound under suitable conditions, and
 - b) detecting binding of the polypeptide of claim 1 to the test compound, thereby identifying a compound that specifically binds to the polypeptide of claim 1.
 - 27. A method of screening for a compound that modulates the activity of the polypeptide of claim 1, the method comprising:
- a) combining the polypeptide of claim 1 with at least one test compound under conditions permissive for the activity of the polypeptide of claim 1,
 - b) assessing the activity of the polypeptide of claim 1 in the presence of the test

compound, and

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c) comparing the activity of the polypeptide of claim 1 in the presence of the test compound with the activity of the polypeptide of claim 1 in the absence of the test compound, wherein a change in the activity of the polypeptide of claim 1 in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide of claim 1.

28. A method of screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence of claim 5, the method
 10 comprising:

- exposing a sample comprising the target polynucleotide to a compound, under conditions suitable for the expression of the target polynucleotide,
- b) detecting altered expression of the target polynucleotide, and
- c) comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.
- 29. A method of assessing toxicity of a test compound, the method comprising:
- a) treating a biological sample containing nucleic acids with the test compound,
- b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide of claim 12 under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide comprising a polynucleotide sequence of a polynucleotide of claim 12 or fragment thereof,
- c) quantifying the amount of hybridization complex, and
- d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.
- 30. A method for a diagnostic test for a condition or disease associated with the expression of SECP in a biological sample, the method comprising:
 - a) combining the biological sample with an antibody of claim 11, under conditions suitable for the antibody to bind the polypeptide and form an antibody:polypeptide complex,

and

b) detecting the complex, wherein the presence of the complex correlates with the presence of the polypeptide in the biological sample.

- 5 31. The antibody of claim 11, wherein the antibody is:
 - a) a chimeric antibody,
 - b) a single chain antibody,
 - c) a Fab fragment,
 - d) a F(ab'), fragment, or
- 10 e) a humanized antibody.

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- 32. A composition comprising an antibody of claim 11 and an acceptable excipient.
- 33. A method of diagnosing a condition or disease associated with the expression of SECP in
 a subject, comprising administering to said subject an effective amount of the composition of claim 32.
 - 34. A composition of claim 32, further comprising a label.
- 35. A method of diagnosing a condition or disease associated with the expression of SECP in
 a subject, comprising administering to said subject an effective amount of the composition of claim 34.
 - 36. A method of preparing a polyclonal antibody with the specificity of the antibody of claim 11, the method comprising:
 - a) immunizing an animal with a polypeptide consisting of an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, or an immunogenic fragment thereof, under conditions to elicit an antibody response,
 - b) isolating antibodies from the animal, and
 - c) screening the isolated antibodies with the polypeptide, thereby identifying a polyclonal antibody which specifically binds to a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-51.
 - 37. A polyclonal antibody produced by a method of claim 36.

- 38. A composition comprising the polyclonal antibody of claim 37 and a suitable carrier.
- 39. A method of making a monoclonal antibody with the specificity of the antibody of claim 11, the method comprising:
 - a) immunizing an animal with a polypeptide consisting of an amino acid sequence selected from the group consisting of SEQ ID NO:1-51, or an immunogenic fragment thereof, under conditions to elicit an antibody response,
 - b) isolating antibody producing cells from the animal,
 - c) fusing the antibody producing cells with immortalized cells to form monoclonal antibody-producing hybridoma cells,
 - d) culturing the hybridoma cells, and
 - e) isolating from the culture monoclonal antibody which specifically binds to a
 polypeptide comprising an amino acid sequence selected from the group consisting of
 SEQ ID NO:1-51.

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- 40. A monoclonal antibody produced by a method of claim 39.
- 41. A composition comprising the monoclonal antibody of claim 40 and a suitable carrier.
- 20 42. The antibody of claim 11, wherein the antibody is produced by screening a Fab expression library.
 - 43. The antibody of claim 11, wherein the antibody is produced by screening a recombinant immunoglobulin library.

- 44. A method of detecting a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-51 in a sample, the method comprising:
 - incubating the antibody of claim 11 with the sample under conditions to allow specific binding of the antibody and the polypeptide, and
- 30 b) detecting specific binding, wherein specific binding indicates the presence of a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-51 in the sample.

45. A method of purifying a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-51 from a sample, the method comprising:

- a) incubating the antibody of claim 11 with the sample under conditions to allow specific binding of the antibody and the polypeptide, and
- separating the antibody from the sample and obtaining the purified polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-51.
- 46. A microarray wherein at least one element of the microarray is a polynucleotide of claim 10 13.
 - 47. A method of generating an expression profile of a sample which contains polynucleotides, the method comprising:
 - a) labeling the polynucleotides of the sample,

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- b) contacting the elements of the microarray of claim 46 with the labeled polynucleotides of the sample under conditions suitable for the formation of a hybridization complex, and
- c) quantifying the expression of the polynucleotides in the sample.
- 48. An array comprising different nucleotide molecules affixed in distinct physical locations on a solid substrate, wherein at least one of said nucleotide molecules comprises a first oligonucleotide or polynucleotide sequence specifically hybridizable with at least 30 contiguous nucleotides of a target polynucleotide, and wherein said target polynucleotide is a polynucleotide of claim 12.
- 49. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to at least 30 contiguous nucleotides of said target polynucleotide.
 - 50. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to at least 60 contiguous nucleotides of said target polynucleotide.
 - 51. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to said target polynucleotide.

52. An array of claim 48, which is a microarray.

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53. An array of claim 48, further comprising said target polynucleotide hybridized to a nucleotide molecule comprising said first oligonucleotide or polynucleotide sequence.

54. An array of claim 48, wherein a linker joins at least one of said nucleotide molecules to said solid substrate.

- 55. An array of claim 48, wherein each distinct physical location on the substrate contains multiple nucleotide molecules, and the multiple nucleotide molecules at any single distinct physical location have the same sequence, and each distinct physical location on the substrate contains nucleotide molecules having a sequence which differs from the sequence of nucleotide molecules at another distinct physical location on the substrate.
- 15 56. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:1.
 - 57. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:2.
 - 58. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:3.
 - 59. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:4.
 - 60. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:5.
- 25 61. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:6.
 - 62. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:7.
 - 63. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:8.
 - 64. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:9.
 - 65. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:10.

66. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:11. 67. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:12. 68. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:13. 5 69. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:14. 70. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:15. 10 71. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:16. 72. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:17. 73. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:18. 15 74. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:19. 75. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:20. 20 76. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:21. 77. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:22. 78. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:23. 25 79. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:24. 80. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:25. 30 81. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:26. 82. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:27.

83. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:28. 84. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:29. 85. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:30. 5 86. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:31. 87. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:32. 10 88. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:33. 89. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:34. 90. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:35. 15 91. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:36. 92. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:37. 20 93. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:38. 94. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:39. 95. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:40. 25 96. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:41. 97. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:42. 30 98. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:43.

99. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:44.

100. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:45.

- 101. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:46.
- 102. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:47.

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- 103. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:48.
- 104. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:49.
- 105. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:50.
- 106. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:51.
- 15 107. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:52.
 - 108. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:53.
 - 109. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:54.
- 110. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:55.
 - 111. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:56.
- 30 112. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:57.
 - 113. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID

NO:58.

114. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:59.

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- 115. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:60.
- 116. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:61.
 - 117. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:62.
- 15 118. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:63.
 - 119. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:64.

- 120. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:65.
- 121. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:66.
 - 122. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:67.
- 30 123. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:68.
 - 124. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID

NO:69.

125. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:70.

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- 126. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:71.
- 127. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID 10 NO:72.
 - 128. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:73.
- 15 129. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:74.
 - 130. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:75.

- 131. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:76.
- 132. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:77.
 - 133. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:78.
- 30 134. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:79.
 - 135. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID

NO:80.

136. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:81.

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- 137. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:82.
- 138. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID 10 NO:83.
 - 139. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:84.
- 15 140. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:85.
 - 141. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:86.

- 142. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:87.
- 143. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:88.
 - 144. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:89.
- 30 145. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:90.
 - 146. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID

NO:91.

147. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:92.

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- 148. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:93.
- 149. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:94.
 - 150. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:95.
- 15 151. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:96.
 - 152. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:97.

- 153. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:98.
- 154. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:99.
 - 155. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:100.
- 30 156. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:101.
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Cys Leu Ile Ile Leu Thr Cys Ile Ala Glu Asp Gln Tyr Ala Asn
                                   220
Ala Phe Leu His Asp Asp Asn Met Asn Phe Arg Val Asn Leu His
                230
                                   235
Arg Met Pro Met Arg His Arg Lys Lys Ala Ala Asp Lys Asn Leu
                245
                                    250
Pro Cys Arg Pro Leu Val Cys Ala Val Leu Asp Leu Met Val Glu
Phe Ile Val Thr His Met Met Lys Glu Phe Pro Met Asp Leu Tyr
                275
                                    280
Ile Arg Cys Ile Gln Val Val His Lys Leu Leu Cys Tyr Gln Lys
                290
                                    295
Lys Cys Arg Val Arg Leu His Tyr Thr Trp Arg Glu Leu Trp Ser
                305
                                   310
Ala Leu Ile Asn Leu Leu Lys Phe Leu Met Ser Asn Glu Thr Val
                320
                                   325
Leu Leu Ala Lys His Asn Ile Phe Thr Leu Ala Leu Met Ile Val
                335
                                    340
                                                        345
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Asn Leu Phe Asn Met Phe Ile Thr Tyr Gly Asp Thr Phe Leu Pro
              350
                                  355
Thr Pro Ser Ser Tyr Asp Glu Leu Tyr Tyr Glu Ile Ile Arg Met
               365
                                   370
His Gln Ser Phe Asp Asn Leu Tyr Ser Met Val Leu Arg Leu Ser
               380
                                   385
Thr Asn Ala Gly Gln Trp Lys Glu Ala Ala Ser Lys Val Thr His
               395
                                   400
Ala Leu Val Asn Ile Arg Ala Ile Ile Asn His Phe Asn Pro Lys
               410
                                   415
Ile Glu Ser Tyr Ala Ala Val Asn His Ile Ser Gln Leu Ser Glu
                425
                                   430
Glu Gln Val Leu Glu Val Val Arg Ala Asn Tyr Asp Thr Leu Thr
Leu Lys Leu Gln Asp Gly Leu Asp Gln Tyr Glu Arg Tyr Ser Glu
               455
                                   460
Gln His Lys Glu Ala Ala Phe Phe Lys Glu Leu Val Arg Ser Ile
               470
                                   475
Ser Thr Asn Val Arg Arg Asn Leu Ala Phe His Thr Leu Ser Gln
               485
                                   490
Glu Val Leu Leu Lys Glu Phe Ser Thr Ile Ser
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Met Gly Glu Leu Ser Pro Leu Gln Arg Pro Leu Ala Thr Glu Gly
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Thr Met Lys Ala Gln Gly Val Leu Leu Lys Leu Ala Leu Leu Ala
Leu Pro Leu Leu Leu Leu Ser Thr Pro Pro Cys Ala Pro Gln
Val Ser Gly Ile Arg Gly Asp Gly Phe Pro Glu Glu Ile Gln Trp
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Leu Ser Lys Phe Leu Pro Arg Leu Glu
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<211> 121
<212> PRT
<213> Homo sapiens
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<221> misc_feature
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Met Ala Leu Pro Pro Gly Pro Ala Ala Leu Arg His Thr Leu Leu
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<210> 13

<211> 290

<212> PRT

<213> Homo sapiens

<220>

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<223> Incyte ID No: 7500688CD1

<400> 13

Met Ala Leu Pro Pro Gly Pro Ala Ala Leu Arg His Thr Leu Leu 10 Leu Leu Pro Ala Leu Leu Ser Ser Gly Gly Pro Gly Thr Pro Arg 20 25 Leu Ala Trp Tyr Leu Asp Gly Gln Leu Gln Glu Ala Ser Thr Ser 35 40 Arg Leu Leu Ser Val Gly Gly Glu Ala Phe Ser Gly Gly Thr Ser 55 Thr Phe Thr Val Thr Ala His Arg Ala Gln His Glu Leu Asn Cys 65 Ser Leu Gln Asp Pro Arg Ser Gly Arg Ser Ala Asn Ala Ser Val Ile Leu Asn Val Gln Phe Lys Pro Glu Ile Ala Gln Val Gly Ala 100 Lys Tyr Gln Glu Ala Gln Gly Pro Gly Leu Leu Val Val Leu Phe Ala Leu Val Arg Ala Asn Pro Pro Ala Asn Val Thr Trp Ile Asp 130 125 Gln Asp Gly Pro Val Thr Val Asn Thr Ser Asp Phe Leu Val Leu 140 145 Asp Ala Gln Asn Tyr Pro Trp Leu Thr Asn His Thr Val Gln Leu 155 160 Gln Leu Arg Ser Leu Ala His Asn Leu Ser Val Val Ala Thr Asn 170 175 Asp Val Gly Val Thr Ser Ala Ser Leu Pro Ala Pro Gly Pro Ser 185 190 Arg His Pro Ser Leu Ile Ser Ser Asp Ser Asn Asn Leu Lys Leu 200 205 Asn Asn Val Arg Leu Pro Arg Glu Asn Met Ser Leu Pro Ser Asn

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215
                                   220
                                                        225
Leu Gln Leu Asn Asp Leu Thr Pro Asp Ser Arg Ala Val Lys Pro
                                   235
                230
Ala Asp Arg Gln Met Ala Gln Asn Asn Ser Arg Pro Glu Leu Leu
                                  250
                245
Asp Pro Glu Pro Gly Gly Leu Leu Thr Ser Gln Gly Phe Ile Arg
                                   265
                260
Leu Pro Val Leu Gly Tyr Ile Tyr Arg Val Ser Ser Val Ser Ser
                                   280
                275
Asp Glu Ile Trp Leu
                290
<210> 14
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Met Ser Ala Leu Ser Leu Leu Ile Leu Gly Leu Leu Thr Ala Val
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Pro Pro Ala Ser Cys Gln Gln Gly Ser Leu His Thr
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Met Leu Asp His Lys Asp Leu Glu Ala Glu Ile His Pro Leu Lys
                 5
                                    10
Asn Glu Glu Arg Lys Ser Gln Glu Asn Leu Gly Asn Pro Ser Lys
                                    25
Asn Glu Asp Asn Val Lys Ser Ala Pro Pro Gln Ser Arg Leu Ser
                 35
Arg Cys Arg Ala Ala Ala Phe Phe Leu Ser Leu Phe Leu Cys Leu
                 50
                                    55
Phe Val Val Phe Val Val Ser Phe Val Ile Pro Cys Pro Asp Arg
                 65
                                    70
Pro Ala Ser Gln Arg Met Trp Arg Ile Asp Tyr Ser Ala Ala Val
                                    85
Ile Tyr Asp Phe Leu Ala Val Asp Asp Ile Asn Gly Asp Arg Ile
                                   100
                 95
Gln Asp Val Leu Phe Leu Tyr Lys Asn Thr Asn Ser Ser Asn Asn
                                  115
Phe Ser Arg Ser Cys Val Asp Glu Gly Phe Ser Ser Pro Cys Thr
                                  130
                125
Phe Ala Ala Ala Val Ser Gly Ala Asn Gly Ser Thr Leu Trp Glu
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140
                                    145
Arg Pro Val Ala Gln Asp Val Ala Leu Val Glu Cys Ala Val Pro
                                   160
                155
Gln Pro Arg Gly Ser Glu Ala Pro Ser Ala Cys Ile Leu Val Gly
                                   175
                170
Arg Pro Ser Ser Phe Ile Ala Val Asn Leu Phe Thr Gly Glu Thr
                                   190
               185
Leu Trp Asn His Ser Ser Ser Phe Ser Gly Asn Ala Ser Ile Leu
                200
Ser Pro Leu Gln Val Pro Asp Val Asp Gly Asp Gly Ala Pro
                215
                                    220
Asp Leu Leu Val Leu Thr Gln Glu Arg Glu Glu Val Ser Gly His
                                    235
Leu Tyr Ser Gly Ser Thr Gly His Gln Ile Gly Leu Arg Gly Ser
                245
Leu Gly Val Asp Gly Glu Ser Gly Phe Leu Leu His Val Thr Arg
                260
                                    265
Thr Gly Ala His Tyr Ile Leu Phe Pro Cys Ala Ser Ser Leu Cys
                275
                                    280
                                                        285
Gly Cys Ser Val Lys Gly Leu Tyr Glu Lys Val Thr Glu Ser Gly
                290
                                   295
Gly Pro Phe Lys Ser Asp Pro His Trp Glu Ser Met Leu Asn Ala
               305
                                    310
Thr Thr Arg Arg Met Leu Ser His Ser Ser Gly Ala Val Arg Tyr
                320
                                    325
Leu Met His Val Pro Gly Asn Ala Gly Ala Asp Val Leu Leu Val
                335
                                    340
Gly Ser Glu Ala Phe Val Leu Leu Asp Gly Gln Glu Leu Thr Pro
                350
                                    355
Arg Trp Thr Pro Lys Ala Ala His Val Leu Arg Lys Pro Ile Phe
                                    370
                365
Gly Arg Tyr Lys Pro Asp Thr Leu Ala Val Ala Val Glu Asn Gly
                380
                                    385
Thr Gly Thr Asp Arg Gln Glu Thr Gly Glu Ala Arg His Ser Leu
                395
                                    400
Tyr Met Phe His Pro Thr Leu Pro Arg Val Leu Leu Glu Leu Ala
Asn Val Ser Thr His Ile Val Ala Phe Asp Ala Val Leu Phe Glu
                425
Pro Ser Arg His Ala Ala Tyr Ile Leu Leu Thr Gly Pro Ala Asp
                                    445
Ser Glu Ala Pro Gly Leu Val Ser Val Ile Lys His Lys Val Arg
Asp Leu Val Pro Ser Ser Arg Val Val Arg Leu Gly Glu Gly Gly
                                    475
Pro Asp Ser Asp Gln Ala Ile Arg Asp Arg Phe Ser Arg Leu Arg
                                    490
                485
Tyr Gln Ser Glu Ala
                500
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<211> 543

<212> PRT

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Leu Asp Leu Gly Thr Gly Ala Val Leu Cys Ser Leu Ala Leu Pro
               395
                                  400
Ser Leu Pro Gly Gly Pro Leu Ser Ala Ser Leu Pro Thr Ala Asp
                                   415
               410
His Arg Ser Ala Phe Phe Phe Trp Gly Leu His Glu Leu Gly Ser
               425
                                   430
Thr Ser Glu Thr Glu Thr Gly Glu Ala Arg His Ser Leu Tyr Met
                                   445
               440
Phe His Pro Thr Leu Pro Arg Val Leu Leu Glu Leu Ala Asn Val
                                   460
               455
Ser Thr His Ile Val Ala Phe Asp Ala Val Leu Phe Glu Pro Ser
                                   475
Arg His Ala Ala Tyr Ile Leu Leu Thr Gly Pro Ala Asp Ser Glu
                                   490
                485
Ala Pro Gly Leu Val Ser Val Ile Lys His Lys Val Arg Asp Leu
                                   505
                500
Val Pro Ser Ser Arg Val Val Arg Leu Gly Glu Gly Pro Asp
                                   520
Ser Asp Gln Ala Ile Arg Asp Arg Phe Ser Arg Leu Arg Tyr Gln
Ser Glu Ala
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Met Gly Val Pro Thr Ala Pro Glu Ala Gly Ser Trp Arg Trp Gly
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                                    10
Ser Leu Leu Phe Ala Leu Phe Leu Ala Ala Ser Leu Gly Pro Ser
                 20
                                    25
Pro Ala Pro Trp Arg Pro Pro Gly Cys Gln His Gln Pro Arg Pro
                 35
                                    40
Gly Ser Ala Pro Arg Ala Gly Val Gly Leu Arg Pro Pro Trp Gln
                                    55
Leu Leu His His His Ala Gln Pro Asp Pro Ala Gly
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Met Ser Trp Pro Arg Arg Leu Leu Leu Arg Tyr Leu Phe Pro Ala
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                                    10
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Leu Leu Leu His Ala Val Lys
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Met Phe Leu Lys Ala Val Val Leu Thr Leu Ala Leu Val Ala Val
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Ala Val Pro Ser Ser Arg Thr Asn Leu Glu Lys
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Met Ile Pro Ala Val Val Leu Leu Leu Leu Leu Val Glu Gln
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Ala Asp Pro Ser Ala Lys Gly Ser Tyr Asn Gln Leu
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Met Pro Gln Met Arg Gln Thr Pro Thr Asp Lys Pro Leu Cys Pro
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Ser Arg Thr His Lys Val Leu Pro Ile Leu Glu Ile Leu Tyr His
                20
                                    25
Val Glu Glu Arg Asn Ser His His Val Tyr Met Ala Leu Ile Ile
                 35
                                    40
Leu Leu Ile Leu Thr Glu Asp Asp Gly Phe Asn Arg Ser Ile His
                                    55
Glu Val Ile Leu Lys Asn Ile Thr Trp Tyr Ser Glu Arg Val Leu
                 65
                                    70
Thr Glu Ile Ser Leu Gly Ser Leu Leu Ile Leu Val Val Ile Arg
                80
                                    85
Thr Ile Gln Tyr Asn Met Thr Arg Thr Arg Asp Lys Tyr Leu His
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100
                                                       105
Thr Asn Cys Leu Ala Ala Leu Ala Asn Met Ser Ala Gln Phe Arg
               110
                                  115
Ser Leu His Gln Tyr Ala Ala Gln Arg Ile Ile Ser Leu Phe Ser
               125
                                   130
Leu Leu Ser Lys Lys His Asn Lys Val Leu Glu Gln Ala Thr Gln
                                   145
               140
Ser Leu Arg Gly Ser Leu Ser Ser Asn Asp Val Pro Leu Pro Asp
                                   160
               155
Tyr Val Ile Ser Phe Phe Ser Ser Arg Leu Clu Ala Gly Ala
                                   175
               170
Glu Leu Ser Val Glu Arg Val Leu Glu Ile Ile Lys Gln Gly Val
                                   190
                185
Val Ala Leu Pro Lys Asp Arg Leu Lys Lys Phe Pro Glu Leu Lys
                200
                                    205
                                                       210
Phe Lys Tyr Val Glu Glu Glu Gln Pro Glu Glu Phe Phe Ile Pro
                                   220
                215
Tyr Val Trp Ser Leu Val Tyr Asn Ser Ala Val Gly Leu Tyr Trp
               230
                                   235
Asn Pro Gln Asp Ile Gln Leu Phe Thr Met Asp Ser Asp
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Leu Leu Thr Asp Leu Tyr Gln Ala Thr Met Ala Leu Gly Tyr Trp
                 20
                                     25
Arg Ala Gly Arg Ala Arg Asp Ala Ala Glu Phe Glu Leu Phe Phe
                 35
                                     40
Arg Arg Cys Pro Phe Gly Gly Ala Phe Ala Leu Ala Ala Gly Leu
                                     55
Arg Asp Cys Val Arg Phe Leu Arg Ala Phe Arg Leu Arg Asp Ala
                 65
                                    70
Asp Val Gln Phe Leu Ala Ser Val Leu Pro Pro Asp Thr Asp Pro
                 80
                                    85
Ala Phe Phe Glu His Leu Arg Ala Leu Asp Cys Ser Glu Val Thr
                                    100
Val Arg Ala Leu Pro Glu Gly Ser Leu Ala Phe Pro Gly Val Pro
                110
                                   115
Leu Leu Gln Val Ser Gly Pro Leu Leu Val Val Gln Leu Leu Glu
                125
                                   130
Thr Pro Leu Cys Leu Val Ser Tyr Ala Ser Leu Val Ala Thr
                140
                                   145
Asn Ala Ala Arg Leu Arg Leu Ile Ala Gly Pro Glu Lys Arg Leu
                                   160
                155
Leu Glu Met Gly Leu Arg Arg Ala Gln Gly Pro Asp Gly Gly Leu
                170
                                    175
                                                        180
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Thr Ala Ser Thr Tyr Ser Tyr Leu Gly Gly Phe Asp Ser Ser Ser
                185
                                   190
Asn Val Leu Ala Gly Gln Leu Arg Gly Val Pro Val Ala Gly Thr
                200
                                    205
Leu Ala His Ser Phe Val Thr Ser Phe Ser Gly Ser Glu Val Pro
                215
                                    220
Pro Asp Pro Met Leu Ala Pro Ala Ala Gly Glu Gly Pro Gly Val
                230
                                    235
Asp Leu Ala Ala Lys Ala Gln Val Trp Leu Glu Gln Val Cys Ala
                245
His Leu Gly Leu Gly Val Gln Glu Pro His Pro Gly Glu Arg Ala
                260
Ala Phe Val Ala Tyr Ala Leu Ala Phe Pro Arg Ala Phe Gln Gly
                                    280
                275
Leu Leu Asp Thr Tyr Ser Val Trp Arg Ser Gly Leu Pro Asn Phe
                290
Leu Ala Val Ala Leu Ala Leu Gly Glu Leu Gly Tyr Arg Ala Val
                305
                                    310
Gly Val Arg Leu Asp Ser Gly Asp Leu Leu Gln Gln Ala Gln Glu
                320
                                   325
Ile Arg Lys Val Phe Arg Ala Ala Ala Ala Gln Phe Gln Val Pro
                335
                                   340
Trp Leu Glu Ser Val Leu Ile Val Val Ser Asn Asn Ile Asp Glu
                350
                                   355
Glu Ala Leu Ala Arg Leu Ala Gln Glu Leu Val Ala Val Gly Gly
                                   370
                365
Gln Pro Arg Met Lys Leu Thr Glu Asp Pro Glu Lys Gln Thr Leu
                380
                                    385
Pro Gly Ser Lys Ala Ala Phe Arg Leu Leu Gly Ser Asp Gly Ser
                395
                                    400
Pro Leu Met Asp Met Leu Gln Leu Ala Glu Glu Pro Val Pro Gln
                410
                                    415
Ala Gly Gln Glu Leu Arg Val Trp Pro Pro Gly Ala Gln Glu Pro
                                    430
                425
Cys Thr Val Arg Pro Ala Gln Val Glu Pro Leu Arg Leu Cys
                440
                                    445
Leu Gln Gln Gly Gln Leu Cys Glu Pro Leu Pro Ser Leu Ala Glu
                455
Ser Arg Ala Leu Ala Gln Leu Ser Leu Ser Arg Leu Ser Pro Glu
His Arg Arg Leu Arg Ser Pro Ala Gln Tyr Gln Val Val Leu Ser
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Glu Arg Leu Gln Ala Leu Val Asn Ser Leu Cys Ala Gly Gln Ser
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Pro
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<211> 127

<212> PRT

<213> Homo sapiens

<220>

<221> misc_feature

<223> Incyte ID No: 7500701CD1

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Pro Pro Pro Leu Tyr Thr Arg His Arg Met Leu Gly Pro Glu Ser
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                                    25
Val Pro Pro Pro Lys Arg Ser Arg Ser Lys Leu Met Ala Pro Pro
                                    40
Arg Ile Gly Thr His Asn Gly Thr Phe His Cys Asp Glu Ala Leu
                                    55
Ala Cys Ala Leu Leu Arg Leu Leu Pro Glu Tyr Arg Asp Ala Glu
                                    70
Ile Val Arg Thr Arg Asp Pro Glu Lys Leu Ala Ser Cys Asp Ile
                                    85
Val Val Asp Val Gly Gly Glu Tyr Asp Pro Arg Arg His Arg Tyr
                                   100
                95
Asp His His Gln Arg Gln Gly Ser Ser Val Gln Trp Ile Trp Phe
               110
                                  115
Lys Arg Ser Phe Cys Arg Asp
               125
<210> 24
<211> 137
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Met Gly His Arg Phe Leu Arg Gly Leu Leu Thr Leu Leu Pro
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Pro Pro Pro Leu Tyr Thr Arg His Arg Met Leu Gly Pro Glu Ser
                20
                                    25
Val Pro Pro Pro Lys Arg Ser Arg Ser Lys Leu Met Ala Pro Pro
                                    40
Arg Ile Gly Thr His Asn Gly Thr Phe His Cys Asp Glu Ala Leu
                                    55
Ala Cys Ala Leu Leu Arg Leu Leu Pro Glu Tyr Arg Asp Ala Glu
                                    70
Ile Val Arg Thr Arg Asp Pro Glu Lys Leu Ala Ser Cys Asp Ile
                 80
                                    85
Val Val Asp Val Gly Gly Glu Tyr Asp Pro Arg Arg His Arg Tyr
                                   100
Asp His His Gln Arg Cys Met Arg Thr Leu Trp Arg Arg Trp Met
               110
                                   115
Leu Trp Thr Met Gly Ser Pro Ser Gly Gln Arg Gly Ser Leu Asp
               125
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Met His
<210> 25
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<213> Homo sapiens

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85

80

Pro	Ser	Lys	Glu	Ser 95	Leu	Lys	Leu	Gln	Gly 100	Val	Phe	Ser	ГЛа	Gln 105
Thr	Va1	Leu	Lys	Ser 110	His	Pro	Leu	Leu	Ser 115	Gln	Ser	Tyr	Glu	Leu 120
Arg	Ala	Glu	Leu	Leu 125	Gly	Arg	Gln	Pro	Val 130	Leu	Glu	Phe	Ser	Leu 135
Glu	Asn	Leu	Arg	Thr 140	Met	Asn	Thr	Ser	Gly 145	Gln	Thr	Ala	Leu	Pro 150
Gln	Ala	Pro	Val	Asn 155	Gly	Leu	Ala	Lys	Lys 160	Leu	Thr	Lys	Ser	Ser 165
Thr	His	Ser	Asp	His 170	Asp	Asn	Ser	Thr	Ser 175	Leu	Asn	Gly	Gly	Lys 180
_				185					190				Gly	195
			_	200		_	_	_	205				Thr	210
		_		215	_				220				Ser	225
				230					235				Gln	240
				245					250				Ser	255
			_	260		_			265	-	_		Pro	270
				275					280	_		_	Ile	285
			_	290			_		295		_		Arg	300
				305					310				Glu	315
				320			_		325		_		Leu	330
-				335					340				Leu	345
				350					355				Ser	360
				365	_				370				Asn Ala	375
				380					385				Asp	390
				395					400				Thr	405
			-	410					415				Ser	420
	_			425					430				Trp	435
				440					445				Arg	450
_				455					460	_			Glu	465
				470					475				Gly	480
				485					490				Arg	495
116	Jer	GIU	261	500	361	1111	n is	Der	505	OIY	uid	Don	ar a	510

Val	Asn	Gly	Val	Ile 515	Asn	Thr	Leu	Gln	Pro 520	Val	Leu	Ala	Asp	His 525
Ile	Pro	Gly	Asp	Ser 530	Ser	Asp	Ala	Glu	Glu 535	Gln	Leu	His	Lys	Lys 540
Gln	Arg	Leu	Asn		Val	Ser	Ser	Ser		Asp	Gly	Thr	Сув	
Ala	Ala	Arg	Thr	Arg	Pro	Val	Leu	Ser	Cys	Lys	Lys	Arg	Arg	Leu
Val	Arg	Pro	Asn	560 Ser	Ile	Val	Pro	Leu	565 Ser	Lys	Lys	Val	His	
Asn	Ser	Thr	Ile	575 Arg	Pro	Gly	Cys	Asp	580 Val	Asn	Pro	Ser	Cys	585 Ala
Leu	Cvs	Glv	Ser	590 Glv	Ser	Ile	Asn	Thr	595 Met	Pro	Pro	Glu	Ile	600 His
				605				Leu	610					615
				620					625					630
				635				Asp	640					645
His	Phe	Gln	Ser	Met 650	Leu	Lys	Ser	Gln	Trp 655	Gln	Asn	Lys	Pro	Phe 660
Asp	Lys	Ile	Lys	Pro 665	Pro	Lys	Lys	Leu	Ser 670	Leu	ГЛа	His	Arg	Ala 675
Pro	Met	Pro	Gly	Ser 680	Leu	Pro	Asp	Ser	Ala 685	Arg	Lys	Asp	Arg	His 690
Lys	Leu	Val	Ser	Ser	Phe	Leu	Thr	Thr		Met	Leu	Lys	His	
Thr	qeA	Met	Ser		Ser	Ser	Tyr	Leu	Ala	Ala	Thr	His	His	Pro
Pro	His	Ser	Pro	710 Leu	Val	Arg	Gln	Leu	715 Ser	Thr	Ser	Ser	Asp	
Pro	Ala	Pro	Ala	725 Ser	Ser	Ser	Ser	Gln	730 V al	Thr	Ala	Ser	Thr	735 Ser
Gln	Gln	Pro	Val	740 Arg	Arg	Arg	Arg	Gly	745 Glu	Ser	Ser	Phe	Asp	750 Ile
				755				Val	760					765
				770				Leu	775				_	780
				785					790					795
				800				Ser	805					810
Glu	Ile	Glu	Asp	Leu 815	Ser	Asp	Ala	Ala	Phe 820	Ala	Ala	Leu	His	Ala 825
Lys	Сув	Glu	Glu	Met 830	Glu	Arg	Ala	Arg	Trp 835	Leu	Trp	Thr	Thr	Ser 840
Val	Pro	Pro	Gln	Arg 845	Arg	Gly	Ser	Arg	Ser 850	Tyr	Arg	Ser	Ser	Asp 855
Gly	Arg	Thr	Thr	Pro 860	Gln	Leu	Gly	Ser	Ala 865	Asn	Pro	Ser	Thr	Pro 870
Gln	Pro	Ala	Ser	Pro	Asp	Val	Ser	Ser	Ser	His	Ser	Leu	Ser	G1u
Tyr	Ser	His	Gly		Ser	Pro	Arg	Ser		Ile	Ser	Pro	Glu	885 Leu
His	Ser	Ala	Pro	890 Leu	Thr	Pro	Val	Ala	895 Arg	Asp	Thr	Leu	Arg	900 His
Leu	Ala	Ser	Glu	905 Asp	Thr	Arg	Суз	Ser	910 Thr	Pro	Glu	Leu	Gly	915 Leu
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Asp Glu Gln Ser Val Gln Pro Trp Glu Arg Arg Thr Phe Pro Leu
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                                  940
Ala His Ser Pro Gln Ala Glu Cys Glu Asp Gln Leu Asp Ala Gln
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                                  955
Glu Arg Ala Ala Arg Cys Thr Arg Arg Thr Ser Gly Ser Lys Thr
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               965
Gly Arg Glu Thr Glu Ala Ala Pro Thr Ser Pro Pro Ile Val Pro
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Leu Lys Ser Arg His Leu Val Ala Ala Ala Thr Ala Gln Arg Pro
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Thr His Arg
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Leu Ala Leu Val Leu Ala Leu Ala Pro Gly Leu Pro Thr Ala Arg
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Ala Gly Gln Thr Pro Arg Pro Ala Glu Arg Gly Pro Pro Val Arg
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Leu Phe Thr Glu Glu Glu Leu Ala Arg Tyr Gly Gly Glu Glu Ser
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Phe Met Asp Glu Glu Pro Pro Thr Met Pro
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Met Leu Leu Trp Pro Leu Leu Leu Leu Leu Leu Leu Pro Thr
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Leu Ala Leu Leu Arg Gln Gln Arg Ser Gln Asp Ala Arg Leu Ser
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Trp Leu Ala Gly Leu Gln His Arg Val Ala Trp Gly Ala Leu Val
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                                    40
Trp Ala Ala Thr Trp Gln Arg Arg Leu Glu Gln Ser Thr Leu
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                                    55
His Val His Gln Ser Gln Gln Gln Ala Leu Arg Trp Cys Leu Gln
                                    70
                 65
Gly Ala Gln Arg Pro His Cys Ser Leu Arg Arg Ser Thr Asp Ile
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85

80

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Ser Thr Phe Arg Asn His Leu Pro Leu Thr Lys Ala Ser Gln Thr
               95 100
Gln Gln Glu Asp Ser Gly Arg Gly Pro Ser Cys Trp Thr Met Ala
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Val Trp Arg Ala Ala Phe Trp Ile Pro Leu Arg Ala Leu Leu Pro
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Thr Thr Arg Cys Leu Trp Arg
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Met Ala Ser Arg Leu Thr Leu Leu Thr Leu Leu Leu Leu Leu Leu
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Ala Gly Gly Leu Gly Arg Thr Pro Lys Gln Thr Trp Arg Ala Ser
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Ser Leu Thr Pro Arg Thr Ser Pro Val Ser Thr Arg Pro
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Met Ala Ser Gly Ser Asn Trp Leu Ser Gly Val Asn Val Val Leu
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Val Met Ala Tyr Gly Ser Leu Val Phe Val Leu Leu Phe Ile Phe
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                20
Val Lys Arg Gln Ile Met Arg Phe Ala Met Lys Ser Arg Arg Gly
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Pro His Val Pro Val Gly His Asn Ala Pro Lys Val Ala Thr Thr
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Ile Cys Ile Gly
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Asp Trp Met Lys Met Lys Val Lys Leu Gly Ser Ala Leu Gly Gly
               305
                        310
Pro Tyr Leu Gly Val His Leu Arg Arg Lys Asp Phe Ile Trp Gly
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His Arg Gln Asp Val Pro Ser Leu Glu Gly Ala Gly Gly Thr Gly
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                                  340
Arg Pro Leu Ala
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Ala Gln Leu Leu Arg His Val Ala Ala Gln Asp Ala Gly Asp Gly
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Leu Arg Gly Gly Arg Ala Ala Ala Gly Leu Tyr Gly Glu Gln
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Gly Leu Leu His Gly
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Met Asp His Cys Gly Ala Leu Phe Leu Cys Leu Cys Leu Leu Thr
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Leu Gln Asn Ala Thr Thr Glu Thr Trp Glu Glu Leu Leu Ser Tyr
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Met Glu Asn Met Gln Val Ser Arg Gly Arg Ser Ser Val Phe Ser
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Ser Arg Gln Glu Val Ser Met Pro Gly Val Ser Thr Pro Cys Ser
Ser Pro Pro Ser
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<220>

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Met Lys Ser Leu Ile Leu Leu Ala Ile Leu Ala Ala Leu Ala Val
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Val Thr Leu Cys Tyr Glu Ser His Glu Ser Met Glu Ser Tyr Glu
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Leu Arg Ser Glu Asn Ala Leu Ser Leu Ser Thr Ser Ser Ile Gly
                                    40
                35
Lys Pro Val Met Thr Thr Asp Phe Ala Asn Ala Thr Pro Trp Phe
                                    55
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Met Asp Thr Met Leu Pro Ile Ile Ala Thr Ser Gly Ser Ala Glu
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Gly Pro Asn Glu Thr Glu Gly Arg Lys Ile Ser Phe Phe Leu
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Glu Ala Gly Thr
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Met Lys Leu Cys Val Thr Val Leu Ser Leu Leu Met Leu Val Ala
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Ala Phe Cys Ser Pro Ala Leu Ser Ala Pro Asn Ser Lys Pro Lys
                                   25
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Glu Ala Ser Lys Ser Val Leu Ile Pro Val Asn Pro Gly Ser Arg
                                   40
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Ser Thr Cys Met Thr Trp Asn
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Met Asp Asn Val Gln Pro Lys Ile Lys His Arg Pro Phe Cys Phe
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Ser Val Lys Gly His Val Lys Met Leu Arg Leu His Cys Leu Val
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Val Phe Met
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Cys Glu Leu Lys Pro Gln Gln Ser Glu Leu Asn Ser Phe Leu Trp
                35
                                    40
Thr Ile Lys Arg Asp Pro Pro Ser Tyr Phe Phe Gly Thr Ile His
                50
                                    55
                                                        60
Val Pro Tyr Thr Arg Val Trp Asp Phe Ile Pro Asp Asn Ser Lys
                65
                                   70
Glu Ala Phe Leu Gln Ser Ser Ile Val Tyr Phe Glu Leu Asp Leu
                80
                                    85
Thr Asp Pro Tyr Thr Ile Ser Ala Leu Thr Ser Cys Gln Met Leu
                95
                                   100
Pro Gln Gly Glu Asn Leu Gln Asp Val Leu Pro Arg Asp Ile Tyr
               110
                                   115
                                                        120
Cys Arg Leu Lys Arg His Leu Glu Tyr Val Lys Leu Met Met Pro
               125
                                   130
Leu Trp Met Thr Pro Asp Gln Arg Gly Lys Gly Leu Tyr Ala Asp
               140
                                   145
                                                       150
Tyr Leu Phe Asn Ala Ile Ala Gly Asn Trp Glu Arg Lys Arg Pro
               155
                                   160
Val Trp Val Met Leu Met Val Asn Ser Leu Thr Glu Val Asp Ile
               170
                                   175
Lys Ser Arg Gly Val Pro Val Leu Asp Leu Phe Leu Ala Gln Glu
               185
                                   190
Ala Glu Arg Leu Arg Lys Gln Thr Gly Ala Val Glu Lys Val Glu
               200
Glu Gln Cys His Pro Leu Asn Gly Leu Asn Phe Ser Gln Val Ile
               215
                                    220
Ser Trp Ala Thr Thr Gln Cys Trp Met Phe Cys Gly Val Lys Ala
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Met Arg
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Met Ala Ser Thr Val Val Ala Val Gly Leu Thr Ile Ala Ala Ala

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Gly Phe Ala Gly Arg Tyr Val Leu Gln Ala Met Lys His Met Glu
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Pro Gln Val Lys Gln Val Phe Gln Ser Leu Pro Lys Ser Pro Tyr
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Cys Gln
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Leu Ile Cys Cys Pro Gly Val Asn Glu Phe Gln Arg Gln Arg Val
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Pro Ala Ser Lys Ala Gly His Pro Asp Thr Ala Thr His Phe Gly
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Gly Cys Gly Gln Val Leu His His
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Ala Glu Tyr Gln Arg Lys Gln Gln Phe Trp Asp Ser Val Arg Leu
Ala Leu Phe Thr Leu Ala Ile Val Ala Ile Ile Gly Ile Ala Ile
Gly Ile Val Thr His Phe Val Val Glu Asp Asp Lys Ser Phe Tyr
                 50
                                    55
Tyr Leu Ala Ser Phe Lys Val Thr Asn Ile Lys Tyr Lys Glu Asn
                 65
                                    70
Tyr Gly Ile Arg Ser Ser Arg Glu Phe Ile Glu Arg Ser His Gln
Ile Glu Arg Met Arg
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<210> 42
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Trp Ala Gln Gly Leu Arg Gln Ala Pro Leu Ala Ser Gly Met Met

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80
                                     85
Thr Gly Thr Ile Glu Thr Thr Gly Asn Ile Ser Ala Glu Lys Gly
                 95
                                    100
Gly Ser Ile Ile Leu Gln Cys His Leu Ser Ser Thr Thr Ala Gln
                110
                                    115
Val Thr Gln Val Asn Trp Glu Gln Gln Asp Gln Leu Leu Ala Ile
                125
                                    130
Cys Asn Ala Asp Leu Gly Trp His Ile Ser Pro Ser Phe Lys Asp
                                    145
Arg Val Ala Pro Gly Pro Gly Leu Gly Leu Thr Leu Gln Ser Leu
                155
                                    160
Thr Val Asn Asp Thr Gly Glu Tyr Phe Cys Ile Tyr His Thr Tyr
                                    175
Pro Asp Gly Thr Tyr Thr Gly Arg Ile Phe Leu Glu Val Leu Glu
                                    190
Ser Ser Val Ala Glu His Gly Ala Arg Phe Gln Ile Pro Leu Leu
                                    205
Gly Ala Met Ala Ala Thr Leu Val Val Ile Cys Thr Ala Val Ile
                215
                                    220
Val Val Val Ala Leu Thr Arg Lys Lys Ala Leu Arg Ile His
                230
                                    235
Ser Val Glu Gly Asp Leu Arg Arg Lys Ser Ala Gly Gln Glu Glu
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                                    250
Trp Ser Pro Ser Ala Pro Ser Pro Pro Gly Ser Cys Val Gln Ala
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                                    265
Glu Ala Ala Pro Ala Gly Leu Cys Gly Glu Gln Arg Gly Glu Asp
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Cys Ala Glu Leu His Asp Tyr Phe Asn Val Leu Ser Tyr Arg Ser
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Leu Gly Asn Cys Ser Phe Phe Thr Glu Thr Gly
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 Met
 Gly
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 Asp
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 Gly
 Leu
 Ala
 Ala
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 Trp
 Cys
 Leu

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Gly	Gly	Cys	Gln	His 110	His	Суз	Val	Gln	Leu 115	Thr	Ile	Thr	Arg	His 120
Arg	Cys	Gln	Cys	Arg	Pro	Gly	Phe	Gln	Leu	Gln	G1u	Asp	Gly	Arg
His	Cys	Val	Arg	125 Arg	Ser	Pro	Cys	Ala		Arg	Asn	Gly	Ser	
				140				_	145		_	_	_	150
Met	His	Arg	Cys	Gln 155	Val	Val	Arg	Gly	Leu 160	Ala	Arg	Cys	Glu	Суз 165
His	Val	Gly	Tyr	Gln	Leu	Ala	Ala	Asp	_	Lys	Ala	Суз	Glu	_
		_		170	_			_	175				_	180
Val	Asp	Glu	Cys	Ala 185	Ala	Gly	Leu	Ala	Gln 190	Cys	Ala	His	Gly	Cys 195
Leu	Asn	Thr	Gln	Gly	Ser	Phe	Lys	Суз		Сув	His	Ala	Gly	
Glu	Leu	Glv	Ala	200 Asp	Glv	Ara	Gln	Cvs	205 T∨r	Ara	Ile	Glu	Met	210 Glu
		,		215	3			-,-	220	5				225
Ile	Val	Asn	Ser	Суз	Glu	Ala	Asn	Asn	Gly	Gly	Сув	Ser	His	Gly
				230					235					240
Суз	Ser	His	Thr	Ser 245	Ala	Gly	Pro	Leu	Cys 250	Thr	Сув	Pro	Arg	Gly 255
Tyr	Glu	Leu	Asp	Thr	Asp	Gln	Arg	Thr	Cys	Ile	Asp	Val	Asp	Asp
		_		260	_				265	_		_	_	270
Cys	Ala	Asp	Ser	Pro 275	Cys	Сув	Gln	Gln	Val 280	Суѕ	Thr	Asn	Asn	Pro 285
Glv	Glv	ጥ _ν ዮ	Glu	Cys	Glv	Cva	Tur	λla		Tvr	Ara	Leu	Ser	
CLY	OLY	-7-	OI u	290	Oly	CJ3	***	nzu	295	.,.	arg	Dea	561	300
Asp	Gly	Суз	Gly	Сув	Glu	Asp	Val	Asp		Сув	Ala	Ser	Ser	
-1	-1	~	61 .	305	•••	_	m)		310		01		D)	315
GIA	GIÀ	Cys	GIu	His 320	His	Cys	Thr	Asn	325	Ala	GIA	Ser	Phe	330
Суз	Ser	Сув	Glu	Ala 335	Gly	Tyr	Arg	Leu	His 340	Glu	Asp	Arg	Arg	Gly 345
Суз	Ser	Pro	Leu	Glu 350	Glu	Pro	Met	Va1	Asp 355	Leu	Asp	Gly	Glu	Leu 360
Pro	Phe	Val	Arg	Pro	Leu	Pro	His	Ile		Val	Leu	Gln	Asp	
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Leu	Pro	Gln	Leu	Phe	Gln	Asp	_					_	Gl.	Glu
Glu	_			380			Asp	Asp		Gly	Ala	Asp	Giu	390
	Ala	Glu	Leu	380 Arg	Gly	_	_	_	385	_		_		390 Val
				Arg 395	_	Glu	His	Thr	385 Leu 400	Thr	Glu	Lys	Phe	Val 405
Суз				Arg	_	Glu	His	Thr	385 Leu 400	Thr	Glu	Lys	Phe	Val 405
	Leu	Asp	Asp	Arg 395 Ser 410 Gly	Phe	Glu Gly	His His	Thr Asp	385 Leu 400 Cys 415 Leu	Thr Ser	Glu Leu	Lys Thr	Phe Cys	Val 405 Asp 420 Cys
Asp	Leu Cys	Asp Arg	Asp Asn	Arg 395 Ser 410 Gly 425	Phe Gly	Glu Gly Thr	His His Cys	Thr Asp Leu	385 Leu 400 Cys 415 Leu 430	Thr Ser Gly	Glu Leu Leu	Lys Thr Asp	Phe Cys Gly	Val 405 Asp 420 Cys 435
Asp	Leu Cys Cys	Asp Arg Pro	Asp Asn Glu	Arg 395 Ser 410 Gly 425 Gly 440	Phe Gly Trp	Glu Gly Thr	His His Cys Gly	Thr Asp Leu Leu	385 Leu 400 Cys 415 Leu 430 Ile 445	Thr Ser Gly Cys	Glu Leu Leu Asn	Lys Thr Asp Glu	Phe Cys Gly Thr	Val 405 Asp 420 Cys 435 Cys 450
Asp	Leu Cys Cys	Asp Arg Pro	Asp Asn Glu	Arg 395 Ser 410 Gly 425 Gly 440 Phe	Phe Gly Trp	Glu Gly Thr	His His Cys Gly	Thr Asp Leu Leu	385 Leu 400 Cys 415 Leu 430 Ile 445 Ser	Thr Ser Gly Cys	Glu Leu Leu Asn	Lys Thr Asp Glu	Phe Cys Gly Thr	Val 405 Asp 420 Cys 435 Cys 450 Cys
Asp Asp Pro	Leu Cys Cys Pro	Asp Arg Pro Asp	Asp Asn Glu Thr	Arg 395 Ser 410 Gly 425 Gly 440 Phe 455	Phe Gly Trp Gly	Glu Gly Thr Thr	His His Cys Gly Asn	Thr Asp Leu Leu Cys	385 Leu 400 Cys 415 Leu 430 Ile 445 Ser 460	Thr Ser Gly Cys Phe	Glu Leu Leu Asn Ser	Lys Thr Asp Glu Cys	Phe Cys Gly Thr	Val 405 Asp 420 Cys 435 Cys 450 Cys 465
Asp Asp Pro	Leu Cys Cys Pro	Asp Arg Pro Asp	Asp Asn Glu Thr	Arg 395 Ser 410 Gly 425 Gly 440 Phe	Phe Gly Trp Gly	Glu Gly Thr Thr	His His Cys Gly Asn	Thr Asp Leu Leu Cys	385 Leu 400 Cys 415 Leu 430 Ile 445 Ser 460	Thr Ser Gly Cys Phe	Glu Leu Leu Asn Ser	Lys Thr Asp Glu Cys	Phe Cys Gly Thr	Val 405 Asp 420 Cys 435 Cys 450 Cys 465
Asp Asp Pro Gln	Leu Cys Cys Pro Asn	Asp Arg Pro Asp Gly	Asp Asn Glu Thr	Arg 395 Ser 410 Gly 425 Gly 440 Phe 455 Thr	Phe Gly Trp Gly Cys	Glu Gly Thr Thr Lys Asp	His His Cys Gly Asn Ser	Thr Asp Leu Leu Cys Val	385 Leu 400 Cys 415 Leu 430 Ile 445 Ser 460 Thr 475	Thr Ser Gly Cys Phe Gly	Glu Leu Leu Asn Ser Ala	Lya Thr Asp Glu Cys	Phe Cys Gly Thr Ser Arg	Val 405 Asp 420 Cys 435 Cys 450 Cys 465 Cys 480
Asp Pro Gln	Leu Cys Cys Pro Asn	Asp Arg Pro Asp Gly	Asp Asn Glu Thr Gly Val	Arg 395 Ser 410 Gly 425 Gly 440 Phe 455 Thr 470 Ser 485	Phe Gly Trp Gly Cys	Glu Gly Thr Thr Lys Asp	His His Cys Gly Asn Ser	Thr Asp Leu Leu Cys Val	385 Leu 400 Cys 415 Leu 430 Ile 445 Ser 460 Thr 475 Glu 490	Thr Ser Gly Cys Phe Gly Asp	Glu Leu Leu Asn Ser Ala Gly	Lya Thr Asp Glu Cys Cys	Phe Cys Gly Thr Ser Arg	Val 405 Asp 420 Cys 435 Cys 450 Cys 465 Cys 480 Lys 495
Asp Pro Gln	Leu Cys Cys Pro Asn	Asp Arg Pro Asp Gly	Asp Asn Glu Thr Gly Val	Arg 395 Ser 410 Gly 425 Gly 440 Phe 455 Thr 470 Ser	Phe Gly Trp Gly Cys	Glu Gly Thr Thr Lys Asp	His His Cys Gly Asn Ser	Thr Asp Leu Leu Cys Val	385 Leu 400 Cys 415 Leu 430 Ile 445 Ser 460 Thr 475 Glu 490	Thr Ser Gly Cys Phe Gly Asp	Glu Leu Leu Asn Ser Ala Gly	Lya Thr Asp Glu Cys Cys	Phe Cys Gly Thr Ser Arg	Val 405 Asp 420 Cys 435 Cys 450 Cys 465 Cys 480 Lys 495
Asp Pro Gln Pro	Leu Cys Cys Pro Asn Pro	Asp Pro Asp Gly Gly	Asp Asn Glu Thr Gly Val	Arg 395 Ser 410 Gly 425 Gly 440 Phe 455 Thr 470 Ser 485 Lys	Phe Gly Trp Gly Cys Gly	Glu Gly Thr Thr Lys Asp Thr Cys	His His Cys Gly Asn Ser Asn	Thr Asp Leu Cys Val Cys	385 Leu 400 Cys 415 Leu 430 Ile 445 Ser 460 Thr 475 Glu 490 Lys 505	Thr Ser Gly Cys Phe Gly Asp Cys	Glu Leu Leu Asn Ser Ala Gly	Lya Thr Asp Glu Cya Cya Cya Cya	Phe Cys Gly Thr Ser Arg Pro	Val 405 Asp 420 Cys 435 Cys 450 Cys 465 Cys 480 Lys 495 Asn 510

Gly	Leu	Tyr	Gly	Arg 530	Phe	Cys	His	Leu	Thr 535	Cys	Pro	Pro	Trp	Ala 540
Phe	Gly	Pro	Gly		Ser	Glu	Glu	Суз		Cys	Val	Gln	Pro	
Thr	Gln	Ser	Сув		Lys	Arg	Asp	Gly		Cys	Ser	Cys	Lys	
Gly	Phe	Arg	Gly		Arg	Суз	Gln	Ala		Cys	Glu	Leu	Gly	
Phe	Gly	Pro	Gly	_	Trp	Gln	Ala	Суз		Сув	Pro	Val	Gly	
Ala	Суѕ	Asp	Ser		Ser	Gly	Glu	Сув		Lys	Arg	Cys	Pro	
Gly	Phe	Gln	Gly		Asp	Суз	Gly	Gln		Суз	Pro	Va1	Gly	
Phe	Gly	Val	Asn		Ser	Ser	Ser	Сув		Суз	Gly	Gly	Ala	
Суз	His	Gly	Val	Thr 650	Gly	Gln	Сув	Arg	Cys 655	Pro	Pro	Gly	Arg	Thr 660
Gly	Glu	Asp	Суз	Glu 665	Ala	Asp	Суз	Pro	Glu 670	Gly	Arg	Trp	Gly	Leu 675
Gly	Суз	Gln	Glu	Ile 680	Суз	Pro	Ala	Суз	Gln 685	His	Ala	Ala	Arg	Cys 690
Asp	Pro	Glu	Thr	Gly 695	Ala	Сув	Leu	Суз	Leu 700	Pro	Gly	Phe	Val	Gly 705
Ser	Arg	Суз	Gln	Asp 710	Val	Cys	Pro	Ala	Gly 715	Trp	Tyr	Gly	Pro	Ser 720
Суз	Gln	Thr	Arg	Cys 725	Ser	Cys	Ala	Asn	Asp 730	Gly	His	Суѕ	His	Pro 735
Ala	Thr	Gly	His	Cys 740	Ser	Сув	Ala	Pro	Gly 745	Trp	Thr	Gly	Phe	Ser 750
_		_	Ala	755	_		_		760	_		_	_	765
			Asn	770					775					780
			Суз	785					790					795
			Cys	800					805					810
			Сув	815					820					825
			Сув	830					835					840
			Ala	845					850					855
			Ala	860			_	_	865					870
	_		Ala	875					880					885
			Thr	890					895					900
		_	Ala	905	-	-			910	-		_		915
			Trp	920	_			_	925			_		930
vrA	vsħ	val	Arg	935	GTÅ	Cys	wrd	nis	940	GIÀ	GIÀ	cha	Ded	945

Gly Gly Leu Cys Asp Pro His Thr Gly Arg Cys Leu Cys Pro Ala 950 955 Gly Trp Thr Gly Asp Lys Cys Gln Ser Pro Cys Leu Arg Gly Trp 965 970 Phe Gly Glu Ala Cys Ala Gln Arg Cys Ser Cys Pro Pro Gly Ala 985 980 Ala Cys His His Val Thr Gly Ala Cys Arg Cys Pro Pro Gly Phe 995 1000 Thr Gly Ser Gly Cys Glu Gln Gly Cys Pro Pro Gly Arg Tyr Gly 1010 1015 Pro Gly Cys Glu Gln Leu Cys Gly Cys Leu Asn Gly Gly Ser Cys 1025 1030 1035 Asp Ala Ala Thr Gly Ala Cys Arg Cys Pro Thr Gly Phe Leu Gly Thr Asp Cys Asn Leu Ser Glu Trp Leu Val Ala Ala Val Leu Ser 1055 1060 Gly Ala Ser Cys Val Ser Arg Pro Val Arg Arg Ala Ala Ser Ala 1070 1075 Pro Thr Ala Pro Thr Cys Val Gly Val Gly Arg Gly Arg Pro Ala 1085 1090 Thr Leu

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<212> PRT

<213> Homo sapiens

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_		_		170					175					180
Val	Asp	Glu	Cys		Ala	Gly	Leu	Ala		Cys	Ala	His	Gly	
T on	Non	Wh-	<i>0</i> 1n	185	C	Phe	7	~	190	~	w:	21-	01	195
Deu	ASII	1111	GIII	200	261	rne	rAs	Cys	205	Cys	nrs	NIG	GTÀ	210
Glu	Leu	Gly	Ala		Glv	Arg	Gln	Cvs		Ara	Ile	Glu	Met	
		_	_	215	•	5			220	5				225
Ile	Val	Asn	Ser	Сув	Glu	Ala	Asn	Asn	Gly	Gly	Сув	Ser	His	
				230					235					240
Сув	Ser	His	Thr		Ala	Gly	Pro	Leu		Thr	Cys	Pro	Arg	_
		Y	•	245		0.7	_	 3	250		_	1	_	255
ıyı	GIU	Leu	Asp	260	Asp	Gln	Arg	Thr	265	TTE	Asp	vaı	Asp	270
Cys	Ala	Asp	Ser		Cvs	Cys	Gln	Gln		Cvs	Thr	Asn	Aan	
-				275					280	-,-				285
Gly	Gly	Tyr	Glu	Cys	Gly	Сув	Tyr	Ala	Gly	Tyr	Arg	Leu	Ser	
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Asp	Gly	Суз	Gly		Glu	Asp	Val	Asp		Cys	Ala	Ser	Ser	
Glv.	O1	~	01	305	***	~	m b	3	310		03		-	315
GTA	GIY	Cys	GIU	320	nıs	Сув	Inr	ASN	325	Ala	втА	Ser	Phe	330
Cys	Ser	Суз	Glu		Gly	Tyr	Ara	Leu		Glu	Asp	Ara	Ara	
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Cys	Ser	Pro	Leu	Glu	Glu	Pro	Met	Val	Asp	Leu	Asp	Gly	Glu	Leu
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Pro	Phe	Val	Arg		Leu	Pro	His	Ile		Val	Leu	Gln	Asp	
Lon	Dro	Gla	Lon	365	C1-	Asp	>	3	370	03	21-	>	o1	375
Deu	FIG	GIII	Dea	380	GIII	ASD	Asp	Asp	385	GIY	Ala	Asp	GIU	390
Glu	Ala	Glu	Leu		Gly	Glu	His	Thr		Thr	Glu	Lvs	Phe	
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CAa	Leu	Asp	Asp	Ser	Phe	Gly	His	Asp	Сув	Ser	Leu	Thr	Cys	Asp
_	_	_	_	410			_	_	415				_	420
Asp	Cys	Arg	Asn	Gly 425	Gly	Thr	Cys	Leu		Gly	Leu	Asp	Gly	-
Asp	Cva	Pro	Glu		Trn	Thr	Glv	Len	430	Cve	Aan	GI.	Th-	435
	-,-			440		****	O1,		445	CJS	ASII	Gid	1111	450
Pro	Pro	Asp	Thr	Phe	Gly	Lys	Asn	Cys		Phe	Ser	Суз	Ser	
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Gln	Asn	Gly	Gly		Cys	Ąsp	Ser	Val		Gly	Ala	Суз	Arg	
Dro	Dwo	01	17-1	470	01	mb	>	~	475	•	61		_	480
FIO	PIO	GIA	val	485	GIY	Thr	ASN	Cys	490	Asp	GIÀ	Cys	Pro	Lys 495
Gly	Tyr	Tyr	Gly	-	His	Cys	Ara	Lvs		Cvs	Asn	Cvs	Ala	
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Arg	Gly	Arg	Cys	His	Arg	Leu	Tyr	Gly	Ala	Суз	Leu	Суз	Asp	Pro
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Gly	Leu	Tyr	Gly		Phe	Сув	His	Leu		Cys	Pro	Pro	Trp	
Pho	G1v	Dro	01	530	C	01	03	^	535	~	17- 1	67	_	540
rne	GLY	PIO	GIY	545	Ser	Glu	GIU	cys	550	сув	vaı	GIN	PTO	н18 555
Thr	Gln	Ser	Cys		Lys	Arg	Asp	Glv		Cvs	Ser	Cvs	Lvs	
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Gly	Phe	Arg	Gly		Arg	Cys	Gln	Ala	Glu	Cys	Glu	Leu	Gly	
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Phe	Gly	Pro	Gly	Суѕ	Trp	Gln	Ala	Cys	Thr	Cys	Pro	Va1	Gly	Val

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Ala	Cys	Asp	Ser		Ser	Gly	G1u	Cys	_	Lys	Arg	Cys	Pro	
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Gly	Phe	Gln	Gly	Glu 620	Asp	Cys	Gly	Gln	G1u 625	Cys	Pro	Val	Gly	Thr 630
Phe	Gly	Val	Asn	Cys 635	Ser	Ser	Ser	Суз	Ser 640	Суѕ	Gly	Gly	Ala	Pro 645
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Gly	Glu	Asp	Cys		Ala	Asp	Сув	Pro		Gly	Arg	Trp	Gly	
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Asp	Pro	Glu	Thr		Ala	Суз	Leu	Суз	Leu	Pro	Gly	Phe	Va1	
Ser	Arg	Cys	Gln		Val	Сув	Pro	Ala		Trp	Tyr	Gly	Pro	
Сув	Gln	Thr	Arg	Сув	Ser	Cys	Ala	Asn	_	Gly	His	Суз	His	
Ala	Thr	Gly	His		Ser	Сув	Ala	Pro		Trp	Thr	Gly	Phe	
Cys	Gln	Arg	Ala		Asp	Thr	Gly	His		Gly	Pro	Asp	Сув	
His	Pro	Cys	Asn		Ser	Ala	Gly	His		Ser	Cys	Asp	Ala	
Ser	Gly	Leu	Cys		Сув	Glu	Ala	Gly	_	Val	Gly	Pro	Arg	-
Glu	Gln	Gln	Сув		Gln	Gly	His	Phe	_	Pro	Gly	Суз	Glu	
Leu	Cys	Gln	Cys		His	Gly	Ala	Ala		Asp	His	Val	Ser	
Ala	Cys	Thr	Сув		Ala	Gly	Trp	Arg		Thr	Phe	Сув	Glu	
Ala	Cys	Pro	Ala		Phe	Phe	Gly	Leu		Суз	Arg	Ser	Ala	
Asn	Cys	Thr	Ala	845 Gly 860	Ala	Ala	Суз	Asp		Val	Asn	Gly	Ser	
Leu	Cys	Pro	Ala		Arg	Arg	Gly	Pro	865 Arg 880	Cys	Ala	Glu	Thr	
Pro	Ala	His	Thr		Gly	His	Asn	Cys	Ser	Gln	Ala	Суз	Ala	
Phe	Asn	Gly	Ala		Cys	Asp	Pro	Val	895 His 910	Gly	Gln	Cys	His	
Ala	Pro	Gly	Trp		Gly	Pro	Ser	Сув		Gln	Glu	Суз	Leu	
Arg	Asp	Val	Arg		Gly	Суѕ	Arg	His		Gly	G1y	Сув	Leu	930 Asn 945
Gly	Gly	Leu	Cys		Pro	His	Thr	Gly		Суз	Leu	Cys	Pro	
Gly	Trp	Thr	Gly		Lys	Суз	Gln	Ser		Сув	Leu	Arg	Gly	Trp
Phe	Gly	Glu	Ala		Ala	Gln	Arg	Cys	Ser	Суз	Pro	Pro	Gly	
Ala	Cys	His	His		Thr	Gly	Ala	Cys 1	985 Arg 1000	Cys	Pro	Pro		
Thr	Gly	Ser	Gly		Glu	Gln	Ala	Cys		Pro	Gly	Ser		.005 Gly

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                                1030
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                                1045
Gly Pro Ser Cys Gln Gln Arg Cys Pro Pro Gly Arg Tyr Gly Pro
              1055
                                1060
Gly Cys Glu Gln Leu Cys Gly Cys Leu Asn Gly Gly Ser Cys Asp
              1070
                                 1075
Ala Ala Thr Gly Ala Cys Arg Cys Pro Thr Gly Phe Leu Gly Thr
                                 1090
Asp Cys Asn Leu Thr Cys Pro Gln Gly Arg Phe Gly Pro Asn Cys
              1100
                                 1105
Thr His Val Cys Gly Cys Gly Gln Gly Ala Ala Cys Asp Pro Val
              1115
                                 1120
Thr Gly Thr Cys Leu Cys Pro Pro Gly Arg Ala Gly Val Arg Cys
              1130
                               1135
Glu Arg Gly Cys Pro Gln Asn Arg Phe Gly Val Gly Cys Glu His
              1145
                                 1150
Thr Cys Ser Cys Arg Asn Gly Gly Leu Cys His Ala Ser Asn Gly
              1160
                                1165
Ser Cys Ser Cys Gly Leu Gly Trp Thr Gly Arg His Cys Glu Leu
              1175
                                 1180
Ala Cys Pro Pro Gly Arg Tyr Gly Ala Ala Cys His Leu Glu Cys
              1190
                                 1195
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                                1210
Arg Cys Gly Pro Gly Phe Tyr Gly Gln Ala Cys Glu His Pro Cys
              1220
                                1225
Pro Pro Gly Phe His Gly Ala Gly Cys Gln Gly Leu Cys Trp Cys
              1235
                                 1240
Gln His Gly Ala Pro Cys Asp Pro Ile Ser Gly Arg Cys Leu Cys
              1250
                                1255
Pro Ala Gly Phe His Gly His Phe Cys Glu Arg Gly Cys Glu Pro
              1265
                                 1270
Gly Ser Phe Gly Glu Gly Cys His Gln Arg Cys Asp Cys Asp Gly
              1280
                                 1285
Gly Ala Pro Cys Asp Pro Val Thr Gly Leu Cys Leu Cys Pro Pro
              1295
                                1300
Gly Arg Ser Gly Ala Thr Cys Asn Leu Gly Gly Pro Leu Arg Leu
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<213> Homo sapiens

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                                    25
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                                    40
Pro Glu Pro Tyr Tyr Pro Glu Ser Gly Trp Asp Arg Leu Arg Glu
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Asn Val Val Asp Lys Leu Glu Pro Val Leu His Glu Gly Leu Glu
                                    40
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Thr Val Asp Asn Thr Leu Lys Gly Ile Leu Glu Lys Leu Lys Val
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                                    55
Asp Leu Gly Val Leu Gln Lys Ser Ser Ala Trp Gln Leu Ala Lys
                                    70
                 65
Gln Lys Ala Gln Glu Ala Glu Lys Leu Leu Asn Asn Val Ile Ser
                80
                                    85
Lys Leu Leu Pro Thr Asn Thr Asp Ile Phe Gly Leu Lys Ile Ser
                95
                                   100
Asn Ser Leu Ile Leu Asp Val Lys Ala Glu Pro Ile Asp Asp Gly
                110
                                   115
Lys Gly Leu Asn Leu Ser Phe Pro Val Thr Ala Asn Val Thr Val
                                   130
Ala Gly Pro Ile Ile Gly Gln Ile Ile Asn Leu Lys Ala Ser Leu
                140
                                   145
Asp Leu Leu Thr Ala Val Thr Ile Glu Thr Asp Pro Gln Thr His
                155
                                 160
Gln Pro Val Ala Val Leu Gly Glu Cys Ala Ser Asp Pro Thr Ser
Ile Ser Leu Ser Leu Leu Asp Asn Gln Lys Cys Ile Glu Ala Gly
                                   190
His Asp Gly Ser Thr Pro Val Ile Pro Ala Leu Trp Glu Ala Glu
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240

235

Val His Ser Gly Gly Val Asn Cys Ile Ser Phe His Pro Ser Gly

230

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Asp Leu Leu Glu Gly Arg Leu Ile Tyr Thr Leu Gln Gly His Thr
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                                   265
Gly Pro Val Phe Thr Val Ser Phe Ser Lys Gly Glu Leu Phe
                                   280
               275
Ala Ser Gly Gly Ala Asp Thr Gln Val Leu Leu Trp Arg Thr Asn
               290
                                   295
Phe Asp Glu Leu His Cys Lys Gly Leu Thr Lys Arg Asn Leu Lys
               305
                                   310
Arg Leu His Phe Asp Ser Pro Pro His Leu Leu Asp Ile Tyr Pro
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                                   325
Arg Thr Pro His Pro His Glu Glu Lys Val Glu Thr Val Glu Thr
Thr Glu Thr Ser Gly Arg Thr Leu Pro Asp Lys Gly Glu Glu Ala
               350
                                   355
Cys Gly Tyr Phe Leu Asn Pro Ser Leu Met Ser Pro Glu Cys Leu
               365
                                   370
Pro Thr Thr Lys Lys Lys Thr Glu Asp Met Ser Asp Leu Pro
               380
                                   385
Cys Glu Ser Gln Arg Ser Ile Pro Leu Ala Val Thr Asp Ala Leu
               395
                                  400
Glu His Ile Met Glu Gln Leu Asn Val Leu Thr Gln Thr Val Ser
               410
                                   415
Ile Leu Glu Gln Arg Leu Thr Leu Thr Glu Asp Lys Leu Lys Asp
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Ser
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 Glu
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 Pro
 Ser
 Gly
 Arg
 Val
 Glu
 Gly
 Pro

 Pro
 Ala
 Trp
 Fro
 Trp
 Pro
 Ser
 Leu
 Pro
 Cys
 Gly
 Pro

 Cys
 Ile
 Pro
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 Met
 Leu
 Val
 Leu
 Ala
 Thr
 Leu
 Ala
 Ala
 Thr
 Leu
 Ala
 Ala
 Thr
 Leu
 Ala
 Ala

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Asn Leu Gly Pro Gln Thr Val Leu Glu Val Pro Ala Arg Ser Thr
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                                    145
                                                       150
Phe Trp Gly Pro Gln Pro Trp Glu Gly Arg Pro Pro Ala Thr Gly
                                                       165
                155
                                   160
Leu Val Ser Trp Ala Glu Pro Glu Gln Arg Pro Glu Ala Ser Val
                                   175
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Gln Phe Gly Ser Pro Gln Ala Arg Arg Gln Arg Pro Gly Ser Pro
                                   190
                185
Asp Pro Glu Trp Gly Leu Gln Pro Arg Val Thr Leu Glu Gln Ile
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cctccccagg agtagggagg aaccaggtgg gctggctggg atgggtggat atttaaagac 240
caggeettgg acgetgcage acttetatet etgettgatg eetgetgeca egtggetggt 300
cctcctcctc ctgctgtggc tgagccttgg ggtgaagaca ggcagctgct cccaacccca 360
gaacetttge tgtettggga eggateacea etgeaagagg ggaagttget aetgtgatga 420
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ctcacataca ggcccgattc cacctacagc aaagctggat gcgatggctg gcagaggcaa 540
accetttgcc tgcacttcag gccaaagccg ggatgtggcc tagatggttc ctaaggtccc 600
tgacaatect gagatettge atettgteta tttcaggtea aagettetea gatgaccaag 660
atggtgctgc agatggtgct gaggatggag aacccaccaa gccccgctag gagccaccta 720
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attaaaagca aggggaggag tggccaggtg caatggctca ctcccataaa cccagaactt 1380
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<213> Homo sapiens

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